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INSECT RESISTANT PLANTS

This is a continuation-in-part of U.S. Patent application ser. no. 535,354, filed September 24, 1983.

FIELD

The present invention is in the fields of genetic engineering, plant husbandry, and bacterial bio-affecting compositions, especially those derived from the genus Bacillus.

BACKGROUND

Insecticidal Protein

Bacillus thuringiensis, a species of bacteria closely related to B. cereus, forms a proteinacious crystalline inclusion during sporulation. This crystal is parasporal, forming within the cell at the end opposite from the developing spore. The crystal protein, often referred to as the δ -endotoxin, has two forms: a nontoxic protoxin of approximate molecular weight (MW) of 130 kilodaltons (kD), and a toxin having an approx. MW of 67 kD. The crystal contains the protoxin protein which is activated in the gut of larvae of a number of insect species. Klowden MJ et al. (1983) Appl. Envir. Microbiol. 46:312-315, have shown solubilized protoxin from B. thuringiensis var. israelensis is toxic to Aedes aegypti adults. During activation, the protoxin is cleaved into two polypeptides, one or both of which are toxic. In vivo, the crystal is activated by being solubilized and converted to toxic form by the alkalinity and proteases of the gut. In vitro the protoxin may be solubilized by extremely high pH (e.g. pH 12), by reducing agents under moderately basic conditions (e.g. pH 10), or by strong denaturants (guanidium, urea) under neutral conditions (pH 7), and once solubilized, may be activated by the action of the protease trypsin. The crystal protein is reported to be antigenically related to proteins within both the spore coat and the vegetative cell wall. Carbohydrate is not involved in the toxic properties of the protein.

B. thuringiensis and its crystalline endotoxin are useful because the crystal protein is an insecticidal protein known to be poisonous to the larvae of over a hundred of species of insects, most commonly those from the orders Lepidoptera and Diptera. Insects susceptible to the action of the B. thuringiensis crystal protein include, but need not be limited to, those listed in Table 1. Many of these insect species are economically important pests. Plants which can be protected by application of the crystal protein include, but need not be limited to, those listed in Table 2. Different varieties of B. thuringiensis, which include, but need not be limited to, those listed in Table 3, have different host ranges (Faust RM et al. (1982) in Genetic Engineering in the Plant Sciences, ed. Panapoulos NJ, pp. 225-254); this probably reflects the toxicity of a given crystal protein in a particular host. The crystal protein is highly specific to insects; in over two decades of commercial application of sporulated B. thuringiensis cells to crops and ornamentals there has been no known case of effects to plants or noninsect animals. The efficacy and safety of the endotoxin have been reviewed by Faust RM et al., supra. Other useful reviews include those by Fast PG (1981) in Microbial Control of Pests and Plant Diseases, 1970-1980, ed.: Burges HD, pp. 223-248, and Huber HE and Luthy P (1981) in Pathogenesis of Invertebrate Microbial Diseases, ed.: Davidson EW, pp. 209-234.

The crystal protein gene usually can be found on one of several large plasmids that have been found in Bacillus thuringiensis, though in some strains it may be located on the chromosome (Kronstad JW et al. (1983) J. Bacteriol. 154:419-428). Several of the genes have been cloned into plasmids that can grow in E. coli. Whiteley's group (Whiteley HR et al. (1982) in Molecular Cloning and Gene Regulation in Bacilli, ed.: Ganesan AT et al., pp. 131-144, Schnepf HE and Whiteley HR (1981) Proc. Natl. Acad. Sci. USA 78:2893-2897, and European pat. application 63,949) reported the cloning of the toxin from B. thuringiensis var. kurstaki strains HD-1-Dipel and HD-73, using the enzymes Sau3AI (under partial digest conditions) and BglII, respectively, to insert large gene-bearing fragments having approximate sizes of 12 kbp and 16 kbp into the BamHI site of the E. coli plasmid vector pBR322. The HD-1 crystal protein was observed to be located on a 6.6 kilobase pair (kbp) HindIII fragment.

Crystal protein from the HD-1-Dipel gene which was toxic to larvae, immunologically identifiable, and the same size as authentic protoxin, was observed to be produced by transformed E. coli cells containing pBR322 clones or subclones. This indicated that the Bacillus gene was transcribed, probably from its own promoter, and translated in E. coli. Additionally, this suggests that the toxic activity of the protein product is independent of the location of its synthesis. That the gene was expressed when the fragment containing it was inserted into the vector in either orientation suggests that transcription was controlled by its own promoter. The transcriptional and translational start sites, as well as the deduced sequence for the amino-terminal 333 amino acids of the HD-1-Dipel protoxin, have been determined by nucleic acid sequencing (Wong HC et al. (1983) J. Biol. Chem. 258:1960-1967). The insecticidal gene was found to have the expected bacterial ribosome binding and translational start (ATG) sites along with commonly found sequences at -10 and -35 (relative to the 5'-end of the mRNA) that are involved in initiation of transcription in bacteria such as B. subtilis. Klier A et al. (1982) EMBO J. 1:791-799, have reported the cloning of the crystal protein gene from B. thuringiensis strain berliner 1715 in pBR322. Using the enzyme BamHI, a large 14 kbp fragment carrying the crystal protein gene was moved into the vector pHV33, which can replicate in both E. coli and Bacillus. In both E. coli and sporulating B. subtilis, the pHV33-based clone directed the synthesis of full-size (130 kD) protoxin which formed cytoplasmic inclusion bodies and reacted with antibodies prepared against authentic protoxin. Extracts of E. coli cells harboring the pBR322 or pHV33-based plasmids were toxic to larvae. In further work, Klier A et al. (1983) Nucleic Acids Res. 11:3973-3987, have transcribed the berliner crystal protein gene in vitro and have reported on the sequence of the promoter region, together with the first 11 codons of the crystal protein. The bacterial ribosome binding and translational start sites were identified. Though the expected "-10" sequence was identified, no homology to other promoters has yet been seen near -35. Held et al. (1982) Proc. Natl. Acad. Sci. USA 77:6065-6069 reported the cloning of a crystal protein gene from the variety kurstaki in the phage λ -based cloning vector Charon4A. E. coli cells infected with one of the Charon clones produced antigen that was the same size as the protoxin (130 kD) and was toxic to

larvae. A 4.6 kbp EcoRI fragment of this Charon clone was moved into pHV33 and an E. coli plasmid vector, pBR328. Again, 130 kD antigenically identifiable crystal protein was produced by both E. coli and B. subtilis strains harboring the appropriate plasmids. A B. thuringiensis chromosomal sequence which cross-hybridized with the cloned crystal protein gene was identified in B. thuringiensis strains which do not produce crystal protein during sporulation.

In addition to the crystal protein, B. thuringiensis produces at least three other toxins. Two of them, the α -exotoxin and γ -exotoxin, are phospholipase enzymes that degrade lipids. B. cereus is also known to produce phospholipases (or lecithinases) which are toxic to insect larvae. Other bacterial enzymes which are involved in insect pathogenesis include, but need not be limited to, hyaluronidases, phosphatases, and proteases. Protease produced by Pseudomonas aeruginosa has been shown to have a specific affinity to proteins of Galleria mellonella larvae (see Lysenko O and Kucera M (1971) in Microbial Control of Insects and Mites, eds.: Burges HD and Hussey HW, pp. 205-227).

Chang S (1983) Trends Biotechnol. 1:100-101; reported that the DNA sequence of a complete HD-1 gene had been publicly presented (ref. 5 therein), and that the HD-1 toxin moiety resides in the amino-terminal 67 kD peptide.

Shuttle Vectors

Shuttle vectors, developed by Ruvkun GB and Ausubel FM (1981) Nature 298:85-88, provide a way to insert foreign genetic materials into position of choice in a large plasmid, virus, or genome. There are two main problems encountered when dealing with large plasmids or genomes. Firstly, the large plasmids may have many sites for each restriction enzyme. Unique site-specific cleavage reactions are not reproducible and multi-site cleavage reactions followed by ligation lead to great difficulties due to the scrambling of the many fragments whose order and orientation one does not want changed. Secondly, the transformation efficiency with large DNA plasmids is very low. Shuttle vectors allow one to overcome these difficulties by facilitating the insertion, often in vitro, of

the foreign genetic material into a smaller plasmid, then transferring, usually by in vivo techniques, to the larger plasmid.

A shuttle vector consists of a DNA molecule, usually a plasmid, capable of being introduced into the ultimate recipient bacteria. It also includes a copy of the fragment of the recipient genome into which the foreign genetic material is to be inserted and a DNA segment coding for a selectable trait, which is also inserted into the recipient genome fragment. The selectable trait ("marker") is conveniently inserted by transposon mutagenesis or by restriction enzymes and ligases.

The shuttle vector can be introduced into the ultimate recipient cell, typically a bacterium of the family Rhizobiaceae (which contains the genus Agrobacterium), by a tri-parental mating (Ruvkin and Ausubel, supra), direct transfer of a self-mobilizable vector in a bi-parental mating, direct uptake of exogenous DNA by Agrobacterium cells ("transformation", using the conditions of Holsters M et al. (1978) Molec. Gen. Genet. 163:181-187), by spheroplast fusion of Agrobacterium with another bacterial cell, by uptake of liposome-encapsulated DNA, or infection with a shuttle vector that is based on a virus that is capable of being packaged in vitro. A tri-parental mating involves the mating of a strain containing a mobilizable plasmid, which carries genes for plasmid mobilization and conjugative transfer, with the strain containing the shuttle vector. If the shuttle vector is capable of being mobilized by the plasmid genes, the shuttle vector is transferred to the recipient cell containing the large genome, e.g. the Ti or Ri plasmids of Agrobacterium strains.

After the shuttle vector is introduced into the recipient cell, possible events include a double cross over with one recombinational event on either side of the marker. This event will result in transfer of a DNA segment containing the marker to the recipient genome replacing a homologous segment lacking the insert. To select for cells that have lost the original shuttle vector, the shuttle vector must be incapable of replicating in the ultimate host cell or be incompatible with an independently selectable plasmid pre-existing in the recipient cell. One common means of arranging this is to provide in the third parent another plasmid which

is incompatible with the shuttle vector and which carries a different drug resistance marker. Therefore, when one selects for resistance to both drugs, the only surviving cells are those in which the marker on the shuttle vector has recombined with the recipient genome. If the shuttle vector carries an extra marker, one can then screen for and discard cells that contain plasmids resulting from a single cross-over event between the shuttle vector and the recipient plasmid resulting in cointegrates in which the entire shuttle vector is integrated with the recipient plasmid. If the foreign genetic material is inserted into or adjacent to the marker that is selected for, it will also be integrated into the recipient plasmid as a result of the same double recombination. It might also be carried along when inserted into the homologous fragment at a spot not within or adjacent to the marker, but the greater the distance separating the foreign genetic material from the marker, the more likely will be a recombinational event occurring between the foreign genetic material and marker, preventing transfer of the foreign genetic material.

If the shuttle vector is used to introduce a phenotypically dominant trait (e.g. a novel expressible insecticide structural gene, but not an inactivated oncogenic T-DNA gene) one need not rely on a double homologous recombination. The cells resulting from a single cross-over event resulting in cointegrate plasmids can transfer the desired trait into plant cells. One may even use a variant shuttle vector having a single interrupted sequence of T-DNA. However, as the resulting T-DNA will now contain a tandem duplication, one must be vigilant regarding a possible rare deletion of the shuttle vector by a single homologous recombination event occurring between the two homologous sequences in either the Agrobacterium or plant cells.

Shuttle vectors have proved useful in manipulation of Agrobacterium plasmids: see Garfinkel DJ et al. (1981) Cell 27:143-153, Matzke AJM and Chilton M-D (1981) J. Molec. Appl. Genet. 1:39-49, and Leemans J et al. (1981) J. Molec. Appl. Genet. 1:149-164, who referred to shuttle vectors by the term "intermediate vectors".

A recently disclosed variation of the shuttle vector system for inserting changes into large DNA molecules is the "suicide vector". In

this system, as described by Puhler A et al.; US application ser. no. 510,370 and Simon R et al. (1983) in press, the shuttle vector is incapable of being maintained within the recipient cell. This property eliminates the need to introduce an incompatible plasmid into the recipient cell in order to exclude the shuttle vector as is commonly done during a triparental mating. All vectors which do not integrate into some already present DNA effectively "commit suicide" by not being replicated. As can be done with traditional types of shuttle vectors, one may distinguish between double and single homologous by screening for an antibiotic resistance gene which is not between the two regions of homology. Use of a pBR322-based suicide vector to transfer DNA sequences into a Ti plasmid has been reported by Van Haute E et al. (1983) EMBO J. 2:411-417; and Comai L et al. (1982) Plant. Molec. Biol. 1:291-300.

An alternative to the use of shuttle vectors for introduction of novel DNA sequences into T-DNA by means of homologous recombination involves bacterial transposons. As described in the section Agrobacterium-Genes on the TIP Plasmids, transposons can "jump" into the T-DNA of a TIP plasmid (e.g. see Gartinkel DJ et al. (1981) Cell 27:143-153). Should the transposon be modified in vitro by the insertion of the novel sequence, that novel DNA can be transferred into the TIP plasmid's T-DNA by the transposon. The TIP can then transfer the novel DNA/transposon/T-DNA combination to a plant cell where it will be stably integrated.

Agrobacterium-Overview

Included within the gram-negative bacterial family Rhizobiaceae in the genus Agrobacterium are the species A. tumefaciens and A. rhizogenes. These species are respectively the causal agents of crown gall disease and hairy root disease of plants. Crown gall is characterized by the growth of a gall of dedifferentiated tissue. Hairy root is a teratoma characterized by inappropriate induction of roots in infected tissue. In both diseases, the inappropriately growing plant tissue usually produces one or more amino acid derivatives, known as opines, not normally produced by the plant which are catabolized by the infecting bacteria. Known opines have been classified into three main families whose type members are octopine, nopaline, and agropine. The cells of inappro-

priately growing tissues can be grown in culture, and, under appropriate conditions, be regenerated into whole plants that retain certain transformed phenotypes.

Virulent strains of Agrobacterium harbor large plasmids known as Ti (tumor-inducing) plasmids in A. tumefaciens and Ri (root-inducing) plasmids in A. rhizogenes. Curing a strain of these plasmids results in a loss of pathogenicity. The Ti plasmid contains a region, referred to as T-DNA (transferred-DNA), which in tumors is found to be integrated into the genome of the host plant. The T-DNA encodes several transcripts. Mutational studies have shown that some of these are involved in induction of tumorous growth. Mutants in the genes for tml, tmr, and tms, respectively result in large tumors (in tobacco), a propensity to generate roots, and a tendency for shoot induction. The T-DNA also encodes the gene for at least one opine synthase, and the Ti plasmids are often classified by the opine which they caused to be synthesized. Each of the T-DNA genes is under control of a T-DNA promoter. The T-DNA promoters resemble eukaryotic promoters in structure, and they appear to function only in the transformed plant cell. The Ti plasmid also carries genes outside the T-DNA region. These genes are involved in functions which include opine catabolism, oncogenicity, agrocin sensitivity, replication, and autotransfer to bacterial cells. The Ri plasmid is organized in a fashion analogous to the Ti plasmid. The set of genes and DNA sequences responsible for transforming the plant cell are hereinafter collectively referred to as the transformation-inducing principle (TIP). The designation TIP therefore includes both Ti and Ri plasmids. The integrated segment of a TIP is termed herein "T-DNA" (transferred DNA), whether derived from a Ti plasmid or an Ri plasmid.

Chilton M-D (June 1983) Sci. Amer. 248(6):50-59, has recently provided an introductory article on the use of Ti plasmids as vectors. Recent general reviews of Agrobacterium-caused disease include those by Merlo DJ (1982), Adv. Plant Pathol. 1:139-178, Ream LW and Gordon MP (1982), Science 218:854-859, and Bevan MW and Chilton M-D (1982), Ann. Rev. Genet. 16:357-384; Kahl G and Schell J (1982) Molecular Biology of Plant Tumors, and Barton KA and Chilton M-D (1983) Meth. Enzymol. 101:527-539.

Agrobacterium-Infection of Plant Tissues

Plant cells can be transformed by Agrobacterium in a number of methods known in the art which include but are not limited to co-cultivation of plant cells in culture with Agrobacterium, direct infection of a plant, fusion of plant protoplasts with Agrobacterium spheroplasts, direct transformation by uptake of free DNA by plant cell protoplasts, transformation of protoplasts having partly regenerated cell walls with intact bacteria transformation of protoplasts by liposomes containing T-DNA, use of a virus to carry in the T-DNA, microinjection, and the like. Any method will suffice as long as the gene is reliably expressed, and is stably transmitted through mitosis and meiosis.

The infection of plant tissue by Agrobacterium is a simple technique well known to those skilled in the art (for an example, see Butcher DN et al. (1980) in Tissue Culture Methods for Plant Pathologists, eds.: Ingram DS and Helgeson JP, pp. 203-208). Typically a plant is wounded by any of a number of ways, which include cutting with a razor, puncturing with a needle, or rubbing with abrasive. The wound is then inoculated with a solution containing tumor-inducing bacteria. An alternative to the infection of intact plants is the inoculation of pieces of tissues such as potato tuber disks (Anand DK and Heberlein GT (1977) *Amer. J. Bot.* 64:153-158) or segments of tobacco stems (Barton KA et al. (1983) *Cell* 32:1033-1043). After induction, the tumors can be placed in tissue culture on media lacking phytohormones. Hormone independent growth is typical of transformed plant tissue and is in great contrast to the usual conditions of growth of such tissue in culture (Braun AC (1956) *Cancer Res.* 16:53-56).

Agrobacterium is also capable of infecting isolated cells and cells grown in culture (Marton L et al. (1979) *Nature* 277:129-131) and isolated tobacco mesophyll protoplasts. In the latter technique, after allowing time for partial regeneration of new cell walls, Agrobacterium cells were added to the culture for a time and then killed by the addition of antibiotics. Only those cells exposed to A. tumefaciens cells harboring the Ti plasmid were capable of forming calli when plated on media lacking

hormone. Most calli were found to contain an enzymatic activity involved in opine anabolism. Other workers (Horsch RB and Fraley RT (18 January 1983) 15th Miami Winter Symposium) have reported transformations by co-cultivation, leading to a high rate (greater than 10%) of calli displaying hormone-independent growth, with 95% of those calli making opines.

Davey MR et al. (1980) in Ingram and Helgeson, supra, pp. 209-219, describe the infection of older cells that had been regenerated from protoplasts.

Plant protoplasts can be transformed by the direct uptake of Ti plasmids. Davey MR et al. (1980) Plant Sci. Lett. 18:307-313, and Davey MR et al. (1980) in Ingram and Helgeson, supra, were able to transform Petunia protoplasts with the Ti plasmid in the presence of poly-L- α -ornithine to a phenotype of opine synthesis and hormone-independent growth in culture. It was later shown (Draper J et al. (1982) Plant and Cell Physiol. 23:451-458, Davey MR et al. (1982) in Plant Tissue Culture 1982, ed: Fujiwara A, pp. 515-516) that polyethylene glycol-stimulated Ti plasmid uptake and that some T-DNA sequences were integrated into the genome. Krens FA et al. (1982) Nature 296:72-74, reported similar results using polyethylene glycol following by a calcium shock, though their data suggests that the integrated T-DNA included flanking Ti plasmid sequences.

An alternative method to obtain DNA uptake involves the use of liposomes. The preparation of DNA containing liposomes is taught by Papanadjopoulos in U.S. Patents 4,078,052 and 4,235,871. Preparations for the introduction of Ti-DNA via liposomes have been reported (Nagata T et al. (1982) in Fujiwara, supra, pp. 509-510, and Nagata T (1981) Mol. Gen. Genet. 184:161-165). An analogous system involves the fusion of plant and bacterial cells after removal of their cell walls. An example of this technique is the transformation of Vinca protoplast by Agrobacterium spheroplasts reported by Hasezawa S et al. (1981) Mol. Gen. Genet. 182:206-210. Plant protoplasts can take up cell wall delimited Agrobacterium cells (Hasezawa S et al. (1982) in Fujiwara, supra pp. 517-518).

T-DNA can be transmitted to tissue regenerated from a fusion of two protoplasts, only one of which had been transformed (Wullems GJ et al. (1980) Theor. Appl. Genet. 56:203-208). As detailed in the section on Regeneration of Plants, T-DNA can pass through meiosis and be transmitted to progeny as a simple Mendelian trait.

Agrobacterium--Regeneration of Plants

Differentiated plant tissues with normal morphology have been obtained from crown gall tumors. Braun AC and Wood HN (1976) Proc. Natl. Acad. Sci. USA 73:496-500, grafted tobacco teratomas onto normal plants and were able to obtain normally appearing shoots which could flower. The shoots retained the ability to make opines and to grow independently of phytohormones when placed in culture. In the plants screened, these tumorous phenotypes were not observed to be transmitted to progeny, apparently being lost during meiosis (Turgeon R et al. (1976) Proc. Natl. Acad. Sci. USA 73:3562-3564). Plants which had spontaneously lost tumorous properties, or which were derived from teratoma seed, were initially shown to have lost all their T-DNA (Yang F-M et al. (1980) In Vitro 16:87-92, Yang F et al. (1980) Molec. Gen. Genet. 177:707-714, Lemmers M et al. (1980) J. Mol. Biol. 144:353-376). However, later work with plants that had become revertants after hormone treatment (1 mg/l kinetin) showed that plants which had gone through meiosis, though losing T-DNA genes responsible for the transformed phenotype, could retain sequences homologous to both ends of T-DNA (Yang F and Simpson RB (1981) Proc. Natl. Acad. Sci. USA 78:4151-4155). Wullems GJ et al. (1981) Cell 24:719-724, further demonstrated that genes involved in opine anabolism were capable of passing through meiosis though the plants were male sterile and that seemingly unaltered T-DNA could be inherited in a Mendelian fashion (Wullems G et al. (1982) in Fujiwara, supra). Otten L et al. (1981) Molec. Gen. Genet. 183:209-213, used Tn7 transposon-generated Ti plasmid mutants in the tms (shoot-inducing) locus to create tumors which proliferated shoots. When these shoots were regenerated into plants, they were found to form self-fertile flowers. The resultant seeds germinated into plants which contained T-DNA and made opines. In further experiments, DeGreve H et al. (1982) Nature 300:752-755, have found that octopine synthase can be

inherited as a single dominant Mendelian gene. However, the T-DNA had sustained extensive deletions of functions other than ocs while undergoing regeneration from callus. Similar experiments with a tmr (root-inducing) mutant showed that full-length T-DNA could be transmitted through meiosis to progeny, that in those progeny nopaline genes could be expressed, though at variable levels, and that cotransformed yeast alcohol dehydrogenase I gene was not expressed (Barton KA et al. (1983) Cell 32:1033-1043). It now appears that regenerated tissues which lack T-DNA sequences are probably descended from untransformed cells which "contaminate" the tumor (Ooms G et al. (1982) Cell 30:589-597). Recent work by Binns AN (1983) Planta 158:272-279, indicates that tumorigenic genes, in this case tmr, can be "shut off" during regeneration and "turned back on" by placing regenerated tissue in culture.

Roots resulting from transformation from A. rhizogenes have proven relatively easy to regenerate directly into plantlets (Chilton M-D et al. (1982) Nature 295:432-434.

Agrobacterium-Genes on the TIP Plasmids

A number of genes have been identified within the T-DNA of the TIP plasmids. About half a dozen octopine plasmid T-DNA transcripts have been mapped (Gelvin SB et al. (1982) Proc. Natl. Acad. Sci. USA 79:76-80, Willmitzer L et al. (1982) EMBO J. 1:139-146) and some functions have been assigned (Leemans J et al. (1982) EMBO J. 1:147-152). Some of these transcripts, specifically those in the region encoding tmr and tms, can also be transcribed in prokaryotic cells (Schroder G et al. (1983) EMBO J. 2:403-409). The four genes of an octopine type plasmid that have been well defined by transposon mutagenesis include tms, tmr, and tml (Garfinkel DJ et al. (1981) Cell 27:143-153). Ti plasmids which carry mutations in these genes respectively incite tumorous calli of Nicotiana tabacum which generate shoots, proliferate roots, and are larger than normal. In other hosts, mutants of these genes can induce different phenotypes (see Bevan MW and Chilton M-D (1982) Ann. Rev. Genet. 16:357-384). The phenotypes of tms and tmr are correlated with differences in the phytohormone levels present in the tumor. The differences in cytokinin:auxin ratios are similar to those which in culture induce shoot or

root formation in untransformed callus tissue (Akiyoshi DE *et al.* (1983) Proc. Natl. Acad. Sci. USA 80:407-411). T-DNA containing a functional gene for either tms or tmr alone, but not functional tml alone, can promote significant tumor growth. Promotion of shoots and roots is respectively stimulated and inhibited by functional tml (Ream LW *et al.* (1983) Proc. Natl. Acad. Sci. USA 80:1660-1664). Mutations in T-DNA genes do not seem to affect the insertion of T-DNA into the plant genome (Leemans *et al.* (1982) *supra*, Ream *et al.* (1983) *supra*). The ocs gene encodes octopine synthase, which has been sequenced by De Greve H *et al.* (1982) J. Mol. Appl. Genet. 1:499-511. It does not contain introns (intervening sequences commonly found in eukaryotic genes which are post-transcriptionally spliced out of the messenger precursor during maturation of the mRNA). It does have sequences that resemble a eukaryotic transcriptional signal ("TATA box") and a polyadenylation site. All of the signals necessary for expression of the ocs gene are found within 295 bp of the ocs transcriptional start site (Koncz C *et al.* (1983) EMBO J. 2:1597-1603).

Nopaline Ti plasmids encode the nopaline synthase gene (nos), which has been sequenced by Depicker A *et al.* (1982) J. Mol. Appl. Genet. 1:561-573. As was found with the ocs gene, nos is not interrupted by introns. It has two putative polyadenylation sites and a potential "TATA box". In contrast to ocs, nos is preceded by a sequence which may be a transcriptional signal known as a "CAT box". All of the signals necessary for expression of the nos gene are found within 261 bp of the nos transcriptional start site (Koncz C *et al.*, *supra*). A gene for agrocinopine synthase and genes equivalent to tms and tmr have been identified on a nopaline-type plasmid (Joos H *et al.* (1983) Cell 32:1057-1067), and a number of transcripts have been mapped (Willmitzer L *et al.* (1983) Cell 32:1045-1056). McPherson JC *et al.* (1980) Proc. Natl. Acad. Sci. USA 77:2666-2670, reported the *in vitro* translation of T-DNA encoded mRNAs from crown gall tissues.

Transcription from hairy root T-DNA has also been detected (Willmitzer L *et al.* (1982) Mol. Gen. Genet. 186:16-22). Functionally, the hairy root syndrome appears to be equivalent of a crown gall tumor incited by a Ti plasmid mutated in tmr (White FF and Nester EW (1980) J. Bacteriol. 144:710-720).

In eukaryotes, methylation (especially of cytosine residues) of DNA is correlated with transcriptional inactivation; genes that are relatively under methylated are transcribed into mRNA. Gelvin SB et al. (1983) *Nucleic Acids Res.* 11:159-174, has found that the T-DNA in crown gall tumors is always present in at least one unmethylated copy. That the same genome may contain numerous other copies of T-DNA which are methylated suggests that the copies of T-DNA in excess of one may be biologically inert. (See also Ooms G et al. (1982) *Cell* 30:589-597.)

The Ti plasmid encodes other genes which are outside of the T-DNA region and are necessary for the infection process. (See Holsters M et al. (1980) *Plasmid* 3:212-230 for nopaline plasmids, and De Greve H et al. (1981) *Plasmid* 6:235-248, Garfinkel DJ and Nester EW (1980) *J. Bacteriol* 144:732-743, and Ooms G (1980) *J. Bacteriol* 144:82-91 for octopine plasmids). Most important are the onc genes, which when mutated result in Ti plasmids incapable of oncogenicity. (These loci are also known as vir, for virulence.) Several onc genes have been accurately mapped and have been found to be located in regions conserved among various Ti plasmids (Klee HJ et al. (1983) *J. Bacteriol.* 153:878-883, Iyer VN et al. (1982) *Mol. Gen. Genet.* 188:418-424). The onc genes function in trans, being capable of causing the transformation of plant cells with T-DNA of a different plasmid type and physically located on another plasmid (Hille J et al. (1982) *Plasmid* 7:107-118, Klee HJ et al. (1982) *J. Bacteriol* 150:327-331, de Framond AJ et al. (1983) *Biotechnol.* 1:262-269). Nopaline Ti DNA has direct repeats of about 25 base pairs immediately adjacent to the left and right borders of the T-DNA which might be involved in either excision from the Ti plasmid or integration into the host genome (Yadav NS et al. (1982) *Proc. Natl. Acad. Sci. USA* 79:6322-6326), and a homologous sequence has been observed adjacent to an octopine T-DNA border (Simpson RB et al. (1982) *Cell* 29:1005-1014). Opine catabolism is specified by the occ and noc genes, respectively, of octopine- and nopaline-type plasmids. The Ti plasmid also encodes functions necessary for its own reproduction including an origin of replication. Ti plasmid transcripts have been detected in *A. tumefaciens* cells by Gelvin SB et al. (1981) *Plasmid* 6:17-29, who found that T-DNA regions were weakly transcribed along with non-T-DNA sequences. Ti plasmid-determined

characteristics have been reviewed by Merlo, supra (see especially Table II), and Ream and Gordon supra.

Agrobacterium-Ti Plasmid DNA

Different octopine-type Ti plasmids are nearly 100% homologous to each other when examined by DNA hybridization (Currier TC and Nester EW (1976) J. Bacteriol. 126:157-165) or restriction enzyme analysis (Sciaky D et al. (1978) Plasmid 1:238-253). Nopaline-type Ti plasmids have as little as 67% homology to each other (Currier and Nester, supra). A survey revealed that different Ri plasmids are very homologous to each other (Costantino P et al. (1981) Plasmid 5:170-182). Drummond NH and Chilton M-D (1978) J. Bacteriol. 136:1178-1183, showed that proportionally small sections of octopine- and nopaline-type Ti plasmids were homologous to each other. These homologies were mapped in detail by Engler G et al. (1981) J. Mol. Biol. 152:183-208. They found that three of the four homologous regions were subdivided into three (overlapping the T-DNA), four (containing some onc genes), and nine (having onc genes) homologous sequences. The uninterrupted homology contains at least one tra gene (for conjugal transfer of the Ti plasmid to other bacterial cells), and genes involved in replication and incompatibility. This uninterrupted region has homology with a Sym plasmid (involved in symbiotic nitrogen fixation) from a species of Rhizobium, a different genus in the family Rhizobiaceae (Prakash RK et al. (1982) Plasmid 7:271-280). The order of the four regions is not conserved, though they are all oriented in the same direction. Part of the T-DNA sequence is very highly conserved between nopaline and octopine plasmids (Chilton M-D et al. (1978) Nature 275:147-149, Depicker A et al. (1978) Nature 275:150-153). Ri plasmids have been shown to have extensive homology among themselves, and to both octopine (White FF and Nester EW (1980) J. Bacteriol. 144:710-720) and nopaline (Risuleo G et al. (1982) Plasmid 7:45-51) Ti plasmids, primarily in regions encoding onc genes. Ri T-DNA contains extensive though weak homologies to T-DNA from both types of Ti plasmid (Willmitzer L et al. (1982) Mol. Gen. Genet. 186:16-22). Plant DNA from uninfected Nicotiana glauca contains sequences, referred to as cT-DNA (cellular T-DNA), that show homology to a portion of the Ri T-DNA (White FF et al. (1983) Nature

301:348-350, Spanó L et al. (1982) Plant Molec. Biol. 1:291-300).
Huffman GA et al. (1983) J. Bacteriol., have mapped the region of cross-hybridization and have shown that Ri plasmid, pRiA4b, is more closely related to a pTiA6 (octopine-type) than pTiT37 (nopaline-type) and that this Ri plasmid appears to carry sequence homologous to tms but not tmr. Their results also suggested that Ri T-DNA may be discontinuous, analogous to the case with octopine T-DNA.

It has been shown that a portion of the Ti (Chilton M-D et al. (1977) Cell 11:263-271) or Ri (Chilton M-D (1982) Nature 295:432-434, White FF et al. (1982) Proc. Natl. Acad. Sci. USA 79:3193-3197, Willmitzer L (1982) Mol. Gen. Genet. 186:16-22) plasmid is found in the DNA of tumorous plant cells. The transferred DNA is known as T-DNA. T-DNA is integrated into the host DNA (Thomashow MF et al. (1980) Proc. Natl. Acad. Sci. USA 77:6448-6452, Yadav NS et al. (1980) Nature 287:458-461) in the nucleus (Nutti MP et al. (1980) Plant Sci. Lett. 18:1-6, Willmitzer L et al. (1980) Nature 287:359-361, Chilton M-D et al. (1980) Proc. Natl. Acad. Sci. USA 77:4060-4064).

Thomashow MF et al. (1980) Proc. Natl. Acad. Sci. USA 77:6448-6452, and Thomashow MF et al. (1980) Cell 19:729-739, found the T-DNA from octopine-type Ti plasmids to have been integrated in two separate sections, TL-DNA and TR-DNA, left and right T-DNAs respectively. The copy numbers of TR and TL can vary (Merlo DJ et al. (1980) Molec. Gen. Genet. 177:637-643). A core of T-DNA is highly homologous to nopaline T-DNA (Chilton et al. (1978) supra, and Depicker et al. (1978) supra), is required for tumor maintenance, is found in TL, is generally present in one copy per cell, and codes for the genes tms, tmr, and tml. On the other hand TR can be totally dispensed with (De Beuckeleer M et al. (1981) Molec. Gen. Genet. 183:283-288, Ooms G et al. (1982) Cell 30:589-597), though found in a high copy number (Merlo et al. (1980) supra). Ooms G et al. (1982) Plasmid 7:15-29, hypothesized that TR is involved in T-DNA integration, though they find that when TR is deleted from the Ti plasmid, A. tumefaciens does retain some virulence. Ooms G et al. (1982) Cell 30:589-597, showed that though T-DNA is occasionally deleted after integration in the plant genome, it is generally stable and that tumors containing a mixture of cells that differ in T-DNA organization are the

result of multiple transformation events. The ocs is found in TL but can be deleted from the plant genome without loss of phenotypes related to tumorous growth. The left border of integrated TL has been observed to be composed of repeats of T-DNA sequences which are in either direct or inverted orientations (Simpson RB et al. (1982) Cell 29:1005-1014).

In contrast to the situation in octopine-type tumors, nopaline T-DNA is integrated into the host genome in one continuous fragment (Lemmers M et al. (1980) J. Mol. Biol. 144:353-376; Zambryski P et al. (1980) Science 209:1385-1391). Direct tandem repeats were observed. T-DNA of plants regenerated from teratomas had minor modifications in the border fragments of the inserted DNA (Lemmers et al., supra). Sequence analysis of the junction between the right and left borders revealed a number of direct repeats and one inverted repeat. The latter spanned the junction (Zambryski et al. (1980) supra). The left junction has been shown to vary by at least 70 base pairs while the right junction varies no more than a single nucleotide (Zambryski P et al. (1982) J. Molec. Appl. Genet. 1:361-370). Left and right borders in junctions of tandem arrays were separated by spacers which could be over 130 bp. The spacers were of unknown origin and contained some T-DNA sequences. T-DNA was found to be integrated into both repeated and low copy number host sequences. H. Joos et al. (1983) Cell 32:1057-1067, have shown that virulence is not eliminated after deletion of either of the usual nopaline T-DNA borders.

Simpson et al. (1982) supra, and Zambryski et al. (1980) supra have suggested that direct repeats in the border regions are involved in integration of T-DNA into plant DNA. That T-DNA having borders from two different Ti plasmids are less specifically integrated than are homologous borders supports this suggestion (Ooms G et al. (1982) Plant Molec. Biol. 1:265-276).

Yadav NS et al. (1982) Proc. Natl. Acad. Sci. USA 79:6322-6326, have found a chi site, which in the bacteriophage λ augments general recombination in the surrounding DNA as far as 10 kilobases away, in a nopaline Ti plasmid just outside the left end of the T-DNA. Simpson RB et al. (1982) Cell 29:1005-1014, have not observed a chi sequence in an octopine Ti plasmid, though the possible range of action does not eliminate the possi-

bility of one being necessary and present but outside of the region sequenced. The significance of the chi in the Ti plasmid is not known. If the chi has a function, it is probably used in Agrobacterium cells and not in the plants, as chi is not found within the T-DNA.

Agrobacterium-Manipulations of the TIP Plasmids

As detailed in the section on Shuttle Vectors, technology has been developed for the introduction of altered DNA sequences into desired locations on a TIP plasmid. Transposons can be easily inserted using this technology (Garfinkel DJ et al. (1981) Cell 27:143-153). Hernalsteen J-P et al. (1980) Nature 287:654-656, have shown that a DNA sequence (here a bacterial transposon) inserted into T-DNA in the Ti plasmid is transferred and integrated into the recipient plant's genome. Though insertion of foreign DNA has been done with a number of genes from different sources, to date foreign genes have not usually been expressed under control of their own promoters. Sources of these genes include alcohol dehydrogenase (Adh) from yeast (Barton KA et al. (1983) Cell 32:1033-1043), AdhI (Bennetzen J, unpublished) and zein from corn, interferon and globin from mammals, and the mammalian virus SV40 (Schell J, unpublished). However, when the nopaline synthase gene was inserted into octopine T-DNA and transformed into plant tissue, it was found to be fully functional (Fink CL (1982) M.S. thesis, University of Wisconsin-Madison). The gene encoding phaseolin, the storage protein found in seeds of the bean Phaseolus vulgaris L., has been transferred into and expressed in sunflower tumors. This latter work constitutes the first example of a transferred plant gene being expressed under control of its own promoter in foreign plant tissue. Transcription started and stopped at the correct positions, and introns were posttranscriptionally processed properly (Hall TC et al., U.S. application ser. no. 485,613, which is hereby incorporated by reference). Holsters M et al. (1982) Mol. Gen. Genet. 185:283-289, have shown that a bacterial transposon (Tn7) inserted into T-DNA could be recovered in a fully functional and seemingly unchanged form after integration into a plant genome.

Deletions can be generated in a TIP plasmid by several methods. Shuttle vectors can be used to introduce deletions constructed by standard

recombinant DNA techniques (Cohen and Boyer, U.S. Pat. 4,237,224). Deletions with one predetermined end can be created by the improper excision of transposons (Koekman BP et al. (1979) Plasmid 2:347-357, and Ooms G et al. (1982) Plasmid 7:15-29). Hille J and Schilperoot R (1981) Plasmid 6:151-154, have demonstrated that deletions having both ends at predetermined positions can be generated by use of two transposons. The technique can also be used to construct "recombinant DNA" molecules in vivo.

The nopaline synthase gene has been used for insertion of DNA segments coding for drug resistance that can be used to select for transformed plant cells. In plant cells, the kanamycin resistance gene from Tn5 is not transcribed under control of its own promoter (Kemp JD et al. (18 May 1982) Beltsville Symp. VII, Beltsville, MD, to be published (1983) in Genetic Engineering: Applications to Agriculture, ed. Owens LD; and Fink CL (1982) supra). Bevan MW et al. (1983) Nature 304:184-187 and Fraley RT et al. (1983) Proc. Natl. Acad. Sci. USA 80:4803-4807, have inserted the kanamycin resistance gene (neomycin phosphotransferase II) from Tn5 behind (i.e. under control of) the nopaline promoter. The construction was used to transform plant cells which in culture displayed resistance to kanamycin and its analogs such as G418. Schell J et al. (18 January 1983) 15th Miami Winter Symp. (see also Marx JL (1983) Science 219:830), reported a similar construction, in which the methotrexate resistance gene (dihydrofolate reductase) from Tn7 was placed behind the nopaline synthase promoter. Transformed cells were resistant to methotrexate. Similarly, Herrera-Estrella L et al. (1983) Nature 303:209-213, have obtained expression in plant cells of enzymatic activity for octopine synthase and chloramphenicol acetyltransferase, an enzyme which in bacteria confers resistance to chloramphenicol, by placing the structural genes for these two enzymes under control of nos promoters.

Hall TC et al., U.S. application ser. no. 485,614, which is hereby incorporated by reference, have fused the ocs promoter and the 5' end of the octopine synthase structural gene to the structural gene for the bean seed protein phaseolin. A fusion protein having the amino terminus of octopine synthase and lacking the amino terminus of phaseolin was produced under control of the T-DNA promoter. The introns, which were contributed by the phaseolin sequences, were posttranscriptionally processed properly.

de Fraimond AJ et al. (1983) Biotechnol. 1:262-269; has reported on the construction a "mini-Ti plasmid". In the nopaline T-DNA there is normally only one site cut by the restriction enzyme KpnI. A mutant lacking the site was constructed and a KpnI fragment, containing the entire nopaline T-DNA, was isolated. This fragment together with a kanamycin resistance gene was inserted into pRK290; thereby resulting in a plasmid which could be maintained in A. tumefaciens and lacked almost all non-T-DNA Ti sequences. By itself, this plasmid was not able to transform plant cells. However when placed in an A. tumefaciens strain containing an octopine Ti plasmid, tumors were induced which synthesized both octopine and nopaline. The mini-Ti plasmid has also been transferred into plant cells when complemented with a Ti plasmid deleted for its own T-DNA. These results indicated that the non-T-DNA functions acted in trans with T-DNA, that the missing nopaline Ti plasmid functions were complemented by the octopine Ti plasmid, and that the nopaline "mini-Ti" was functional in the transformation of plant cells. A similar pair of complementing plasmids, each containing either octopine T-DNA or onc genes, has been constructed by Hoekema A et al. (1983) Nature 303:179-180.

Chilton et al. (18 January 1983) 15th Miami Winter Symp., also reported on the construction of a "micro-Ti" plasmid made by resectioning the mini-Ti with SmaI to delete essentially all of T-DNA but the nopaline synthase gene and the left and right borders. The micro-Ti was inserted into a modified pRK290 plasmid that was missing its SmaI site, and was employed in a manner similar to mini-Ti, with comparable results.

SUMMARY OF THE INVENTION

One object of this invention is to confer pest resistance, specifically insect resistance, to a plant. In pursuance of this goal, other objects are to stably insert a gene coding for an insecticidal protein into the genome of the plant cell, to have this gene expressed in plant tissues, for the expression to be either regulated or constitutive, and for the plant tissues to be in a normal plant. Another object is to provide novel specialized insecticidal tissues for a plant, in particular a means for producing on a normal dicot a gall which contains within its

tissue an insecticidal protein. Other objects and advantages will become evident from the following description.

The invention disclosed herein provides a plant comprising a genetically modified plant cell having an insecticide structural gene introduced and expressed therein under control of a plant expressible promoter. Further, the invention provides plant tissue comprising a plant cell whose genome includes T-DNA comprising an insecticide structural gene inserted in such orientation and spacing with respect to a plant expressible promoter as to be expressible in the plant cell under control of that promoter. Also provided are novel strains of bacteria containing and replicating T-DNA, as defined herein, the T-DNA being modified to contain an inserted insecticide structural gene in such orientation and spacing with respect to a plant expressible promoter as to be expressible in a plant cell under control of said promoter. Further, the invention provides novel plasmids having the ability to replicate in E. coli and comprising T-DNA, and further comprising an insecticide structural gene inserted within T-DNA contained within the plasmid, in such manner as to be expressible in a plant cell under control of a plant expressible promoter. Additionally, this invention discloses novel plasmids wherein the insecticide structural gene is capable of expression in E. coli or Bacillus subtilis, and furthermore discloses strains of bacteria harboring said bacterial expression plasmids.

The invention is exemplified in one of its embodiments by the insertion of the full length structural gene of Bacillus thuringiensis insect toxic protein into a sub-Ti plasmid so that the toxin gene is placed under the control of T-DNA plant active regulation sequences (ORF24). The sub-Ti plasmid containing the plant expressible crystal protein gene was introduced into tobacco cells. Tobacco plants regenerated from these transformed cells were found to express crystal protein at levels measurable by ELISA techniques but also the leaves of these plants were found to be toxic to insect larvae.

The present invention comprises an insecticide structural gene under control of a promoter expressible in plant cells, said promoter/gene combination being inserted into a plant cell by any means known to the art.

More specifically, in its preferred embodiment the invention disclosed herein further comprises expression in plant cells of an insecticide structural gene under control of a plant expressible promoter, after introduction via T-DNA, that is to say, by inserting the insecticide structural gene into T-DNA under control of a plant expressible promoter and introducing the T-DNA containing the insert into a plant cell using known means.

The invention is useful for genetically modifying plant tissues and whole plants by inserting useful insecticide structural genes from various bacterial species or strains. Such useful insecticide structural genes include, but are not limited to, the genes coding for insecticidal proteins as defined below, especially the crystal protein of Bacillus thuringiensis, related proteins, and the like. The invention is exemplified by introduction and expression of a structural gene for a crystal protein from B. thuringiensis var. kurstaki HD-73 into cotton or tobacco plant cells. Once plant cells expressing an insecticide structural gene under control of a plant expressible promoter are obtained, plant tissues and whole plants can be regenerated therefrom using methods and techniques well known in the art. The regenerated plants are then reproduced by conventional means and the introduced genes can be transferred to other strains and cultivars by conventional plant breeding techniques.

The introduction and expression of the structural gene for an insecticidal protein can be used to protect a crop from infestation with insect larvae such as hornworm (Manduca sp.) or European corn borer (Ostrinia nubilalis). Other uses of the invention, exploiting the properties of other insecticide structural genes introduced into other plant species will be readily apparent to those skilled in the art. The invention in principle applies to any introduction of an insecticide structural gene into any plant species into which foreign DNA (in the preferred embodiment T-DNA) can be introduced and in which said DNA can remain stably replicated. In general these taxa presently include, but are not limited to, gymnosperms and dicotyledenous plants, such as sunflower (family Compositae), tobacco (family Solanaceae), alfalfa, soybeans and other legumes (family Leguminosae), cotton (family Malvaceae), and most vegetables. Pests which may be controlled by means of the present invention

and the crops that may be protected from them include, but are not limited to, those listed in Tables 1 and 2, respectively. Because of its susceptibility to a number of larvae, cotton is an ideal choice for the insertion of an insecticidal protein gene. Each of the following is a major cotton pest and is also susceptible to the B. thuringiensis insecticidal protein: Heliothis zea (cotton bollworm), Pectinophora gossypiella (pink bollworm), Heliothis virescens (tobacco budworm), Trichoplusia ni (cabbage looper). Application of insecticidal protein prepared from sporulating B. thuringiensis does not control insects such as the pink bollworm in the field because of their particular life cycles and feeding habits. A plant containing in its tissues insecticidal protein will control this recalcitrant type of insect, thus providing advantage over prior insecticidal uses of B. thuringiensis. By incorporation of the insecticidal protein into the tissues of a plant, the present invention additionally provides advantage over such prior uses by eliminating instances of nonuniform application and the costs of buying and applying insecticidal preparations to a field. Also, the present invention eliminates the need for careful timing of application of such preparations since small larvae are most sensitive to insecticidal protein and the protein is always present, minimizing crop damage that would otherwise result from preapplication larval foraging.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 presents the sequence of the crystal protein gene of p123/58-10, described in Example 1.

Figure 2 presents a map of restriction sites and transcripts of the T-DNA of pTi15955.

Figure 3 is a diagram of a construction described in Example 2 of a recombinant DNA vector carrying an insecticide structural gene under control of a plant expressible promoter.

DETAILED DESCRIPTION OF THE INVENTION

The following definitions are provided, in order to remove ambiguities to the intent or scope of their usage in the specification and claims.

T-DNA: A segment of DNA derived from the transformation-inducing principle (TIP) which becomes integrated in the plant genome. As used herein, the term includes DNA originally derived from any tumor-inducing strain of Aerobacterium including A. tumefaciens and A. rhizogenes, the inserted segment of the latter sometimes referred to in the prior art as R-DNA. In addition, as used herein the term T-DNA includes any alterations, modifications, mutations, substitutions, insertions and deletions either naturally occurring or introduced by laboratory procedures, a principal structural requirement and limitation to such modifications being that sufficient of the right and left ends of naturally-occurring T-DNAs be present to insure the expected function of stable integration in the transformed plant cell genome which is characteristic of T-DNA. The T-DNA may itself be a composite or segments derived from a plurality of sources, naturally occurring or synthetic. In addition, the T-DNA must contain at least one plant expressible promoter, 5' or "upstream" from the site of insertion of the insecticide structural gene, in sufficiently complete form to control initiation of transcription and initiation of translation of an inserted insecticide structural gene. This promoter may be derived from a T-DNA gene, a plant gene, or any other gene having a promoter that is functional within a plant cell in at least one tissue and at least one developmental stage. Preferably, an insertion site will be provided "downstream" in the direction of transcription and translation initiated by the promoter (3' to the promoter), so located with respect to the promoter to enable an insecticide structural gene inserted therein to be expressed under control of the promoter, either directly or as a fusion protein. The T-DNA may also include a 3'-untranslated region downstream from the site of insertion of the insecticide structural gene, which may function to regulate termination of transcription, polyadenylation, and post-transcriptional RNA processing. Optionally, a fusion protein may also be formed between the insecticide structural gene and the 3' end of

the structural gene donating the 3'-untranslated region. The promoter and 3'-untranslated region elements may be derived from the same, or different pre-existing genes and may be derived from the same or different plant, T-DNA, or other sources. For example, an insecticide structural gene, as exemplified herein, could be nested between a plant gene promoter and 3' sequence from the same gene, or it could be a construct comprising the 3'-untranslated region of one gene and the promoter of another, derived from the same or different plant species or T-DNA. The coding region of a plant gene, as herein defined, may include a cDNA copy of the structural portion of a plant gene. The promoter and 3'-untranslated regions may also include modifications, either naturally or artificially induced, and may include chemically synthesized segments.

Plant promoter: As used herein includes regulatory elements of a plant gene, and may further include structural elements, of a plant gene said elements being exogenous to the genes of T-DNA itself. These include, but are not limited to, promoters of the genes for phaseolin and the small subunit of ribulose-1,5-bisphosphate carboxylase. Furthermore, a plant gene promoter is a region of the gene which provides for and may regulate the initiation of transcription and the initiation of translation. Additionally, the plant structural gene sequences (the region which codes for a protein in part or in whole and which may or may not contain one or more introns) may be introduced into T-DNA. (An intron is a region of a gene transcript which is posttranscriptionally removed before the mRNA is translated.) Expression under control of a plant promoter may take the form of direct expression in which the structural gene normally controlled by the promoter is removed in part or in whole and replaced by the inserted insecticide structural gene, a start codon being provided either as a remnant of the plant structural gene or as part of the inserted insecticide structural gene, or by fusion protein expression in which part or all of the insecticide structural gene is inserted in correct reading frame phase within the existing plant structural gene. In the latter case, the expression product is referred to as a fusion protein. The promoter segment may itself be a composite of segments derived from a plurality of sources, naturally occurring or synthetic. Sources of a plant promoter include, but are not limited to, plants listed in Table 2.

T-DNA promoter: Refers to any of the naturally occurring promoters commonly associated with integrated T-DNA. These include, but are not limited to, promoters of the "1.6" transcript, octopine synthase gene (ocs), nopaline synthase gene (nos), tms, tml, and tmr genes, and may depend in part on the TIP source of the T-DNA. Expression under control of a T-DNA promoter may take the form of direct expression in which the structural gene normally controlled by the promoter is removed in part or in whole and replaced by the inserted insecticide structural gene, a start codon being provided either as a remnant of the T-DNA structural gene or as part of the inserted insecticide structural gene, or by fusion protein expression in which part or all of the plant structural gene is inserted in correct reading frame phase within the existing T-DNA structural gene. In the latter case, the expression product is referred to as a fusion protein. The promoter segment may itself be a composite of segments derived from a plurality of sources, naturally occurring or synthetic.

Plant expressible promoter: As used herein includes the definitions for T-DNA promoter and plant promoter, supra. However, an essential aspect of the promoter component of the present invention is that the insecticide structural gene be under control of a promoter expressible in a plant cell. Therefore, plant expressible promoter additionally refers to any promoter expressible in a plant cell which is expressed in at least one tissue during at least one developmental stage. Sources might include, but need not be limited to, plant viruses (e.g. the promoters for the 35S and 19S transcripts of cauliflower mosaic virus, CaMV); animal viruses, nonplant eukaryotes (e.g. animals, yeast), or plastids (e.g. a chloroplast or prokaryotic promoter if the insecticide gene is to be inserted into chloroplast DNA). Properties and components of a promoter that is derived from a source that is not a plant DNA or T-DNA (e.g. "TATA boxes", ATG translational start sites, intron splicing sites, etc.) are the same as described supra for T-DNA promoters and plant promoters are also included within the present definition. The promoter segment may itself be a composite of segments derived from a plurality of sources, naturally occurring or synthetic.

Insecticide structural gene: As used herein includes that portion of an insecticide gene comprising a DNA segment coding for an insecticidal protein, polypeptide or portion thereof, possibly including a translational start codon, but lacking other functional elements of a bacterial gene that regulate initiation of transcription and initiation of translation, commonly referred to as the promoter region. (Note that in the present invention such bacterial functional elements may be present after transfer of the insecticide structural gene into T-DNA. However, because they are not functional within a plant cell, such elements are not referred to by the term "insecticide structural gene"). An insecticide structural gene may be derived in whole or in part from plasmid DNA, genomic DNA, cDNA and chemically synthesized DNA. It is further contemplated that an insecticide structural gene may contain one or more modifications in either the coding segments or untranslated regions which could affect the biological activity or chemical structure of the expression product, the rate of expression or the manner of expression control. Such modifications could include, but are not limited to, mutations, insertions, deletions, substitutions, and "silent" modifications that do not alter the chemical structure of the expression product but which affect intercellular localization, transport, excretion or stability of the expression product. The structural gene may constitute an uninterrupted coding sequence or it may include one or more introns, bounded by the appropriate plant functional splice junctions, which may be obtained from synthetic or a naturally occurring source. The structural gene may be a composite of segments derived from a plurality of sources, naturally occurring or synthetic, coding for a composite protein, the composite protein being insecticidal or being derived in part from an insecticidal protein.

Insecticidal protein: As used herein includes a bacterial protein toxic in any way to insects. This includes a protein or peptide that is directly or indirectly toxic or growth inhibitory under any circumstances to any insect. This also includes proteins that are toxic upon contact, ingestion, or respiration, where alone or in combination with other material, at any time within the life cycle of an insect, including egg, larva, pupa, nymph, and adult stages. This includes proteins toxic to

insects, especially those of the families Lepidoptera and Diptera, and those of the genera Ostrinia, Heliothis, Pectinophora, and Trichoplusia, e.g. M. sexta, O. nubilalis, H. zea, H. virescens, P. gossypiella, and I. ni. Other taxa that might be chosen as targets include, but are not limited to, those listed in Table 1. Examples of insecticidal proteins include, but are not limited to various varieties, listed in Table 3, of Bacillus thuringiensis or of other species of Bacillus, e.g. B. cereus, B. popilliae, and B. sphaericus. Genes that are used to construct or otherwise encode sequences encoding proteins toxic to insects include, but are not limited to, phospholipases, hyaluronidases, phosphatases, proteases, and the various crystal proteins of B. thuringiensis. The term crystal protein should be understood to refer to both the full-length protoxin and toxin forms, to toxic proteins related to the protein which is found in the crystalline inclusion bodies of Bacillus thuringiensis, and to artificial modifications of naturally occurring crystal proteins. Related proteins might be identified by nucleic acid or protein structural or sequence homology, immunological cross-reactivity, or cross-hybridization of nucleic acids.

Plant tissue: Includes differentiated and undifferentiated tissues of plants including, but not limited to roots, shoots, pollen, seeds, tumor tissue, such as crown galls, and various forms of aggregations of plant cells in culture, such as embryos and calluses. The plant tissue may be in planta or in organ, tissue, or cell culture, and may be derived from plants which include, but are not limited to; those listed in Table 2.

Plant cell: As used herein includes plant cells in planta and plant cells and protoplasts in culture, and may be derived from plants which include, but are not limited to those listed in Table 2.

Production of a genetically modified plant expressing an insecticide structural gene introduced via T-DNA combines the specific teachings of the present disclosure with a variety of techniques and expedients known in the art. In most instances, alternatives expedients exist for each stage of the overall process. The choice of expedients depends on variables such as the choice of the basic TIP or other vector systems for the intro-

duction and stable maintenance of the expressible insecticide structural gene, the plant species to be modified and the desired regeneration strategy, and the particular insecticide structural gene to be used, all of which present alternative process steps which those of ordinary skill are able to select and use to achieve a desired result. For instance, although the starting point for obtaining an insecticide structural gene is exemplified in the present application by DNA isolated from B. thuringiensis var. kurstaki HD-73, DNA of other insecticidal protein gene-carrying bacterial strains or recombinant DNA molecules might be substituted as long as appropriate modifications are made to the gene isolation and manipulation procedures. As novel means are developed for the controlled expression and/or stable insertion of foreign genes in plant cells, those of ordinary skill in the art will be able to select among those alternate process steps to achieve a desired result. The fundamental aspects of the invention are the nature and structure of the insecticide structural gene and its means of insertion and expression in a plant genome. The remaining steps of the preferred embodiment for obtaining a genetically modified plant include inserting the promoter/insecticide structural gene combination into T-DNA; transferring the modified T-DNA to a plant cell wherein the modified T-DNA becomes stably integrated as part of the plant cell genome, techniques for in vitro culture and eventual regeneration into whole plants, which may include steps for selecting and detecting transformed plant cells and steps of transferring the introduced gene from the originally transformed strain into commercially acceptable cultivars.

A principal feature of the present invention in its preferred embodiment is the construction of T-DNA having an inserted insecticide structural gene under control of a plant expressible promoter, or, most preferably, a T-DNA promoter, as these terms have been defined, supra. The insecticide structural gene must be inserted in correct position and orientation with respect to the desired promoter. Position has two aspects. The first relates to which side of the promoter the structural gene is inserted. It is known that the majority of promoters control initiation of transcription and translation in one direction only along the DNA. The region of DNA lying under promoter control is said to lie

"downstream" or alternatively "behind" or "3' to" the promoter. Therefore, to be controlled by the promoter, the correct position of plant structural gene insertion must be "downstream" from the promoter. (It is recognized that a few known promoters exert bidirectional control, in which case either side of the promoter could be considered to be "downstream" therefrom.) The second aspect of position refers to the distance, in base pairs, between known functional elements of the promoter, for example the transcription initiation site, and the translational start site of the structural gene. Substantial variation appears to exist with regard to this distance, from promoter to promoter. Therefore, the structural requirements in this regard are best described in functional terms. As a first approximation, reasonable operability can be obtained when the distance between the promoter and the inserted insecticide structural gene is similar to the distance between the promoter and the T-DNA gene it normally controls. Orientation refers to the directionality of the structural gene. That portion of a structural gene which ultimately codes for the amino terminus of the plant protein is termed the 5'-end of the structural gene, while that end which codes for amino acids near the carboxyl end of the protein is termed the 3'-end of the structural gene. Correct orientation of the insecticide structural gene is with the 5'-end thereof proximal to the promoter. An additional requirement in the case of constructions leading to fusion protein expression is that the insertion of the insecticide structural gene into the promoter-donated structural gene sequence must be such that the coding sequences of the two genes are in the same reading frame phase; a structural requirement which is well understood in the art. An exception to this requirement, of relevance to the present invention, exists in the case where an intron separates coding sequences derived from an insecticidal protein gene from the first coding segment of the insecticide structural gene. In that case, the insecticide structural gene must be provided with a splice site compatible with the upstream splice junction contributed by the noninsecticidal coding sequences, and the intron splice sites must be so positioned that the correct reading frame for the promoter-donated structural gene and the insecticide structural gene are restored in phase after the intron is removed by post-transcriptional processing. Differences in rates of expression or developmental control may be observed when a given

insecticide structural gene is inserted under control of different plant expressible promoters. Different properties, including, but not limited to such properties as stability, intercellular or intracellular localization or excretion, solubility, target specificity, and other functional properties of the expressed protein itself may be observed in the case of fusion proteins depending upon the insertion site, the length and properties of the segment of T-DNA protein included within the fusion protein and mutual interactions between the components of the fusion protein that effect folded configuration thereof; all of which present numerous opportunities to manipulate and control the functional properties of the insecticidal protein product, depending upon the desired physiological properties within the plant cell, plant tissue, and whole plant.

Location of the promoter/insecticide structural gene combination insertion site is not critical as long as the transfer function sequences immediately surrounding the T-DNA borders are not disrupted, since these regions appear from prior art studies to be essential for insertion of the modified T-DNA into the plant genome. Preferred insertion sites are those which lie in areas that are most actively transcribed, in particular the tnl gene and an area designated "1.6" lying in the HindIII-f fragment; and equivalent to transcript 24, as shown in Fig. 2. The term "1.6" is used herein to designate this actively transcribed region of T-DNA. The T-DNA into which the promoter/insecticide gene combination is inserted, is obtained from any of the TIP plasmids. The insecticide gene is inserted by standard techniques well known to those skilled in the art. The orientation of the inserted plant gene, with respect to the direction of transcription and translation of endogenous T-DNA genes is not critical, either of the two possible orientations is functional. Differences in rates of expression may be observed when a given gene is inserted at different locations within T-DNA, possibly because of such factors as DNA methylation and chromatin structure. Readily detectable levels of expression of a plant promoter from the phaseolin gene have been obtained where that gene was inserted into pTi15955, an octopine-type plasmid of A. tumefaciens at a SmaI site found within the tnl gene or a HpaI site found within tmr.

A convenient means for inserting a promoter/insecticide structural gene combination into T-DNA involves the use of a shuttle vector, as described supra, having segments of T-DNA (those segments between which insertion is desired) incorporated into a plasmid capable of replicating in E. coli. The T-DNA segment contains a restriction site, preferably one which is unique within the shuttle vector. The insecticide structural gene can be inserted at the unique site in the T-DNA sequences and the shuttle vector is transferred into cells of the appropriate Agrobacterium strain, preferably one whose T-DNA is homologous with the T-DNA segments of the shuttle vector. The transformed Agrobacterium strain is preferably grown under conditions which permit selection of a double-homologous recombination event which results in replacement of a pre-existing segment of the Ti plasmid with a segment of T-DNA of the shuttle vector. However, it should be noted that the present invention is not limited to the introduction of the promoter/insecticide structural gene combination into T-DNA by a double homologous recombination mechanism; a homologous recombination event with a shuttle vector (perhaps having only a single continuous region of homology with the T-DNA) at a single site or an insertion of a promoter/gene-carrying bacterial transposon will also prove an effective means for inserting that combination into T-DNA.

Following the strategy just described, the modified T-DNA can be transferred to plant cells by any technique known in the art. For example, this transfer is most conveniently accomplished either by direct infection of plants with the novel Agrobacterium strain containing an insecticide gene incorporated within T-DNA, or by cocultivation of the Agrobacterium strain with plant cells. The former technique, direct infection, results in due course in the appearance of a tumor mass or crown gall at the site of infection. Crown gall cells can be subsequently grown in culture and, under appropriate circumstances known to those of ordinary skill in the art, regenerated into whole plants that contain the inserted T-DNA segment. Using the method of cocultivation, a certain proportion of the plant cells are transformed; that is to say have T-DNA transferred therein and inserted in the plant cell genome. In either case, the transformed cells must be selected or screened to distinguish them from untransformed cells. Selection is most readily accomplished by

providing a selectable marker incorporated into the T-DNA in addition to the insecticide structural gene. Examples include either dihydrofolate reductase or neomycin phosphotransferase expressed under control of a nopaline synthase promoter. These markers are selected by growth in medium containing methotrexate or kanamycin, respectively, or their analogs. In addition, the T-DNA provides endogenous markers such as the gene or genes controlling hormone-independent growth of Ti-induced tumors in culture, the gene or genes controlling abnormal morphology of Ri-induced tumor roots, and genes that control resistance to toxic compounds such as amino acid analogs, such resistance being provided by an opine synthase. Screening methods well known to those skilled in the art include assays for opine production, specific hybridization to characteristic RNA or T-DNA sequences, or immunological assays for specific proteins, including ELISAs (acronym for "enzyme linked immunosorbant assay"), radioimmune assays and "western" blots. Additionally the toxic properties of expressed insecticidal protein can be used to identify transformed tissue.

An alternative to the shuttle vector strategy involves the use of plasmids comprising T-DNA or modified T-DNA, into which an insecticide structural gene is inserted, said plasmids being capable of independent replication in an Agrobacterium strain. Recent evidence reviewed in the Background indicates that the T-DNA of such plasmids can be transferred from an Agrobacterium strain to a plant cell provided the Agrobacterium strain contains certain trans-acting genes whose function is to promote the transfer of T-DNA to a plant cell. Plasmids that contain T-DNA and are able to replicate independently in an Agrobacterium strain are herein termed "sub-TIP" plasmids. A spectrum of variations is possible in which the sub-TIP plasmids differ in the amount of T-DNA they contain. One end of the spectrum retains all of the T-DNA from the TIP plasmid, and is sometimes termed a "mini-TIP" plasmid. At the other end of the spectrum, all but the minimum amount of DNA surrounding the T-DNA border is deleted, the remaining portions being the minimum necessary to be transferrable and integratable in the host cell. Such plasmids are termed "micro-TIP". Sub-TIP plasmids are advantageous in that they are small and relatively easy to manipulate directly, eliminating the need to transfer the gene to T-DNA from a shuttle vector by homologous recombination. After the

desired structural gene has been inserted, they can easily be introduced directly into a plant cell containing the trans-acting genes that promote T-DNA transfer. Introduction into an Agrobacterium strain is conveniently accomplished either by transformation of the Agrobacterium strain or by conjugal transfer from a donor bacterial cell, the techniques for which are well known to those of ordinary skill. For purposes of introduction of novel DNA sequences into a plant genome, TIP plasmids and sub-TIP plasmids should be considered functionally equivalent.

Although the preferred embodiment of this invention incorporates a T-DNA-based Agrobacterium-mediated system for incorporation of the insecticide gene into the genome of the plant which is to be made insect resistant, other means for transferring and incorporating the gene are also included within the scope of this invention. Other means for the stable incorporation of the insecticide gene into a plant genome additionally include, but are not limited to, use of vectors based upon viral genomes, minichromosomes, transposons, and homologous or nonhomologous recombination into plant chromosomes. Alternate forms of delivery of these vectors into a plant cell additionally include, but are not limited to, direct uptake of nucleic acid, fusion with vector-containing liposomes, microinjection, and encapsidation in viral coat protein followed by an infection-like process. Systems based on Agrobacterium cells and TIPs can be used to transform dicots and gymnosperms by transfer of DNA from a bacterium to a plant cell; systems based on alternate vectors or means for vector delivery may be used to transform all gymnosperms and all angiosperms, including both monocots and dicots.

Regeneration of transformed cells and tissues is accomplished by resort to known techniques. An object of the regeneration step is to obtain a whole plant that grows and reproduces normally but which retains integrated T-DNA. The techniques of regeneration vary somewhat according to principles known in the art, depending upon the origin of the T-DNA, the nature of any modifications thereto and the species of the transformed plant. Plant cells transformed by an Ri-type T-DNA are readily regenerated, using techniques well known to those of ordinary skill, without undue experimentation. Plant cells transformed by Ti-type T-DNA can be regenerated, in some instances, by the proper manipulation of hormone levels in

culture. Preferably, however, the Ti-transformed tissue is most easily regenerated if the T-DNA has been mutated in one or both of the tmr and tms genes. Inactivation of these genes returns the hormone balance in the transformed tissue towards normal and greatly expands the ease and manipulation of the tissue's hormone levels in culture, leading to a plant that is readily regenerated because of its more normal hormone physiology. It is important to note that if the mutations in tmr and tms are introduced into T-DNA by double homologous recombination with a shuttle vector, the incorporation of the mutations must be selected in a different manner than the incorporation of the promoter/insecticide structural gene. For example, in the former instance one might select for chloramphenicol resistance while the latter selection might be for resistance to kanamycin. The inactivation of the tms and tmr loci may be accomplished by an insertion, deletion, or substitution of one or more nucleotides within the coding regions or promoters of these genes, the mutation being designed to inactivate the promoter or disrupt the structure of the protein. (The construction of suitable mutations has been exemplified by Hall TC et al., ser. nos. 485,613 and 485,614.) In some instances, tumor cells are able to regenerate shoots which carry integrated T-DNA and express T-DNA genes, such as nopaline synthase, and which also express an inserted insecticide structural gene. The shoots can be maintained vegetatively by grafting to rooted plants and can develop fertile flowers. The shoots thus serve as parental plant material for normal progeny plants carrying T-DNA and expressing the insecticide structural gene inserted therein.

The genotype of the plant tissue transformed is often chosen for the ease with which its cells can be grown and regenerated in in vitro culture. Should a cultivar of agronomic interest be unsuitable for these manipulations, a more amenable variety is first transformed. After regeneration, the newly introduced foreign insecticidal protein gene is readily transferred to the desired agronomic cultivar by techniques well known to those skilled in the arts of plant breeding and plant genetics. Sexual crosses of transformed plants with the agronomic cultivars yielded initial hybrids. These hybrids can then be back crossed with plants of the desired genetic background. Progeny are continuously screened and selected for the continued presence of integrated T-DNA or for the new

phenotype resulting from expression of the inserted insecticidal protein gene. In this manner, after a number of rounds of back crossing and selection, plants can be produced having a genotype essentially identical to the agronomically desired parents with the addition of the inserted insecticidal protein gene.

In an alternative method for conferring insect resistance to a crop, one may infect plants within a field which is to be protected with an Agrobacterium cell harboring a TIP plasmid having undisabled T-DNA which carries an expressible insecticidal protein gene. We have found that larvae will feed on crown gall tissue. When insect larvae infesting the field eat transformed tissue containing an insecticide gene, they will be affected by the insecticidal protein within that tissue. The Agrobacterium and TIP might additionally encode genes for insect attractants. The presence of such attractants in transformed tissue will increase the insects' preference for such tissue as a food source relative to the rest of the crop material in the field.

EXAMPLES

The following Examples utilize many techniques well known and accessible to those skilled in the arts of molecular biology and manipulation of TIPs and Agrobacterium; such methods are fully described in one or more of the cited references if not described in detail herein. Enzymes are obtained from commercial sources and are used according to the vendor's recommendations or other variations known to the art. Reagents, buffers and culture conditions are also known to those in the art. Reference works containing such standard techniques include the following: Wu R, ed. (1979) Meth. Enzymol. 66, Wu R et al., eds. (1983) Meth. Enzymol. 100 and 101, Grossman L and Moldave K, eds. (1980) Meth. Enzymol. 65, Miller JH (1972) Experiments in Molecular Genetics, Davis R et al. (1980) Advanced Bacterial Genetics, Schleif RF and Wensink PC (1982) Practical Methods in Molecular Biology, and Maniatis T et al. (1982) Molecular Cloning. Additionally, Latche RF et al. (1983) Genet. Engin. 4:1-56, make useful comments on DNA manipulations.

Textual use of the name of a restriction endonuclease in isolation, e.g. "BclI", refers to use of that enzyme in an enzymatic digestion, except in a diagram where it can refer to the site of a sequence susceptible to action of that enzyme, e.g. a restriction site. In the text, restriction sites are indicated by the additional use of the word "site", e.g. "BclI site". The additional use of the word "fragment", e.g. "BclI fragment", indicates a linear double-stranded DNA molecule having ends generated by action of the named enzyme (e.g. a restriction fragment). A phrase such as "BclI/SmaI fragment" indicates that the restriction fragment was generated by the action of two different enzymes, here BclI and SmaI, the two ends resulting from the action of different enzymes. Note that the ends will have the characteristics of being "blunt" or "sticky" (i.e. having a single-stranded protuberance capable of base-pairing with a complementary single-stranded oligonucleotide) and that the sequence of a sticky-end will be determined by the specificity of the enzyme which produces it.

In these Examples, special symbols are used to make sequences more easily understood. Sequences that code for proteins are underlined, and codons are separated with slashes (/). The positions of cuts or gaps in each strand caused by restriction endonucleases or otherwise are indicated by the placement of asterisks (*).

Plasmids, and only plasmids, are prefaced with a "p", e.g., pTi15955 or pKS-4, and strain parenthetically indicate a plasmid harbored within, e.g., A. tumefaciens (pTi15955) or K802(pKS-4). Table 4 provides an index useful for identifying plasmids and their interrelationships. Table 5 provides a list of deposited strains.

Example 1

The first step in developing an insect resistant crop was to clone the insecticidal protein gene of B. thuringiensis var. kurstaki HD-73, which is on deposit with the Agricultural Research Culture Collection, Northern Regional Research Laboratory, Peoria, IL, and has NRRL number B-4488.

1.1 Cloning the *Bacillus thuringiensis* insecticidal protein gene

The 50 megadalton (MD) plasmid was enriched from HD-73 using sucrose gradient centrifugation. A HD-73 library was constructed by first digesting this plasmid with HindIII. The resulting fragments were mixed with and ligated to HindIII-linearized pBR322 (Bolivar F et al. (1978) Gene 2:95-113) and transformed into E. coli HB101. Ampicillin-resistant tetracycline-sensitive transformants were screened by digesting isolated plasmid DNA with HindIII and choosing those clones with 6.6 kilobase pair (kbp) inserts. Colonies containing plasmids p123/58-3 and p123/58-10 were selected from the HD-73 library for further analysis using an insect bioassay (see Example 8). These clones were grown in L-broth and a 250 fold concentrated cell suspension was sonicated and the extract applied to the surface of insect diet. Neonatal Manduca sexta (tobacco hornworm) larvae were placed on the diet for one week. Insect larvae fed extracts of strains harboring p123/58-3 or p123/58-10 did not grow and all larvae died in 2 to 5 days. There was no apparent difference between the larvae fed these extracts and those fed insecticidal protein purified from cells of B. thuringiensis.

Restriction enzyme analysis of p123/58-3 and p123/58-10 showed that the two plasmids were identical except for having the 6.6 kbp B. thuringiensis DNA fragment inserted into the pBR322 vector in opposite orientations. Note that either of these two plasmids can be converted to the other by digestion with HindIII, religation, and transformation into HB101 followed by appropriate selection and screening steps.

p123/58-10 was used to further probe the transformants from the HD-73 plasmid library. Sixteen of the 572 colonies hybridized to the insert of clone p123/58-10 and all had the characteristic 6.6 kbp HindIII fragment. Further restriction enzyme analysis showed these clones to all be one of the two possible orientations in pBR322 of the same DNA fragment. This suggested there could be a single crystal protein gene in strain HD-73. That these clones represent the only insecticidal protein gene in HD-73 was confirmed by hybridizing labeled p123/58-10 probe to Southern blots of HD-73 plasmid DNA digested with HindIII, BglII or

SalI. None of these enzymes has a restriction site in our cloned crystal protein gene. Hybridization results showed a single band of B. thuringiensis cellular DNA hybridized with p123/58-10 and further indicated that HD-73 has a single insecticidal crystal protein gene. We have identified a number of other clones by hybridization with a probe made from p123/58-10. Restriction mapping has shown that these clones are all identical to either p123/58-3 or p123/58-10, further supporting the conclusion that the HD-73 has a single crystal protein gene.

1.2 Immunological analysis

Analyses on the protein produced in the E. coli clones shows that p123/58-3 and p123/58-10 encoded protein that formed a precipitin band with antiserum to B. thuringiensis insecticidal protein in Ouchterlony diffusion slides. Cell extracts were analyzed on 10% SDS-poly-acrylamide gels, transferred to nitrocellulose, and immunological reactions done with antibody and ¹²⁵I-protein A (Western blots, Example 7). No band was found at 130 kD (kilodalton) where denatured protoxin is observed; however, a peptide of about 67 kD was seen that binds crystal protein antibody (Western blots done as in Example 7), and was identical in size to activated toxin. This peptide accounted for approximately 0.1% of the total E. coli protein.

1.3 Sequence analysis

We compared our DNA sequence results (Fig. 1), obtained by methods well known to those skilled in the art of DNA sequencing (e.g. see Maxam AM and Gilbert W (1980) Meth. Enzymol. 65:499-560), with published sequences (see Background). The published sequences showed only partial homology with our own sequence. An open reading frame of about 2.8 kbp was observed which was bounded at the 5'-end by a translational start signal (ATG) and did not stop before encountering the HindIII site at the junction between the B. thuringiensis DNA and the pBR322 vector. The size of the protein encoded by this open reading frame from the ATG to the HindIII site is greater than that of the 67 kD protein that we observed to be translated in E. coli cells but less than what is needed to encode the 130 kD native crystal protein. That the exact means of translational ter-

mination in the pBK322 encoded read-thru peptide was not important was suggested by the finding that insecticidal activity was encoded by B. thuringiensis DNA inserts having either orientation within the pBK322 vector. Presumably the initially translated amino acid residues carboxy-terminal to the ultimate carboxy-terminus of the translated polypeptide were removed in E. coli by a proteolytic process similar to that which naturally activates the crystal protein.

Example 2

This example teaches the insertion of the Bacillus thuringiensis insecticide gene between a T-DNA gene promoter and a polyadenylation (poly(A) addition) signal, the transfer of the insecticide gene to various plant species via a Ti plasmid, and the regeneration of plants expressing this gene under control of the T-DNA promoter. A large part of the strategy used in this construction is diagrammed in Fig. 3, which represents plasmids schematically and is not necessarily drawn to scale.

2.1 Introduction of BamHI site into the insecticidal protein gene

A BamHI site is introduced into the insecticidal protein gene of p123/58-10 at a location just 5' to the start of the coding sequence. The wild type base sequence (b) and the changed bases in an oligonucleotide primer (a) are as follows:

- BamHI
- a) 5' AGATGGAG*GATCCTT ATG GAT AAC AAT 3'
- b) ...AGATGGAG GTAACIT/ATG/GAT/AAC/...
- Met Asp Asn Asn

The changed bases are the underlined ATC sequence in (a). Note that good hybridization properties are insured because only three out of 27 base-pairs are changed.

p123/58-10 is digested with HindIII and is mixed with and ligated to HindIII-linearized mWB2344 RF (replicative form) DNA. The mixture is transformed into JM103 and transformed colonies are screened by plasmid isolation followed by restriction analysis for the presence of insertion

of a single copy of the insecticidal protein gene-bearing fragment. Vectors containing the two possible orientations are labeled M13-Bt-A and M13-Bt-S. They have the antisense and sense strands, respectively, of the insecticide structural gene when in viral form.

M13-Bt-A is hybridized with the oligonucleotide primer, 5'AGATGGAGGATCCTTATGGATAACAAT3', previously synthesized as described in Example 10.1. The oligonucleotide: M13-Bt-A hybrid is incubated with the Klenow fragment of *E. coli* DNA polymerase I, covalently closed circular DNA (cccDNA) is enriched, and the mixture is transformed into JM103. The virions produced by transformants are isolated and used to infect cells at a low multiplicity of infection. RF DNA is isolated from a number of the infected colonies and is characterized by restriction mapping. Clones derived from the mutant oligonucleotide-primed strand are identified by the presence of a novel BamHI site at the 5'-end of the insecticide structural gene, and one such vector is designated M13-Bt-A(Bam).

M13-Bt-A(Bam) RF DNA is digested with BamHI and HindIII, and is mixed with and ligated to a linker, synthesized as described in Example 10.1, having the following structure:

```

      HindIII          BamHI
      5'AGCTAGCTGACTAG3'
      3'TCGACTGATCCTAG5'

```

Note that this linker contains translational stop signals (underlined) in all three possible reading-phases. The linkers are trimmed by digestion with BamHI and an insecticide structural gene-bearing DNA fragment is purified by agarose gel electrophoresis.

2.2 Construction and modification of a promoter vehicle

The T-DNA "1.6" gene is summarized as follows:

```

      Clal          960 bp 250 bp Clal 60 bp 50 bp
      5'...TACACCAAT*CG/ATG/GAC/ATG/.../TGA/...AT*CGAT....AAATAA...AAATAA...3'
      promoter      Met Asp met ....stop      polyadenylation signals

```

By removing the ClaI fragment, the promoter region of the "1.6" gene can be brought next to the 3'-downstream region of the gene. This 3' region includes polyadenylation signals. The resulting structure is summarized as follows:

```

      ClaI           60 bp           50 bp
5'...ATACACCAAAT*CGATAGT.....AAATAA.....AAATAAAA...3'
   promoter                polyadenylation signals

```

pKS111, which is a pRK290 clone corresponding to the T-DNA clone p403 (which encodes the "1.6" gene which was described in the Detailed Description, transcript 24 in Fig. 2, see also Fink CF (1982) M.S. thesis, University of Wisconsin-Madison), is digested with ClaI and then religated. The ligation mix is transformed into E. coli K802 (Wood WB (1966) J. Mol. Biol. 16:116) and selected for tetracycline resistance. Plasmids are isolated by doing "minipreps" (Plasmid preparations from small volume cell cultures) and restriction maps are obtained to prove the structure. The new vehicle, pKS-proI (see Hall TC et al., US application ser. no. 485,614), can be linearized by ClaI.

pKS-proI grown in K802 was cut with ClaI. After converting sticky-ends to blunt-end with the Klenow fragment of E. coli DNA polymerase I, the DNA was mixed with and ligated to a BamHI linker. The resulting mixture was digested with ClaI to remove religated pKS-proI, and transformed into K802. Plasmids from tetracycline resistant transformants are screened by restriction analysis and a plasmid having the ClaI site at the ATG translational start replaced with a BamHI site is designated pKS-proI(Bam).

2.3 Introduction of a Kanamycin Resistance Gene into pKS-proI(Bam)

It is advantageous to have a kanamycin resistance (kan) gene inserted next to the promoter/insecticide gene combination so as to allow selection of double homologous recombinants after a triparental mating. The source of kan was pKS-4 (Example 2.5). In pKS-4 the kan gene is flanked on one side by a ClaI site. In order to remove a kan gene bearing fragment from pKS-4 with ClaI (i.e. on a "ClaI/kan" fragment) it is necessary to

introduce a ClaI site into that plasmid on the opposite side of kan from the existing ClaI site. This is accomplished by converting a conveniently positioned BamHI site (5'...G*GATCC...3') to the specificity of ClaI (5'...AT*CGAT...3').

pKS-4 is linearized by digestion with BamHI, thereby generating sticky-ends having the following structures:

```

5'...G          GATCC...3'
3'...CCTAG      C...5'

```

The recessed ends of this structure is filled in by incubation with the Klenow fragment of E. coli DNA polymerase I, forming the following blunt-ends:

```

...GGATC      GATCC...
...CCTAG      CTAGG...

```

When these ends were blunt end ligated together, the resulting suture has the following sequence:

```

      ClaI
...GGA1*CG ATCC...
...CCTA GC*TAGG...

```

Note that the resulting structure is susceptible to the action of ClaI but not to that of BamHI.

Alternatively to the above construction, one may convert the BamHI site, or another conveniently located restriction site, into a ClaI site by use of the appropriate linkers. pKS-4 was digested with SmaI mixed with and ligated to ClaI/blunt-ended linkers having the sequence 5'CATCGATG3', digested with ClaI, religated, and transformed into K802. Plasmids isolated from transformants resistant to kanamycin were screened for presence of a novel ClaI site in the position formally occupied by a SmaI site. A ClaI/kan fragment can be isolated from such a plasmid. The plasmid is designated pKS4.2.

When grown in E. coli K802, pKS-proI(Bam) is methylated at two remaining ClaI sites: one is about 145 bases from the the promoter-polyadenylation junction (this is about 30 bases past the second polyadenylation site), the other is about 200 bases from the right hand p403 EcoRI site (see Fig. 2). Methylation blocks cutting by the ClaI restriction endonuclease at an otherwise susceptible site. Therefore, these methyla-

tions protect these sites and effectively direct action of the ClaI enzyme to other sites. pKS-proI(Bam) is transferred to and grown in E. coli GM33, a strain that does not methylate adenosine residues in DNA, so that the otherwise methylated ClaI sites can be cut. After purification of that plasmid from GM33 (pKS-proI(Bam)), a partial digestion is done with ClaI and the resulting mixture is ligated with the ClaI/kan fragment described above. After transformation into E. coli K802, transformants are selected on tetracycline and kanamycin containing media. After plasmid isolation and restriction mapping, a clone having the desired construction is identified and the plasmid found in this clone is labeled p11-83a (Fig. 3).

p11-83a has a kan gene-bearing fragment ligated into the "middle" ClaI site about 30 bp past the second polyadenylation site. The BamHI fragment of the insecticide gene, isolated from the modified vector constructed in Example 2.1, is now ligated into the BamHI site of BamHI-linearized p11-83a that has been transferred to and grown in K802 and is methylated. After transformation into K802, tetracycline and kanamycin selection, plasmid isolation, and restriction enzyme mapping, the desired construction having the insecticide structural gene inserted between the pTi15955 "1.6" promoter and polyadenylation site is identified, and the plasmid harbored therein is labeled p11-83b (Fig. 3).

2.4 Introduction of p11-83b into Ti Plasmids

p11-83b is introduced into pTi15955, pTiA66 (equivalent to pTi15955 but having a nonfunctional tms gene), and mutants deleted in gene affecting regeneration by homologous recombination (Example 10). Tobacco plants are transformed by a system described in Example 6, and transformants are identified by Southern and Northern blots (techniques well known to those skilled in the art) with appropriate probes and by the presence of octopine and crystal protein. Transformed tobacco tissue is lethal to tobacco hornworms. Tobacco plants are regenerated from transformed cells as described in Example 6, and entered into breeding programs. Fields of regenerated plants and their insecticidal protein-containing descendants are resistant to infestation by larvae of insects such as tobacco hornworm by virtue of the toxic effect such larvae experience when eating tissue from such plants.

2.5 Cloning and isolation of a kanamycin resistance gene

pRZ102 (Jorgenson RA et al. (1979) Mol. gen. Genet. 177:65-72), a ColE1 plasmid carrying a copy of the transposon Tn5, was digested with BamHI and HindIII, mixed with pBR322 previously linearized with the same two enzymes, ligated, and transformed into K802. Plasmids, isolated from transformants selected for resistance to both ampicillin and kanamycin were restriction mapped and one having the structure shown in Fig. 3 was labeled pKS-4. pKS-4 DNA may be isolated from E. coli C600 (pKS-4) which has been deposited as NRRL B-15394.

Example 3

This example teaches another method of inserting an expressible gene for the B. thuringiensis insecticidal protein into a plant genome. The shuttle vector is similar to that used by Fink CL (1982) M.S. thesis, University of Wisconsin-Madison, to put the nos gene into an octopine Ti plasmid. In the present invention, the protein coding sequences for nos are removed and replaced with an insecticidal gene before insertion into the Ti plasmid. The eventual result is an octopine-type Ti plasmid carrying an insecticide gene expressible in plant cells under control of a nopaline synthase promoter.

3.1 Moving the nos gene into M13mp7

pCF44 DNA (Fink, supra) was digested with XhoI, religated to itself, and transformed back into K802. Plasmid DNA isolated from ampicillin-resistant transformants was analyzed with restriction enzymes. A plasmid having a single XhoI site within its Ti plasmid-derived DNA sequences was designated pCF44A. The single XhoI site was the result of the deletion of a DNA fragment between the two pCF44 XhoI sites. Deletion of this XhoI fragment resulted in the complete removal of two inconvenient ClaI sites.

pCF44A DNA was digested with HindIII and BamHI, mixed with and ligated to pBR322 which had been digested with the same two restriction enzymes and transformed into K802. Ampicillin resistant transformants

were selected and screened by restriction enzyme analysis of plasmid DNA and a colony was identified which contained a nos gene-containing plasmid, labeled pNS5.

pNS5 DNA, having been isolated from a nonmethylating host, was digested with BclI and BamHI and was mixed with and ligated to a double-stranded circular replicative form (RF) of the single-stranded DNA vector M13mp7 which had been linearized with BamHI. After transformation of the mixture into JM103 and selection of white plaques, two colonies were identified by restriction mapping after RF isolation, designated M13-1 and M13-3, contained the sense and antisense strands, respectively, when in single-stranded form.

3.2 Placement of an NcoI site behind the nos promoter

An oligonucleotide primer having the sequence 5'AGTCTCATACTCACTCTCAATCCAAATAATCTGCCCATGGAT3' was synthesized as described in Example 10.1. This oligonucleotide was changed at the underlined base from the naturally occurring sequence at the 5'-end of the nos structural gene. The change resulted in the introduction of an NcoI site, 5'...C*CATGG...3', at the AUG translational start of the nos gene. The oligonucleotide was hybridized to circular single-stranded M13-3 DNA isolated from virions which had been sedimented out of culture medium. The oligonucleotide:M13-3 hybrid was incubated with DNA ligase and the Klenow fragment of E. coli DNA polymerase I, covalently closed circular DNA (cccDNA) was enriched, and the mixture was transformed into JM103. The virions produced by transformants were isolated and used to infect cells at a low multiplicity of infection. RF DNA was isolated from a number of these infected colonies and characterized by restriction mapping. Clones derived from the mutant oligonucleotide-primed strand were identified by the presence of a single NcoI site, which allowed them to be linearized by that enzyme. The mutated clones were further characterized to localize the NcoI site by digestion with ClaI, BamHI (to identify linearized molecules), and ClaI together with NcoI. The mutated M13-3 vector was labeled M13-3A/B18a.

3.3 Moving the insecticide gene into M13mp8

p123/58-10 DNA (Example 1.1) was digested with EcoRI and mixed with and ligated to EcoRI-linearized M13mp8 RF DNA. After transformation of the mixture into JM103 and selection of white plaques, two colonies having the insecticide gene-carrying fragment inserted in opposite orientations, were identified by restriction mapping. They were labeled MBT14 and MBT3 and respectively had the sense and antisense strands when in single stranded form.

3.4 Placement of an NcoI site at the insecticide gene translation start

An oligonucleotide primer having the sequence 5'GAGGTAACCCATGGATAACAAT3' is synthesized as described in Example 10.1. This oligonucleotide is changed at the two underlined bases from the naturally occurring sequence at the 5'-end of the insecticide structural gene. The change results in the introduction of an NcoI site, 5'...C*CATGG...3', at the ATG translational start of the insecticide gene. The oligonucleotide is hybridized to circular single-stranded MBT3 DNA isolated from virions which had been sedimented out of culture medium. The oligonucleotide:MBT3 hybrid is incubated with DNA ligase and the Klenow fragment of E. coli DNA polymerase II; cccDNA is enriched, and the mixture is transformed into JM103. The virions produced by the transformants are isolated and used to infect cells at a low multiplicity of infection. RF DNA is isolated from a number of these infected colonies and characterized by restriction mapping. Clones derived from the mutant oligonucleotide-primed strand are identified by the presence of a single coI site which allows them to be linearized by that enzyme. The mutated clone is further characterized by restriction enzyme analysis and the presence of the mutant sequence is confirmed by sequencing. The plasmid having the desired sequence is labeled MBT3(Nco).

3.5 Assembly of a plant expressible insecticide gene in a shuttle vector

NcoI- and HindIII-digested MBT3(Nco) RF DNA is mixed with and ligated to a linker, synthesized as described in Example 11.1, having the following structure:

HindIII-end BamHI
5'AGCTGACTAACTAG3'
3'CTGATTGATCCTAG5'

This linker encodes stop codons (underlined) in all three reading phases, and is ended by a functional BamHI site and a HindIII compatible sticky-end incapable of reconstructing a HindIII site. The insecticide gene-bearing DNA fragment is then trimmed by digestion with NcoI and BamHI and is isolated by agarose gel electrophoresis.

pKS111-N (Fink, supra) is a plasmid having a nos gene inserted in Tn5 DNA (from pKS-4) which has a functional kan gene, which is itself inserted in the T-DNA of pKS111. pKS111-N is linearized with SstII and digested to completion with BamHI. M13-3A/B13a is digested with NcoI and SstII and the SstII/NcoI promoter fragment is isolated by agarose gel electrophoresis. The SstII/NcoI promoter and NcoI/BamHI gene fragments are mixed with and ligated to the pKS111-N SstII/BamHI reaction products. The ligation mixture is then transformed into E. coli K802. Plasmids isolated from transformants resistant to kanamycin and tetracycline are subjected to restriction enzyme analysis and colonies harboring plasmids identical to pKS111-N except for replacement of a 5'-portion of the nos gene with an insecticide structural gene are identified. Such a plasmid is designated pKS111-NpBt.

3.6 Insertion into TIP plasmids, plant infection and regeneration

E. coli K802(pKS111-NpBt) is mated with A. tumefaciens as described in Example 9. The Agrobacterium strains chosen harbor TIP plasmids, based on pTi15955, containing mutations, such as those described in the Background, which facilitate regeneration. Homologous recombinants are selected as described in Example 9 and characterized by restriction mapping. The efficacy of the construction is quickly tested by infection of

sunflower stems. The resulting galls are assayed by ELISA and Western blots as described in Example 7 and by bioassay as described in Example 8. As described in Example 6, the Agrobacterium strains are used to infect tobacco cells which are then regenerated. The resulting plants are used as breeding stock to be crossed with various commercial varieties for which insect resistance properties are desired. Regenerated plants and fields of their insecticidal protein-containing descendants are resistant to infestation by larvae of insects such as tobacco hornworm by virtue of the toxic effect such larvae experience when eating tissue from such plants.

Example 4

This example teaches another method of inserting an expressible gene for the B. thuringiensis insecticidal protein into a plant genome. The strategy is similar to that described in Example 3 but differs in that a plant promoter is used instead of a T-DNA promoter. The plant gene providing the promoter is phaseolin, which has been shown to be active in species other than its source, the bean Phaseolus vulgaris L.

4.1 Moving the phaseolin gene into M13mp7

BamHI linkers, having the sequence 5'GGATCC3', are annealed to form double-stranded structures, and blunt-end ligated to form concatamers. These concatamers are partially digested with BamHI to expose 5'GATC...3' sticky-ends, which are compatible with sticky-ends generated by the enzymes BamHI, BclI, BglII, MboI, Sau3AI, and XhoII (5'GATC...3'). A Charon 24A phage clone of phaseolin, 177.4 (Sun SM et al. (1981) Nature 289:37-41, Slighton JL et al. (1983) Proc. Natl. Acad. Sci. USA 80:1897-1901, also designated AG-PVPh177.4) is digested with BglII and BamHI, mixed with and ligated to the concatenated linkers, and completely digested with BamHI to trim the linkers and expose the BamHI sticky-ends, a 3.8 kbp fragment containing the phaseolin gene and 5'- and 3'-flanking sequences is isolated by agarose gel electrophoresis followed by elution. This fragment has BamHI sites at either end as the BamHI/BglII linkage site is not susceptible to the action of either enzyme. The 3.8 kbp BglII/BamHI fragment can also be obtained from p8.8; a pBR322-based subclone of 177.4.

The 3.8 kbp fragment is mixed with and ligated to BamHI-linearized M13mp7 RF. After transformation of the mixture into JM103 and selection of white plaques, two colonies are selected after characterization by restriction and hybridization analysis of RFs and phage DNA. The viral forms of M13-3.8A and M13-3.8S are respectively found to be carrying the antisense and sense strands of the phaseolin gene.

4.2 Placement of an NcoI site behind the phaseolin promoter

The Phaseolus DNA of M13-3.8A has an NcoI site about 450 bp upstream from the phaseolin transcriptional start. The presence of this site will be inconvenient when one wishes to cleave the plasmid at the NcoI site to be introduced at the phaseolin translational start. Isolated M13-3.8A RF DNA is linearized with NcoI and the 5'-protruding ends are filled in by the action of the Klenow fragment of E. coli DNA polymerase I. After blunt-end ligation and transformation into JM103, RF DNAs are isolated and characterized by restriction mapping. A colony is chosen which harbors a vector, labeled M13-3.8Ac, which is lacking the NcoI site of the Phaseolus DNA but is otherwise unchanged from M13-3.8A.

An oligonucleotide primer having the sequence 5'ATACTACTCTACCATGGTGAGAGCAAGGG3' is synthesized as described in Example 10.1. This oligonucleotide is changed at the underlined bases from the naturally occurring sequence at the 5'-end of the phaseolin gene. The oligonucleotide is hybridized to circular single-stranded M13-3.8Ac DNA isolated from virions which had been sedimented out of culture medium. The oligonucleotide:M13-3.8Ac hybrid is incubated with DNA ligase and the Klenow fragment of E. coli DNA polymerase I, cccDNA is enriched, and the mixture is transformed into JM103. The virions produced by transformants are isolated and used to infect cells at a low multiplicity of infection. RF DNA is isolated from a number of these infected colonies and characterized by restriction mapping. Clones derived from the mutant oligonucleotide-primed strand are identified by the presence of a novel NcoI site positioned at the 5'-end of the coding sequence. The mutated clones are further characterized to localize the NcoI site by digestion with ClaI and ClaI together with NcoI. The mutated M13-3.8Ac vector is labeled M13-3.8Aa.

4.3 Placement of a HindIII site at the 3'-end of the phaseolin gene

To conveniently introduce the insecticide gene into the phaseolin gene, two additional changes must be made to that phaseolin gene. The first change involves addition of HindIII site (5'...A*AGCTT...3') 5' to the polyadenylation site and near the 3'-end of the phaseolin gene. The other important change involves placing translational stop codons (e.g. TAA, TAG, or TGA, underlined below) in all three reading frames. When the oligonucleotide 5'AGGGTGCAITTTGAAGCTTGAATAAGTAAGAAGTAAAATGC3' (a) is compared with the 3'-end of the phaseolin gene coding sequence (b), it can be seen to have the desired properties as follows:

```

      HindIII
a) 5'AGGGTGCAITTTGA*AGCTTGAATAAGTAAGAAGTAAAATGC3'
b) ...AGGGTGCAITTTGT G1ACTGAATAAGTATGAAC1AAAATGC...
mismatches:      ↑ ↑↑↑↑      ↑
```

Note also that this 38-mer has only 6 mismatches, thus insuring good hybridization properties during priming.

The oligonucleotide 5'AGGGTGCAITTTGAAGCTTGAATAAGTAAGAAGTAAAATGC3', synthesized as described in Example 10.1, is hybridized to single-strand circular M13-3.8Aa DNA purified from virions isolated by centrifugation of culture medium. The oligonucleotide:M13-3.8Aa hybrid is incubated with DNA ligase and the Klenow fragment of *E. coli* DNA polymerase I, cccDNA is enriched, and the mixture is transformed into JM103. The virions produced by the transformants are isolated and used to infect cells at a low multiplicity of infection. RF DNA is isolated from a number of the infected colonies and characterized by restriction enzyme analysis. Clones derived from the mutant oligonucleotide-primed strand are identified by the presence of a HindIII site mapping at the 3'-end of the phaseolin gene, and the presence of mutant sequences at both ends of the structural gene is confirmed by sequencing. A vector containing the desired sequences is labeled M13-3.8Ab.

4.4 Inserting the insecticide gene

MBT3(Nco) RF DNA is digested with NcoI and HindIII and is mixed with and ligated to NcoI- and HindIII-digested M13-3.8Ab DNA. The mixture is transformed into K802 and plasmid DNA from kanamycin and/or tetracycline resistant transformants is isolated and characterized by restriction enzyme analysis. A plasmid having the insecticide structural gene inserted between the phaseolin promoter and polyadenylation site is labeled M13-PpBt, and a colony harboring it is chosen.

4.5 Moving the modified phaseolin gene into a shuttle vector

pKS111-K (Fink, supra) has the Tn5 kan gene from pKS-4 inserted between the HindIII sites of pKS111 T-DNA. M13-PpBt/RF DNA is digested with BamHI and mixed with and ligated to BamHI-linearized pKS111-K (Fink, supra). Plasmids from K802 transformants resistant to kanamycin and/or tetracycline are isolated and characterized by restriction mapping. A colony is selected which harbors a plasmid, labeled pKS111-PpBt, which contains the phaseolin promoter/insecticide structural gene/polyadenylation site combination which, together with a kan gene, is surrounded by octopine T-DNA.

4.6 Insertion into TIP plasmids, plant infection and regeneration

E. coli K802(pKS111-PpBt) is mated with A. tumefaciens as described in Example 9. The Agrobacterium strains chosen harbor TIP plasmids, based on pTil5955, containing mutations, such as those described in the Background, which facilitate regeneration. Homologous recombinants are selected as described in Example 9 and characterized by restriction mapping. The efficacy of the construction is quickly tested by infection of sunflower stems. The resulting galls are assayed by ELISA and Western blots as described in Example 7 and by bioassay as described in Example 8. As described in Example 6, the Agrobacterium strains are used to infect tobacco cells which are then regenerated. The resulting plants are used as breeding stock to be crossed with various commercial varieties for which insect resistance properties are desired. Fields of regenerated plants and their insecticidal protein-containing descendants are resistant

to infestation by larvae of insects such as tobacco hornworm by virtue of the toxic effect such larvae experience when eating tissue from such plants.

Example 5

Regeneration in this Example involves carrot tumors incited by Ri-based TIP plasmids and is effected essentially as described by Chilton M-D et al. (1982) Nature 295:432-434.

5.1 Infection with hairy root

Carrot disks are inoculated with about 10^9 bacteria in 0.1 ml of water. One to 1.5 cm segments of the ends of the roots obtained are cut off, placed on solid (1-1.5% agar) Monier medium lacking hormones (Tepfer DA and Tempe JC (1981) *Cr. hebdomadaire de l'Académie des Sciences*, Paris 295:153-156), and grown at 25°C to 27°C in the dark. Cultures uncontaminated by bacteria are transferred every 2 to 3 weeks and are subcultured in Monier medium lacking hormones and agar. Transformed roots can be recognized by their aberrant morphology and selected.

5.2 Regeneration of roots to plants

The cultured root tissue described in Example 5.1 is placed on solidified (0.8% agar) Monier medium supplemented with 0.36 μ M 2,4-D and 0.72 μ M kinetin. After 4 weeks, the resulting callus tissue is placed in liquid Monier medium lacking hormones. During incubation at 22 to 25°C on a shaker (150 rpm) for one month, the callus disassociates into a suspension culture from which embryos differentiate, which, when placed in Petri dishes containing Monier medium lacking hormone, develop into plantlets. These plantlets are grown in culture, and after "hardening" by exposure to atmospheres of progressively decreasing humidity, are transferred to soil in either a greenhouse or field plot.

5.3 Use of non-hairy root vectors

Ti-based vectors which do not have functional tnr genes are used instead of the Ri-based vectors as described by Hall TC et al., U.S. applications, ser. nos. 485,613 and 485,614. Construction of suitable mutants can be done by those skilled in the art, and is reviewed in the Background.

Example 6

Regeneration in this Example involves tobacco tumors incited by a Ti-based TIP plasmid and is effected essentially as described by Barton KA et al. (1983) Cell 32:1033-1043.

6.1 Infection with crown gall

Tobacco tissue is transformed using an approach utilizing inverted stem segments first described by Braun AC (1956) Canc. Res. 16:53-56. Stems are surface sterilized with a solution that was 7% commercial Chlorox and 80% ethanol, rinsed with sterile distilled water, cut into 1 cm segments, placed basal end up in Petri dishes containing agar-solidified MS medium (Murashige T and Skoog F (1962) Physiol. Plant. 15:473-497) lacking hormones. Inoculation is effected by puncturing the cut basal surface of the stem with a syringe needle and injecting bacteria. Stems are cultured at 25°C with 16 hours of light per day. The calli which develop are removed from the upper surface of the stem segments, are placed on solidified MS medium containing 0.2 mg/ml carbenicillin and lacking hormones, are transferred to fresh MS-carbenicillin medium three times at intervals of about a month, and are tested to ascertain whether the cultures had been ridden of bacteria. The axenic tissues are maintained on solidified MS media lacking supplements under the culture conditions (25°C; 16 hr.:8 hr. light:dark) described above.

6.2 Culture of transformed tissue

Clones are obtained from the transformed axenic tissues as described by Binns A and Meins F (1979) *Planta* 145:365-369. Calli are converted into suspensions of cells by culturing in liquid MS having 0.02 mg/l naphthalene acetic acid (NAA) at 25°C for 2 or 3 days while being shaken at 135 rpm, and filtering in turn through 543 and 213 µm stainless steel meshes. The passed filtrate is concentrated, plated in 5 ml of MS medium containing 0.5% melted agar, 2.0 mg/l NAA, 0.3 mg/l kinetin and 0.4 g/l Difco yeast extract at a density of about 8×10^3 cells/ml. Colonies reaching a diameter of about 1mm are picked by scalpel point, placed onto and grown on solidified MS medium having 2.0 mg/l NAA, 0.3 mg/l kinetin and about 10 µg/ml S-(2-aminoethyl)-L-cysteine (AEC). (A range of concentrations of AEC, between about 2 µg/ml and about 30 µg/ml, is tried as the exact concentration effective for selection will depend on the variety of tobacco used and the growth conditions to which the source plant and tissues derived from it are subjected.) AEC has been shown to be an agent capable of selecting tissue containing octopine synthase (Dahl GA and Tempe J (1983) *Theor. Appl. Genet.*, in press). Alternatively, the filtrate is plated at low density (several hundred cells per ml) on a filter paper overlaying a feeder layer of tobacco cells growing on the solidified MS/NAA/kinetin/yeast extract medium. When 1 mm colonies have formed the entire filter paper is transferred to a petri dish containing the solidified MS/NAA/kinetin/AEC medium. The resulting calli which do not show the effects of AEC toxicity are selected, split into pieces, and tested for other transformed phenotypes such as production of octopine and hormone independent growth.

6.3 Regeneration of plants

Transformed clones are placed onto solidified MS medium having 0.3 mg/l kinetin, and cultured as described in Example 6.1. The shoots which form are rooted by putting them on a solid (1.0% agar) medium containing 1/10 strength MS medium salts, 0.4 mg/l thiamine, lacking sucrose and hormones, and having a pH of 7.0. Rooted plantlets are grown in culture, hardened as described in Example 5.2, and are transferred to soil in either a greenhouse or field plot. Plants are screened for retention

of the transformed phenotype methods, well known to those skilled in the art, such as Southern, Northern and dot blots with appropriate probes, octopine assays, immunological (see Example 7) or biological (Example 8) assays for presence of crystal protein.

6.4 Vectors used

Constructions described by Hall TC et al., U.S. application ser. nos. 485,613 and 485,614 are suitable Ti-based vectors lacking functional tmr genes. The method described in Example 6.1 for infection of inverted stem segments is often useful for the establishment of TIP-transformed plant cell lines.

Example 7

Anti-insecticidal protein antibody was produced by methods well known to those skilled in the art of immunology. "Western" blots, to detect antigens after SDS-polyacrylamide gel electrophoresis, were done essentially as described by Legocki RP and Verma DPS (1981) Analyt. Biochem 111:385-392.

Micro-ELISA (enzyme-linked immuno-sorbant assay) assays are done using Immulon-2 type plates having 96 wells by the following steps:

7.1 Binding antibody to plates

On day 1, the wells are coated with 1:1000 dilution of antibody (rabbit anti-insecticidal protein IgG) in coating buffer. 200 μ l/well is incubated at 37°C for 2-4 hours. The plates are covered with Saran Wrap during this incubation. Afterwards the plates are rinsed three times with phosphate buffered saline-Tween (PBS-Tween) allowing a 5 minute waiting period between each rinse step. Then 1% bovine serum albumin (BSA) is added to rinse and, after addition to the well, left to sit for 20 minutes before discarding. Rinsing is repeated five times more with PBS-Tween.

7.2 Tissue homogenization

The tissue is sliced up into small pieces and then homogenized with a polytron using 1gm of tissue/ml phosphate buffered saline-Tween-2% polyvinyl pyrrolidone-40 (PBS-Tween-2% PVP-40). All samples are kept on ice before and after grinding and standard curves were obtained. One standard curve is done in tissue homogenates and one standard curve is also done in buffer to check the recovery of insecticidal protein from homogenized tissue or cells. Following centrifugation of the homogenized samples, 100µl of each sample is placed in a well and left overnight at 4°C. To avoid errors, duplicates of each sample are done. The plates are sealed during incubation.

7.3 Binding enzyme

After the overnight incubation, the antigen is discarded and the wells are washed five times with PBS-Tween allowing 5 minutes between each rinse.

A conjugate (rabbit anti-insecticidal protein IgG alkaline phosphatase-linked) is the diluted 1:3000 in PBS-Tween-2% PVP containing 0.2% BSA and 150 is added to each well; followed by incubation for 3-6 hours at 37°C. After the incubation, the conjugate is discarded and the wells are rinsed five times with PBS-Tween, allowing five minutes between each rinse as before.

7.4 Assay

Immediately before running the assay, a 5 mg tablet of p-nitrophenyl phosphate (obtained from Sigma and stored frozen in the dark) is added per 10 ml substrate and vortexed until the tablet is dissolved. 200 µl of the room temperature solution is quickly added to each well. The reaction is measured at various times, e.g. t=0, 10, 20, 40, 60, 90 and 120 minutes, using a Dynatech Micro-ELISA reader. When p-nitrophenyl phosphate, which is colorless, is hydrolysed by alkaline phosphatase to inorganic phosphate and p-nitrophenol, the latter compound gives the solution a yellow color, which can be spectrophotometrically read at 410 nm.

Example 8

Insects were obtained from commercial sources and kept essentially as described by Bell RA and Joachim FG (1976) Ann. Entomol. Soc. Amer. 69:365-373, or Yamamoto RT (1969) J. Econ. Entomol. 62:1427-1431. Bioassays for insecticidal protein were done by feeding extracts to larvae of Manduca sexta essentially as described by Schesser JH et al. (1977) Appl. Environ. Microbiol. 33:878-880.

Example 9

Triparental matings were generally accomplished as described below; other variations known to those skilled in the art are also acceptable. E. coli K802 (pRK290-based shuttle vector) was mated with E. coli (pRK2013) and a TIP plasmid harboring A. tumefaciens strain resistant to streptomycin. The pRK2013 transferred to the shuttle vector carrying strain and mobilized the shuttle vector for transfer to the Agrobacterium. Growth on a medium containing both streptomycin and the drug to which the shuttle vector is resistant, often either kanamycin or chloramphenicol, resulted in the selection of Agrobacterium cells containing shuttle vector sequences. A mating of these cells with E. coli (pPH1J1) resulted in the transfer of pPH1J1 to the Agrobacterium cells. pPH1J1 and pRK290-based shuttle vectors cannot coexist for long in the same cell. Growth on gentamycin, to which pPH1J1 carries a resistance gene, resulted in selection of cells having lost the pRK290 sequences. The only cells resistant to streptomycin, gentamycin, and kanamycin are those which have Ti plasmids that have undergone double-homologous recombination with the shuttle vector and now carry the desired construction. pRK290 and pRK2013 were disclosed by Ditta G et al. (1980) Proc. Natl. Acad. Sci. USA 77:7347-7357, and pPH1J1 by Hirsh PR (1978) Thesis, Univ. E. Anglia.

Example 10

This Example describes techniques for the synthesis and use of synthetic oligonucleotides. Other useful references can be found in the list of works cited in the section introductory to these Examples.

10.1 Oligonucleotide synthesis

The techniques for chemical synthesis of DNA fragments used in these Examples utilize a number of techniques well known to those skilled in the art of DNA synthesis. The modification of nucleosides is described by Schaller H et al. (1963) J. Amer. Chem. Soc. 85:3821-3827. The preparation of deoxynucleoside phosphoramidites is described by Beaucage SL and Caruthers MH (1981) Tetrahedron Lett. 22:1859. Preparation of solid phase resin is described by Adams SP et al. (1983) J. Amer. Chem. Soc. Hybridization procedures useful during the formation of double-stranded synthetic linkers are described by Rossi JJ et al. (1982) J. Biol. Chem. 257:9226-9229.

10.2 Use for oligonucleotides

Use of synthetic oligonucleotides to reconstruct a deleted segment of a gene has been exemplified by Hall et al., U.S. application ser. no. 485,614. Use of synthetic oligonucleotides to link otherwise incompatible restriction site sticky-ends has been exemplified by Hall et al., U.S. application ser. no. 485,614 and is well known to those skilled in the art of recombinant DNA manipulations.

10.3 Oligonucleotide-Directed Mutagenesis

General methods of directed mutagenesis have been reviewed recently by Shortle D et al. (1981) Ann. Rev. Genet. 15:265-294. Of special utility in manipulation of genes is oligonucleotide-directed site-specific mutagenesis, reviewed recently by Zoller MJ and Smith M (1983) Meth. Enzymol. 100:466-500 and Smith M and Gillam S (1981) in Genetic Engineering: Principles and Methods, Vol. 3, eds.: Setlow JK and Hollaender A, and Smith M (1982) Trends in Biochem. 7:440-442. This tech-

nique permits the change of one or more base pairs in a DNA sequence or the introduction of small insertions or deletions. Recent examples of use of oligonucleotide-directed mutagenesis include Zoller MJ and Smith M (1983) supra, Zoller MJ and Smith M (1982) *Nucleic Acids Res.* 10:6487-6500, Dalbadie-McFarland G et al. (1982) *Proc. Natl. Acad. Sci. USA* 79:6409-6413, Simons GFM et al. (1982) *Nucleic Acids Res.* 10:821-832, and Hutchison III CA et al. (1978) *J. Biol. Chem.* 253:6551-6560. Useful M13-based vectors (e.g. mW82344) have been reported by Barnes WM et al. (1983) *Meth. Enzymol.* 101:93-122, and Barnes WM and Bevan M (1983) *Nucleic Acids Res.* 11:349-368.

The sequence to be modified usually is moved into a single-stranded bacteriophage vector, here one derived from M13, by standard techniques well known to those in the art. The vector DNA is generally in the double-stranded replicative form (RF), as the single-stranded viral form cannot ordinarily be "cut and spliced" by restriction enzymes and ligases. After in vitro ligation of the fragment into the RF, transformation into a suitable host, and production single-stranded DNA (ssDNA) as part of the life cycle of the vector. ssDNA is isolated from phage particles and hybridized to an oligonucleotide having sufficient length and sequence homology to hybridize to the vector in the appropriate location. The oligonucleotide should have the sequence desired as an end product and otherwise differ in no way from the sequence to be changed. Once a hybrid is formed comprising a ssDNA circle base paired to the oligonucleotide carrying the mutant sequence, the oligonucleotide primes synthesis of a complementary strand of DNA by the Klenow fragment of E. coli DNA polymerase I, a polymerase lacking a 5'-to-3' exonuclease activity. The vector is optionally incubated with DNA ligase and the polymerase and ligase reactions may be done simultaneously. Preferentially covalently closed-circular double-stranded DNA (cccDNA) molecules can be selected before transformation by techniques which include alkaline sucrose gradient centrifugation, extraction with phenol under alkaline conditions, and incubation with S1 nuclease. The vector can now be transformed into an appropriate bacterial host cell. Virus particles from this initial infection are isolated and used to form plaques by infecting a lawn of bacteria. In cases where one is changing a restriction site, one

may readily screen RFs by restriction enzyme analysis. One may also screen by hybridization under carefully selected conditions using the synthetic mutant oligonucleotide primer as a probe, or by DNA sequencing. When a clone containing the desired change has been isolated, one may manipulate the now mutant DNA as desired using techniques well known to those skilled in the art.

Example 11

This Example teaches isolation of a clone having the 3'-end of the insecticide gene carried by p123/58-10, and the reconstruction of a full-length HD-73 crystal protein gene.

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end

11.1 Cloning of an insecticide gene 3'-end

Immunodetection of electrophoretically separated peptides on protein blots and DNA sequencing showed that p123/58-10 and p123/58-3 each contained a partial protoxin gene. To reconstruct a complete protoxin gene, flanking DNA restriction sites were identified by Southern blots of restriction digests, a well known technique, and overlapping clones were selected from a PstI library made from 50 MD plasmid-enriched DNA as follows. 50 MD plasmid DNA, enriched by sucrose gradient centrifugation as above, was digested to completion with PstI, mixed with and ligated to PstI-linearized pBR322, and transformed into HB101. Tetracycline-resistant transformants were screened essentially as described by W. D. Benton and R. W. Davis (1977) Science 196:180-182, using a probe nick-translated from the 6.7 kbp HindIII insert of p123/58-10. Plasmid DNAs isolated from strains which bound the probe were characterized by restriction enzyme analysis. A strain chosen for further work harbored pBt73-161, which contains the 3'-end of a crystal protein gene.

11.2 Construction of a full-length insecticide gene

The 5'- and 3'-ends of the protoxin gene were fused together at the unique HindIII site to form a complete protoxin gene. p123/58-10 DNA was digested with BamHI, ligated to itself, and transformed into HB101. Plasmid DNAs from ampicillin-resistant transformants were characterized by

restriction enzyme analysis and a strain was identified that harbored a plasmid, designated pBt73-10(Bam), having single BamHI and HindIII sites due to deletion of a small HindIII site-bearing BamHI fragment. A 5 kbp HindIII fragment of pBt73-10i, isolated by agarose gel electrophoresis, was mixed with and ligated to HindIII-digested dephosphorylated (by bacterial alkaline phosphatase) pBt73-10(Bam) DNA. After the ligation mixture was transformed into HB101, plasmid DNA isolated from ampicillin-resistant tetracycline-sensitive transformants was characterized by restriction enzyme analysis. A transformant was identified that harbored a plasmid, designated pBt73-10, carrying a complete protoxin gene. E. coli HB101 (pBt73-10) is on deposit at the Northern Regional Research Center, 1815 N. University St., Peoria, Illinois 61604 USA, as NRRL B-15759.

Example 12

This Example teaches the insertion of the full-length Bacillus thuringiensis insecticide gene between a T-DNA gene promoter and a polyadenylation (poly(A) addition) signal, the transfer of the insecticide gene to tobacco via a sub-Ti plasmid, the regeneration of plants containing the gene, and the expression of this gene under control of the T-DNA promoter in plant cells.

12.1 Preparation of pH400, a sub-Ti plasmid

pH4-1 DNA can be isolated from E. coli K802 (pH4-1), which has been deposited as NRRL B-18009. pH4-1 is a sub-Ti plasmid based on pSUP106 (Priefer UB et al. (1985) J. Bacteriol. 163:324-330; E. coli CSH52 (pSUP106) is deposited as NRRL B-15486). Inserted into pSUP106's HindIII site is pTi15955 DNA having border A (the left border of T_L) a plant-expressible selectable marker, the 5'-end of tml, a complete ocs gene, and border B (the right border of T_L). The plant-expressible selectable marker is sutured to the adjoining pTi15955 sequences at BglII sites made up of a Tn5 neomycin phosphotransferase II (NPT2) structural gene capable of detoxifying kanamycin and related antibiotics (a kun gene). The NPT2 structural gene is transcribed in plant cells under control of a cauliflower mosaic virus (CaMV; for a review of CaMV see Hohn T et al. Curr. Top. Microbiol. Immunol. 96:193-236) 19S transcript promoter and a

pTi15955 ORF26 (as defined by Barker *et al.* (1983) *Plant Mol. Biol.* 2:335-350) polyadenylation site. The NPT2 structural gene is oriented so that it is transcribed in the same direction as the *ocs* gene. In other words, the T-DNA of pH4-1 is essentially identical to T_L-DNA of pTi15955 except for substitution of a CaMV 19S promoter/NPT2 structural gene/ORF26 polyadenylation site combination for T-DNA between the position 1617 *Bgl*II site and the position 11207 *Sma*I site, T-DNA positions being defined by Barker *et al.*, *supra*.

pH4-1 has two *Bgl*II sites, both of which flank the *kan* selectable marker. One of the *Bgl*II sites was removed, thereby leaving a unique *Bgl*II site useful for insertion of extraneous coding DNA. pH4-1 DNA was linearized by being partially digested with *Bgl*II and full-length, linear DNA was electrophoretically isolated. The *Bgl*II sticky-ends were then removed by incubation with the Klenow fragment of *E. coli* DNA polymerase I. The resulting blunt-ended DNA was ligated to itself and transformed into *E. coli*. Plasmid DNAs isolated from transformants resistant to chloramphenicol were screened by restriction analysis and a colony was identified which harbored a plasmid designated pH400. pH400 is identical to pH4-1 except for the absence of the *Bgl*II site between the *kan* gene and the ORF1 promoter, the unique pH400 *Bgl*II site being located between the *kan* gene and the *ocs* gene. The single *Bgl*II site is a convenient place to insert DNA, in particular if that DNA has compatible 5'GATC...3' sticky-ends resulting from the action of *Bgl*II, *Bcl*I, or *Bam*HI.

12.2 Preparation of pDOB513

pDOB512, carrying cauliflower mosaic virus (CaMV) transcription controlling sequences (obtained from Dr. Ken Richards, Centre National de la Recherche Scientifique, Institute de Biologie Moléculaire et Cellulaire, 15, rue Descartes, F-67084 Strasbourg, France) was constructed as follows: A *Hind*III fragment carrying the CaMV 19S RNA promoter region (CaMV nucleotides 5376-5851) was inserted into pBR322 and was trimmed back to within one base pair of the 19S transcript cap site. An adapter molecule having both a *Sma*I site and a *Bam*HI (the structure being 5'CCCGGGGATCCGG3':5'CCGGA1CCCGGG3', see below) was then ligated to the trimmed DNA. A *Hinc*II fragment carrying the CaMV 19S transcript termina-

tor (CaMV nucleotides 7018-7794) to which BamHI linkers had been added was then inserted behind the 19S promoter, the promoter and terminator being separated by the SmaI/BamHI linker. The resulting plasmid is designated p00B412. p00B412 DNA was digested with BglII and SalI, filled in by incubation with the Klenow fragment of E. coli DNA polymerase I, and religated, thereby deleting DNA, which includes BamHI and HindIII sites, between the CaMV position 7644 BglII site and the pBR322 position 650 SalI site and regenerating a BglII site. The resultant plasmid was designated p00B512.

The sticky-ends of HindIII-linearized p00B512 DNA were converted to blunt-ends. The blunt-ended p00B512 DNA was mixed with and ligated to commercially available BglII linkers. The ligation mix was transformed into E. coli K802 and an ampicillin-resistant transformant was isolated which harbored a plasmid, designated p00B513 (Fig. 3). p00B513 has CaMV 19S transcription controlling sequences on a BglII fragment. SmaI and BamHI sites are found between the DNA segments having the promoter and the polyadenylation site in both p00B412, p00B512, and p00B513, thereby providing a convenient location for insertion of foreign DNA that is to be a template for a transcript. The sequence of this suture is as follows:

SmaI BamHI
5'...GAGAAAATCAGCCCGGGGATCCGGAACA...3'

The first underlined base, an A, corresponds to the first base of the 19S mRNA, transcription being towards the right, and the second and third underlined bases, respectively a G and an A, are derived from CaMV sequences at positions 5765 and 7018, respectively, as defined by Hohn et al., supra.

12.3 Preparation of T-DNA ORF24 transcription controlling sequences

pKS-prol was prepared essentially as described in Example 2.2. pKS111 is a recombinant plasmid having T15955 T-DNA which spans the sequence between EcoRI sites at positions 16,202 and 21,631 (as defined by Barker et al., supra) inserted into the EcoRI site of pRK290 (Ditta G et al. (1980) Proc. Natl. Acad. Sci. USA 77:7347-7351). E. coli K802

(Wood WB (1966) J. Mol. Biol. 16:118) methylates DNA at many, but not all, ClaI sites, thereby specifically protecting certain sites from the action of ClaI. The only susceptible ClaI sites of pKS111 are the sites at T-DNA positions 18,892 and 20,128. These sites define a DNA fragment which includes the ORF24 (as defined by Barker et al., supra) structural gene but does not include the ORF24 promote or polyadenylation site. ORF24 is associated with mannopine synthesis. Plasmid DNA isolated from E. coli K802 (pKS111) was digested with ClaI, ligated to itself, and transformed into K802. A tetracycline-resistant transformant was identified by restriction analysis which harbored a plasmid, designated pKS-proI (or alternatively, pTR-proI), identical to pKS111 except for deletion of a fragment which spanned the ClaI sites at positions 18,892 and 20,128.

pTR-proI DNA was isolated and cleaved at its single ClaI site (the suture between the sites at positions 18,892 and 20,128) by incubation with ClaI. After the ClaI sticky-ends were then removed by incubation with the Klenow fragment of E. coli DNA polymerase I, the blunt-ended DNA was mixed with and ligated to commercially available BamHI linkers having the self-complementary sequence 5'CGGATCCG3'. After digestion of the BamHI-linked DNA with BamHI, religation to itself, and transformation of the resulting DNA into K802, plasmid DNAs were isolated from transformants resistant to tetracycline were screened by restriction mapping. A colony was identified which harbored a plasmid, designated pTR-proI(Bam), lacking a ClaI site at the T-DNA positions 18,892/20,128 suture but, instead, having a BamHI site at the former location of that ClaI site.

pBR322 has a BamHI site which will prove inconvenient for later manipulations. Therefore, the T-DNA of the ClaI-deleted pKS111-derivative was transferred to a pBR325 derivative which was lacking a BamHI site. pBR325 DNA, isolated from E. coli GM33 (pBR325), was digested with both BamHI and BclI, was ligated to itself and was transformed into E. coli. GM33 (Marinus MG and Morris RH (1974) J. Mol. Biol. 85:309-322) does not methylate adenine residues of DNA. Plasmid DNA isolated from transformants sensitive to tetracycline and resistant to chloramphenicol and ampicillin were characterized by restriction mapping and a colony was identified which harbored a plasmid, designated pBR325aBB, which could not be cleaved with either BamHI or BclI. pTR-proI DNA was digested with

EcoRI and was mixed with and ligated to EcoRI-linearized, dephosphorylated pBR325aBB DNA. After transformation into E. coli, plasmid DNAs isolated from transformants resistant to ampicillin and sensitive to both chloramphenicol and tetracycline were characterized by restriction mapping. A colony was identified which harbored a plasmid, designated p403B, having the ClaI-deleted T-DNA with BamHI linkers in the former ClaI site, recombined with the pBR325aBB vector.

pKS4.2 DNA (Example 2.3) was digested with ClaI and a fragment carrying the kan gene was electrophoretically isolated. This fragment was mixed with and ligated to ClaI-linearized pBR322 and transformed into E. coli. Plasmid DNAs isolated from transformants resistant to ampicillin and kanamycin were screened by restriction analysis and a colony was identified which harbored a plasmid designated pKS4.3. The pKS4.3 kan gene was oriented with its 5'-end and 3'-end respectively proximal to the pBR322 EcoRI and BamHI sites. In this orientation, the kan gene may be removed on a HindIII fragment.

BamHI-linearized pBR322 DNA was mixed with and ligated to BglII-digested bacteriophage lambda DNA. Transformation into E. coli cells was followed by selection for resistance to ampicillin and screening for sensitivity to tetracycline. Plasmid DNAs were then isolated and screened by restriction analysis and a colony was identified which harbored a plasmid, designated pBR322Bam^r, which could not be cleaved with BamHI.

pKS4.3 DNA was digested with HindIII and the kan gene-bearing fragment was electrophoretically isolated. The kan fragment was then mixed with and ligated to HindIII-linearized pBR322Bam^r DNA. The ligation mixture was then transformed into E. coli. Plasmid DNAs isolated from transformants resistant to both kanamycin and ampicillin were screened by restriction analysis and a colony was identified which harbored a plasmid, designated p11-83c, having the kan fragment inserted into pBR322 Bam^r's HindIII site. In p11-83c the kan gene was oriented so that the kan gene was transcribed in the same direction as the ampicillin resistance (amp) gene.

pTR-proI(Ban) was digested with EcoRV and an approximately 2.26 kbp fragment bearing the ORF24 ("1.6" transcript) promoter and polyadenylation site, but lacking the ORF24 structural gene, was electrophoretically isolated. The EcoRV sticky-ends were then converted to blunt-ends by the action of the Klenow fragment of E. coli DNA polymerase I. The blunt-ended fragment was then mixed with and ligated to commercially available BglII linkers. After trimming off excess linkers by digestion with BglII, the fragment was mixed with and ligated to BglII-linearize p11-83c DNA. The ligation mixture was then transformed into E. coli. Plasmid DNAs isolated from ampicillin-resistant, kanamycin-sensitive transformants were restriction mapped and a colony was identified which harbored a plasmid, designated pCG116, having a BglII fragment bearing the ORF24 promoter and polyadenylation site inserted into Tn5 DNA between the kan promoter and kan structural gene.

pDOB513 DNA was digested with BglII, religated to itself, and transformed into K802. Colonies which harbored a plasmid, designated pDOB514, deleted for CamV transcription controlling sequences were identified by restriction mapping of the harbored plasmids.

pCG116 DNA, which had, on a 2.26 kbp BglII fragment, a BamHI site between the ORF24 promoter, and transcript terminator, was then digested with BglII, which cleaves at sites which in T-DNA correspond to positions 18,027 and 21,522 EcoRI sites. The restriction digested DNA was mixed with and ligated to BglII-linearized pDOB514 DNA. Plasmid DNAs of ampicillin-resistant transformants were characterized by restriction mapping, and a colony was identified which harbored a plasmid, designated pMAN514, having a 2.29 kbp BglII fragment carrying an ORF24 promoter and polyadenylation site separated by a BamHI site.

12.4 Preparation of the insecticide gene

After pBt73-16 DNA was digested with NdeI, resulting NdeI sticky-ends were filled in by incubating the mixture with T4 DNA polymerase and all four dNTPs. The blunt-ended Bacillus DNA was mixed with and ligated to double-stranded, SmaI-linearized, M13mp19 RF DNA (J. Norrander *et al.* (1983) Gene 26:101-106). The ligation mixture was transformed into E. coli JM105. DNAs isolated from plaques that were "clear" when plated

on indicator plates were characterized by restriction analysis and a plaque was identified which harbored a vector, designated 1.6.4, having a 3.6 kbp Bacillus DNA oriented such that single-stranded form was complementary to crystal protein mRNA (i.e. the phage carried the antisense strand).

A BamHI site was introduced into the Bacillus DNA immediately 5'- to the crystal protein translational start site essentially as described in Example 10. Sequences of the unmutated Bacillus DNA and the oligonucleotide primer are as follows:

Bacillus: 5'...GAGATGGAGG TAA CTTATGGATAAC...3'
 Primer: 5'GAGATGGAGGATCCTTATGGATAAC3'
 BamHI MetTyrHis...

Putative mutant DNAs were screened for hybridization under stringent conditions to a ³²P-label derivative of the oligonucleotide primer and for the presence of a novel BamHI site. A plaque was identified harboring a vector designated 1.6.4B-3.8.3, having a BamHI site immediately 5'- to the translational start. Fidelity of the in vitro mutation was confirmed by DNA sequencing. The insecticidal protein structural gene may be removed from double-stranded 1.6.4B-3.8.3 RF DNA on a 3.75 kbp BamHI fragment resulting from cleavage at the novel site in the Bacillus sequence and a BamHI site which is 3'-to the structural gene in the M13mp19 polylinker.

12.5 Combination of an insecticidal protein structural gene with ORF24 transcription controlling sequences

Double-stranded 1.6.4B-3.8.3 RF DNA and pCG116 DNA, both having been digested with BamHI, were mixed with each other and ligated together. After the ligation mixture was transformed into E. coli, plasmid DNAs isolated from ampicillin-resistant transformants were characterized by restriction analysis. A colony was identified which harbored a plasmid, designated pCJ101, having a full-length insecticidal protein structural gene placed between the ORF24 promoter and ORF24 polyadenylation site in an orientation such that the mRNA transcribed under control of the ORF24 promoter encodes the insecticidal protein.

12.6 Insertion of a plant-expressible insecticide gene into a sub-Ti plasmid

pCJ161 and pH400 DNAs, having been digested with BglII, were mixed with each other and ligated together. After the ligation mixture was transformed into E. coli, plasmid DNAs isolated from ampicillin-resistant transformants were characterized by restriction analysis. A colony was identified which harbored a plasmid, designated pH450, having the ORF24 promoter/crystal protein structural gene/ORF24 polyadenylation site combination inserted into the pH400 BglII site, which is located between the ocs gene and the plant-expressible kan gene. The plant-expressible crystal protein gene of pH450 is oriented so that it is transcribed in the same direction, i.e. off of the same DNA stand, as both the ocs gene and the kan gene.

12.7 Plant transformation

pH450 was transferred into A. tumefaciens LBA4404 (Ooms G et al. (1981) Gene 14:33-50), a vir gene-bearing, micro-Ti-mobilizing strain, by the triparental mating technique. Tobacco leaf tissues were inoculated using a modification of the method of R.B. Horsch et al. (1985) Science 227:1229-1231. Leaf segments were dipped in the inoculating suspension of bacteria for 1-3 min. The inoculating bacterial suspensions had titres of 10^7 - 10^8 ml⁻¹; the exact concentration was found to be unimportant. The medium was a tobacco regenerating medium having MS salts, a mixture well known to the art, supplemented with 0.1 mg/l p-chlorophenoxyacetic acid (pCPA), 7.5 mg/l 6-(2,2-dimethylallylamino)-purine (2iP), 300 mg/l kanamycin, and 125 mg/l cloxacillin, mefoxin and 500 mg/l carbenicillin. Generally 75-80% of the shoots were able to root in a kanamycin-containing medium and all were transformed. Some kanamycin-resistant shoots did not make measurable quantities of octopine.

12.8 Expression in plant tissues: immunoassays

Micro-ELISA double antibody sandwich assays were performed using a modification of the method of Example 7.

Antibodies were prepared by well-known methods. Coating antibody was partially purified mouse polyclonal antisera which was raised to B. thuringiensis HD73 crystal protein. The mouse antisera was partially purified by precipitation with 50% (w/v) saturated ammonium sulfate at pH 7.4. The milky precipitate that forms during 30 min. on ice was pelleted by centrifugation (20 min., 10,000 rpm). The pellet was then resuspended in phosphate buffered saline (PBS, pH 7.4) containing 137mM NaCl, 1.5mM KH_2PO_4 , 2.7mM KCl and 8mM NaH_2PO_4 . The resulting coating antibody sample was then dialyzed overnight in PBS at 4°C.

Primary antibody was partially purified rabbit polyclonal antisera raised to B. thuringiensis HD73 crystal protein. This antisera was cleared of antibodies that react with tobacco leaf protein. Tobacco leaf protein was bound to CNBr activated Sepharose 4B (Pharmacia, Upsalla, Sweden) (10mg protein/ml gel). Rabbit serum was diluted 1:4 (v/v) with PBS and added to the swollen Sepharose gel to which the leaf protein was bound. About 2-3 ml of gel was used for 4 ml of diluted serum. The serum-gel mixture was incubated with gentle shaking for about 1 hr. at room temperature. Antibodies that react with leaf protein remain bound to the gel. The liquid containing unbound antibodies was removed from the gel matrix by aspiration. The antibody solution was then partially purified by precipitation with 50% saturated ammonium sulfate followed by overnight dialysis against PBS at 4°C.

Secondary antibody was commercial goat anti-rabbit IgG (H+L) labelled with alkaline phosphatase (Kirkegaard and Perry, Gaithersburg, Maryland). Sterile, distilled water (1ml) was added to 0.5mg of lyophilized pellet. The wells of disposable flexible 96-well/flat-bottom microtitration plates (Dynatech, Alexandria, Virginia) were coated (100µl/well) with coating antibody (1:2000 dilution of coating antibody in coating buffer). Coating buffer contained 5mM Na_2HCO_3 and 35mM NaHCO_3 at pH 9.6. After addition of the coating antibody solution, the plates

were covered and stored at 4°C for at least 12 hours. The contents of the wells were then discarded and the entire plate was rinsed about 3-4 times with a wash buffer of PBS containing 0.05% polyoxyethylenesorbitan monolaurate (Tween-20). Plates were then filled with wash buffer and placed on a shaker for 5 min. followed by a final rinse. The plate was then drained for 30 seconds. It is important that at no point in the assay that the wells dry for a prolonged period. In between assay steps the wells were filled with wash buffer.

The plates were next treated with blocking solution to prevent non-specific binding of proteins. Plate wells were filled with blocking solution containing 1% (w/v) BSA (bovine serum albumin); 1% (w/v) Casein; acid hydrolysate, type 1; and 0.05% (v/v) polyoxyethylenesorbitan monolaurate (Tween-20) and allowed to stand at room temperature for 30 min. to 1 hr.

Tobacco plant tissue samples were prepared from leaf material. Generally, 1 or 2 leaves from the apex of the plant were combined with 4 or 5 leaf disks (7mm) from the basal and marginal regions of the next youngest leaves. Tissue samples of about 100mg were used. After weighing, tissue samples were quick frozen in liquid nitrogen and ground in a tissue grinder. The sample was allowed to warm to room temperature during grinding. After the tissue is completely ground, PBS was added to make a solution containing 0.3mg fresh weight/ μ l (for a 100mg sample, 500 μ l of PBS was added). Callus tissue samples were prepared in the same manner, except that PBS was added to ground tissue to give a solution of 0.5mg fresh weight/ μ l. Samples were then mixed thoroughly and allowed to stand at 4°C (about 20 min.) for tissue to settle out. Samples (10-15 μ l) of the supernatant were taken for total protein assays. Samples were then diluted with an equal volume of a 2x concentration of sample buffer. Sample buffer (1x) contained PBS (pH 7.4) with 2% (v/v) polyethylene glycol, 1% (v/v) polyvinyl pyrrolidone, 0.1% (w/v) BSA, 0.1% (w/v) Casein, acid hydrolysate, 10 μ g/ml Aprutinin (Sigma), a protease inhibitor, 0.05% (v/v) polyoxyethylenesorbitan monolaurate and 0.006% (v/v) β -mercaptoethanol. Control tissue (non-transformed) samples were prepared in a similar manner. Samples were spun in a microfuge for 1 min. before loading into wells of the prepared microtitre plates. Sample supernatant

was loaded (100 μ l/well) in replicates (3 or 4). Plates containing samples are incubated for 2-3 hrs. at room temperature. For quantitation, standards of B. thuringiensis crystal protein can be included.

After incubation the contents of the wells were removed and the plate was rinsed carefully with wash buffer (4-5 times). The plate was filled with wash buffer and agitated for 5 min. Care should be taken in this step to avoid cross-contamination of samples, controls and standards.

A 1:2000 dilution of the primary antibody in antibody buffer was then loaded into the wells (100 μ l/well). Plates were covered and stored at 4°C overnight (about 12 hours). Antibody buffer contained PBS (pH 7.4) with 1% (v/v) goat serum and 0.05% polyoxyethylenesorbitan monolaurate. Plates were then washed (3-4 times) with wash buffer (as above). Plates were then blotted dry.

A 1:2500 dilution of labelled secondary antibody (goat anti-rabbit IgG (H+L)-alkalinephosphatase-labelled) was then loaded into wells (100 μ l/well). Plates were incubated for 1-2 hours at room temperature and thoroughly washed. Plates were washed 4-5 times with wash buffer. Plates were filled with wash buffer and placed on a shaker for at least 10 minutes. Plates were rinsed again and blotted dry.

An NADH substrate was then employed to detect bound labelled antibody. NADP (β -Nicotinamide Adenine Dinucleotide Phosphate) 0.25mg/ml in 0.50mM MgCl₂, 0.05mM diethanolamine (pH 9.5) was loaded into the assay wells (100 μ l/well). Plates were covered and stored at 4°C for about 1 1/2 hours.

Reagent solution was prepared by dissolving p-Iodonitrotetrazolium violet (0.31mg/ml) in 0.05 M sodium phosphate buffer (pH 7.2) containing 1.9% (v/v) ethanol. To this solution Diaphorase "NADH/dye oxidoreductase" (Boehringer Mannheim, Mannheim, West Germany) to 0.035 mg/ml and alcohol dehydrogenase (baker's yeast) to 0.035 mg/ml were added. The reagent solution was incubated at 4°C for 45 minutes after which it was added to the NADP solution in each assay well (150 μ l/well).

The color reaction (red) should begin immediately. Optical density measurements were read at 492nm.

Adjusted ELISA values of crystal protein in leaf tissue, expressed as absorbance at 492 nm, are reported in Table 7. The values were adjusted by subtracting from each measurement the value obtained for a Nicotiana tabacum var. "Xanthi" control. Many plants were observed to contain measurable amounts of antigen which bound anti-crystal protein antibody.

12.9 Expression in plant tissues: bioassays

Tobacco tissue transformed by pH450 was cloned and maintained in a growth chamber. For bioassay, leaves were cut off with a razor blade and placed in a 9 cm petri dish lined with moist filter paper. Newly hatched tobacco hornworm larvae (Maniaca sexta) were put on the leaves. Mortality was recorded.

In trial 1, three clones from each type were sampled with five larvae per dish (Table 8). The data can be summarized as follows:

<u>Clone</u>	<u>Total</u>	<u>Dead</u>	<u>% Dead</u>
100	14	5	36
103	15	1	7
106	15	0	0
109	15	5	33
111	15	3	20

Trial 2 was essentially done as described for trial 1, but here two larvae were placed in each dish. The data are reported in Table 9 and can be summarized as follows:

<u>Clone</u>	<u>Total</u>	<u>Dead</u>	<u>% Dead</u>
100	14	9	64
103	18	4	22
106	24	12	50
109	26	7	27
111	18	8	44

Leaves that had killed both larvae by day 2 were then reinfested with three newly hatched larvae. While the numbers were not large enough to make comparisons between treatments, it was clear that these leaves are still toxic. For example, three dishes from clone 100 received a total of 9 larvae, 6 of which were dead 2 days later.

As protein concentration is highest in small leaves, expression might vary with the size of the leaf, i.e., with the degree of expansion of the cells. Therefore in trial 3 leaves were measured before bioassay. For this test, only plants that had not been sampled before of clones 100 and 103 were sampled. The data were ranked from smallest to largest leaves, but the sizes for the two clones were not aligned. Different numbers of dishes were set up for each plant because the plant of clone 100 had fewer leaves. Three larvae were placed in each dish. Number of dead larvae per dish are reported below.

<u>Rank</u>	<u>Clone 100</u>	<u>Clone 103</u>
1	1	0
2	3	0
3	1	0
4	1	0
5	1	0
6	3	0
7	3	0
8		0
9		2
10		1

It is clear that both small and large leaves are able to kill the larvae. Furthermore, the difference between different plant clones, i.e. 100 and 103, is apparent: 62% vs. 10% mortality, at day 2.

In trial 4, randomly assigned labels were used to avoid biases. Five petri dishes from each clone were prepared with three larvae per dish. In addition, a Xanthi plant that originated in tissue culture was used as a control. The data can be summarized as follows:

<u>Summary</u>	<u>Dead/Total</u>	<u>% Mortality</u>
control	2/15	13
100	11/15	73
103	8/15	53
106	2/15	13
109	1/15	7
111	3/15	20

Clone 100 gave the consistently high mortality. Clone 103 also gave high

mortality explained by the subsequently discovered fact that "clone" 103 was not in fact a clone; the original transformed plant has now been shown to have been a chimeric plant. It has not proven to be particularly insecticidal in any other of the five trials.

After the trial 4 leaves had been fed on for two days, ELISAs were performed on some of the lethal and nonlethal leaves. No crystal protein antigen was detected because of the low levels of protein in these leaves.

In trial 5, four plants were propagated from clone 100 and five from clone 103. Five leaves were sampled from each of these plants. Each leaf was placed in an individual petri dish with three newly hatched larvae.

The data can be summarized as follows:

<u>Summary</u>	<u>Dead/Total</u>	<u>% Mortality</u>
100	8/14	57
100	3/14	21
100	10/15	67
100	5/14	36
103	3/15	20
103	1/15	7
103	2/15	13
103	2/15	13

Thus, clone 100 is consistently more toxic than clone 103.

Southern blot analysis of DNA isolated from presuptive transformants showed that cloned plant tissue having insecticidal activity in bioassays and containing crystal protein antigen generally had pH450's T-DNA. Northern blot analysis of RNA generally demonstrated the presence of mRNA having crystal protein sequences. These mRNA molecules were not the expected size of about 3.8 kbp, but were about 1.7 kbp in size. This was sufficient to encode the toxic portion of the crystal protein. As predicted from the fact that crystal protein levels in leaves as estimated by ELISA were below the limit of detection on western blots, western blot analysis of proteins extracted from transformed tissues did not reveal any antigens that cross-reacted with anti-crystal protein antibodies. Polypeptides that bind anti-crystal protein antibodies have been detected by western blot analysis of both transformed callus and young shoot tissues.

TABLE 1

Insects susceptible to *B. thuringiensis* insecticidal protein

COLEOPTERA

Popillia japonica (Japanese beetle)

Sitophilus granarius (granary weevil)

DIPTERA

Aedes aegypti (yellow-fever mosquito)

A. atlanticus

A. cantans

A. capsus

A. cinereus

A. communis

A. detritus

A. dorsalis

A. dupreei

A. melanimon

A. nigromaculis (pasture mosquito)

A. punctor

A. sierrensis (western treehole mosquito)

A. sollicitans (brown salt marsh mosquito)

Aedes sp.

A. taeniorhynchus (black salt marsh mosquito)

A. tarsalis

A. tormentor

A. triseriatus

A. vexans (inland floodwater mosquito)

Anopheles crucians

A. freeborni

A. quadrimaculatus (common malaria mosquito)

A. sergentii

A. stephensi

Anopheles sp.

Chironomus plumosus (*Chironomus*: midges, biting)

Chironomus sp.
 C. tummi
 Culex erraticus
 C. inornata
 C. nigripalus
 C. peus
 C. pipiens (northern house mosquito)
 C. quinquefasciatus (C. pipiens fatigans) (southern house mosquito)
 C. restuans
 Culex sp.
 C. tritaeniorhynchus
 C. tarsalis (western encephalitis mosquito)
 C. territans
 C. univittatus
 Culiseta incidens (Culiseta: mosquitos)
 C. inornata
 Diamessa sp.
 Dixa sp. (Dixa: midges)
 Eusimulium (Simulium) latipes (Eusimulium: gnats)
 Goeldichironomus holoprasinus
 Haematobia irritans (horn fly)
 Hippelates collusor
 Odagmia ornata
 Pales pavidia
 Polpomyia sp. (Polpomyia: midges, biting)
 Polypedilum sp. (Polypedilum: midges)
 Psorophora ciliata
 P. columiae (confinnis) (Florida Glades mosquito, dark rice field mosquito)
 P. ferox
 Simulium alcocki (Simulium: black flies)
 S. argus
 S. cervicornutum
 S. damnosum
 S. jenningsi
 S. piperi

S. tescorum
S. tuberosum
S. unicornutum
S. venustum
S. verecundum
S. vittatum
Uranotaenia inguiculata
U. lowii
Wyeomyia mitchellii (Wyeomyia: mosquitos)
W. vanduzeei

HYMENOPTERA

Athalia rosae (as colibri)
Nematus (Pteronidea) ribesii (imported currantworm)
Neodiprion banksianae (jack-pine sawfly)
Priophorus tristis
Pristiphora erichsonii (larch sawfly)

LEPIDOPTERA

Achaea janata
Achroia grisella (lesser wax moth)
Achyra rantalis
Acleris variana (black-headed budworm)
Acrobasis sp.
Acrolepia alliella
Acrolepiopsis (Acrolepia) assectella
Adoxophyes orana (apple leaf roller)
Aegeria (Sanninoidea) exitiosa (peach tree borer)
Aglais urticae
Agriopsis (Erannis) aurantiaria (Erannis: loopers)
A. (E.) leucophaearia
A. marginaria
Agrotis ipsilon (as ypsilon) (black cutworm)
A. segetum
Alabama argillacea (cotton leafworm)
Alsophila aescularia

A. pometaria (fall cankerworm)
Amorbia essigana
Anadevidia (*Plusia*) *peponis*
Anisota senatoria (orange-striped oakworm)
Anomis flava
A. (*Cosmophila*) *sabulifera*
Antheraea pernyi
Anticarsia gemmatilis (velvetbean caterpillar)
Apocheima (*Biston*) *hispidaria*
A. pilosaria (*pedaria*)
Aporia crataegi (black-veined whitemoth)
Archips argyrospilus (fruit-tree leaf roller)
A. cerasivoranus (ugly-nest caterpillar)
A. crataegana
A. podana
A. (*Cacoecia*) *rosana*
A. xylosteana
Arctia caja
Argyrotaenia mariana (gray-banded leaf roller)
A. velutinana (red-banded leaf roller)
Ascia (*Pieris*) *monuste orseis*
Ascotis selenaria
Atteva aurea (alioanthus webworm)
Autographa californica (alfalfa looper)
A. (*Plusia*) *gamma*
A. nigrisigna
Autoplusia egea (bean leaf skeletonizer)
Azochis gripusalis
Bissetia steniella
Bombyx mori (silkworm)
Brachionycha sphinx
Bucculatrix thurberiella (cotton leaf perforator)
Bupalus piniarius (*Bupalus*: looper)
Cacoecimorpha pronubana
Cactoblastis cactorum
Caloptilia (*Gracillaria*) *invariabilis*

C. (G) syringella (lilac leaf miner)
C. (G.) theivora
Canephora asiatica
Carposina niponensis
Ceramidia sp.
Cerapteryx graminis
Chilo auricilius
C. sacchariphagus indicus
C. suppressalis (rice stem borer)
Choristoneura fumiferana (spruce budworm)
C. murinana (fir-shoot roller)
Chrysodeixis (Plusia) chalcites
Clepsis spectrana
Cnaphalocrocis medinalis
Coleotechnites (Recurvaria) milleri (lodgepole needle miner)
C. nanella
Colias eurytheme (alfalfa caterpillar)
C. lesbia
Colotois pennaria
Crambus bonifatellus (fawn-colored lawn moth, sod webworm)
C. sperryellus
Crambus spp.
Cryptoblabes gnidiella
Cydia funebrana
C. (Grapholitha) molesta (oriental fruit moth)
C. (Laspeyresta) pomonella (codling moth)
Datana integerrima (walnut caterpillar)
D. ministra (yellow-necked caterpillar)
Dendrolimus pini
D. sibiricus
Depressaria marcella (a webworm)
Desmia funeralis (grape leaf folder)
Diachrysia (Plusia) orichalcea (a semilooper)
Diacrisia virginica (yellow woollybear)
Diaphania (Margaronia) indica
D. nitidalis (pickleworm)

Diaphora mendica
 Diatraea grandiosella (southwestern corn borer)
 D. saccharalis (sugarcane borer)
 Dichomeris marginella (juniper webworm)
 Drymonia ruficornis (as chaonia)
 Drymonia sp.
 Dryocampa rubicunda
 Earias insulana
 Ectropis (Boarmia) crepuscularia
 Ennomos subsignarius (elm spanworm)
 Ephestia (Cadra) cautella (almond moth)
 E. elutella (tobacco moth)
 E. (Anagasta) kuehniella (Mediterranean flour moth)
 Epinotia tsugana (a skeletonizer)
 Epiphyas postvittana
 Erannis defoliaria (mottled umber moth)
 E. tiliaria (linden looper)
 Erinnyis ello
 Eriogaster henkei
 E. lanestris
 Estigmene acrea (salt marsh caterpillar)
 Eublemma amabilis
 Euphydryas chalcedona
 Eupoecilia ambiguella
 Euproctis chrysorrhoea (Nygmi phaeorrhoea) (brown tail moth)
 E. fraterna
 E. pseudoconspersa
 Eupterote fabia
 Eutromula (Simaethis) pariana
 Euxoa messoria (dark-sided cutworm)
 Galleria mellonella (greater wax moth)
 Gastropacha quercifolia
 Halisdota argentata
 H. caryae (hickory tussock moth)
 Harrisina brillians (western grape skeletonizer)
 Hedya nubiferana (fruit tree tortrix moth)

Heliothis (Helicoverpa) armigera (*Heliothis* = *Chloridea*) (gram pod borer)
H. (H.) assulta
Heliothis peltigera
H. virescens (tobacco budworm)
H. viriplaca
H. zea (cotton bollworm, corn earworm, soybean podworm, tomato fruitworm, sorghum headworm, etc.)
Hellula undalis (cabbage webworm)
Herpetogramma phaeopteralis
Heterocampa guttivitta (saddled prominent)
H. manteo (variable oak leaf caterpillar)
Holcocera pulverea
Homoeosoma electellum (sunflower moth)
Homona magnanima
Hyloicus pinastri
Hypeuryntis coricopa
Hyphantria cunea (fall webworm)
Hypogymna morio
Itame (Thamnonoma) wauaria (a spanworm)
Junonia coenia (buckeye caterpillars)
Kakivoria flavofasciata
Keiferia (Gnorimoschema) lycopersicella (tomato pinworm)
Lacanobia (Polia) oleracea
Lamdina athasaria pellucidaria
L. fiscellaria fiscellaria (hemlock looper)
L. fiscellaria lugubrosa
L. fiscellaria somniaria
Lampides boeticus
Leucoma (Stilpnotia) salicis (satin moth)
L. wiltshirei
Lobesia (= Polychrosis) botrana
Loxostege commixtalis (alfalfa webworm)
L. sticticalis (beet webworm)
Lymantria (Porthetria) dispar (gypsy moth) (*Lymantria*: tussock moths)
L. monacha (nun-moth caterpillar)
Malacosoma americana (eastern tent caterpillar)

M. disstria (forest tent caterpillar)
M. fragilis (= fragile) (Great Basin tent caterpillar)
M. neustria (tent caterpillar, lackey moth)
M. neustria var. *testacea*
M. pluviale (western tent caterpillar)
Mamestra brassicae (cabbage moth)
Manduca (*Inotoparce*) *quinquemaculata* (tomato hornworm)
M. (I.) sexta (tobacco hornworm)
Maruca testulalis
Melanolophia imitata
Mesographe forficalis
Mocis repanda (*Mocis*: semilooper)
Molippa sabina
Monema flavescens
Mythimna (*Pseudaletia*) *unipuncta* (armyworm)
Nephantis serinopa
Noctua (*Triphaena*) *pronuba*
Nomophila noctuella (lucerne moth)
Nymphalis antiopa (mourning-cloak butterfly)
Oiketicus moyanoi
Ommatopteryx texana
Operophtera brumata (winter moth)
Opsophanes sp.
O. fagata
Orgyia (*Hemerocampa*) *antiqua*
O. leucostigma (white-marked tussock moth)
O. (H.) pseudotsugata (Douglas-fir tussock moth)
O. thyellina
Orthosia gothica
Ostrinia (*Pyrausta*) *nubilalis* (European corn borer)
Paleacrita vernata (spring cankerworm)
Pammene juliana
Pandemis dumetana
P. pyrusana
Panolis flammea
Papilio cressphontes (orange dog)

P. demoleus
P. philenor
Paralipsa (Aphemia) gularis
Paralobesia viteana
Paramyelois transitella
Parnara guttata
Pectinophora gossypiella (pink bollworm)
Pericallia ricini
Peridroma saucia (variegated cutworm)
Phalera bucephala
Phlogophora meticulosa
Phryganidia californica (California oakworm)
Phthorimaea (= Gnprimoschema) operculella (potato tuberworms)
Phyllonorycter (Lithocolletis) blancardella
Pieris brassicae (large white butterfly)
P. canidia sordida
P. rapae (imported cabbageworm, small white butterfly)
Plathypena scabra (green cloverworm)
Platynota sp.
P. stultana
Platyptilia carduidactyla (artichoke plume moth)
Plodia interpunctella (Indian-meal moth)
Plutella xylostella as *maculipennis* (diamondback moth)
Prays citri (citrus flower moth)
P. oleae (olive moth)
Pseudoplusia includens (soybean looper)
Pygaera anastomosis
Rachiplusia ou
Rhyacionia buoliana (European pine shoot moth)
Sabulodes caberata
Samia cynthia
Saturnia pavonia
Schizura concinna (red-humped caterpillar)
Schoenobius bipunctifer
Selenephra lunigera
Sesamia inferens

Sibine apicalis
Sitotroga cerealella (Angoumois grain moth)
Sparganothis pilleriana
Spilonota (Tmetocera) *ocellana* (eye-spotted budmoth)
Spilosoma lubricipeda (as *menthastri*)
S. virginica
Spilosoma sp.
Spodoptera (*Prodenia*) *eridania* (southern armyworm)
S. exigua (beet armyworm, lucerne caterpillar)
S. frugiperda
S. littoralis
S. litura
S. mauritia
S. (P.) ornithogalli (yellow-striped armyworm)
S. (P.) praefica
Syllepte derogata
S. silicalis
Symmerista canicosta
Thaumetopoea pityocampa (pine processionary caterpillar)
T. processionea
T. wauaria (currant webworm)
T. wilkinsoni
Thymelicus lineola (European skipper)
Thyridopteryx ephemeriformis (bagworm)
Tineola bisselliella (webbing clothes moth)
Tortrix viridana (oak tortricid)
Trichoplusia ni (cabbage looper)
Udea profundalis (celery leaf tier)
U. rubigalis
Vanessa cardui (painted-lady)
V. io
Xanthopastis timais
Xestia (*Amathes*, *Agrotis*) *c-nigrum* (spotted cutworm)
Yponomeuta cognatella (= *Y. evonymi*) (*Yponomeuta* = *Hyponomeuta*)
Y. evonymella
Y. mahalebella

Y. malinella (small ermine moth)

Y. padella (small ermine moth)

Y. rorrella

Zeiraphera diniana

MALLOPHAGA

Bovicola bovis (cattle biting louse)

B. crassipes

B. limbata

B. ovis

Lipeurus caponis (wing louse)

Menacnathus stramineus

Menopon gallinae (shaft louse)

TRICHOPTERA

Hydropsyche pellucida

Potamophylax rotundipennis

TABLE 2

Plants recommended for protection by B. thuringiensis insecticidal protein

alfalfa	escarole	potatoes
almonds	field corn	radishes
apples	filberts	rangeland
artichokes	flowers	raspberries
avocados	forage crops	safflower
bananas	forest trees	shade trees
beans	fruit trees	shingiku
beets	garlic	small grains
blackberries	grapes	soybeans
blueberries	hay	spinach
broccoli	kale	squash
brussels sprouts	kiwi	stonefruits
cabbage	kohlrabi	stored corn
caneberries	lentils	stored grains
carrots	lettuce	stored oilseeds
cauliflower	melons	stored peanuts
celery	mint	stored soybeans
chard	mustard greens	stored tobacco
cherries	nectarines	strawberries
chinese cabbage	onions	sugarbeets
chrysanthemums	oranges	sugar maple
citrus	ornamental trees	sunflower
collards	parsley	sweet corn
cos lettuce	pasture	sweet potatoes
cotton	peaches	tobacco
cranberries	peanuts	tomatoes
crop seed	pears	turf
cucumbers	peas	turnip greens
currants	pecans	walnuts
dewberries	peppers	watermelons
eggplant	pome fruit	
endive	pomegranite	

TABLE 3

Varieties of *B. thuringiensis*

alesti
aizawai
canadensis
dakota
darmstadiensis
dendrolimus
entomocidus
finitimus
fowleri
galleriae
indiana
israelensis
kenyae
kurstaki
kyushuensis
morrisoni
ostrinae
pakistani
sotto
thompsoni
thuringiensis
tolworthi
toumanoffi
wuhanensis

TABLE 4

Index of plasmids and strains

Strain or Plasmid	Constructed or Used in Example	See Figure	Made From (α Comments)
<u>A. tumefaciens</u>	6		(ubiquitous)
<u>A. rhizogenes</u>	5		(also see background)
<u>B. thuringiensis</u> var.			
<u>kurstaki</u> HD-73	1.1	1	
ColE1	2.5		
<u>E. coli</u> GM33	2.3		
<u>E. coli</u> HB101	1.1		
<u>E. coli</u> JM103	2.1		
<u>E. coli</u> K802	2.2		
MBT3	3.3		M13mp8, p123/58-10
MBT3(<u>Nco</u>)	3.4		MBT3
MBT14	3.3		M13mp8, p123/58-10
mWB2344	2.1		
M13-Bt-A	2.1		mWB2344, p123/58-10
M13-Bt-A(<u>Bam</u>)	2.1		M13-Bt-A
M13-Bt-S	2.1		mWB2344, p123/58-10
M13mp7	3.1		
M13mp8	3.3		
M13-PpBt	4.4		MBT3(<u>Nco</u>), M13-3.8Ab
M13-1	3.1		M13mp7, pNS5
M13-3	3.1		M13mp7, pNS5
M13-3A/B18a	3.2		M13-3
M13-3.8A	4.1		M13mp7, 177.4
M13-3.8Aa	4.2		M13-3.8Ac
M13-3.8Ab	4.3		M13-3.8Aa
M13-3.8Ac	4.2		M13-3.8A
M13-3.8S	4.1		M13mp7, 177.4
pBR322	1.1		

PCF44	3.1		pBR322, pTic58
pCF44A	3.1		pCF44
pKS- <u>proI</u>	2.2	3	pKS111, = pTR- <u>proI</u>
pKS- <u>proI</u> (<u>Bam</u>)	2.2	2.2	pKS- <u>proI</u>
pKS-4	2.5	2	pBR322, pRZ102
pKS111	2.2, 12.3	2, 3	pRK290, pTi15955
pKS111-K	4.5		pKS4(pRZ102), pKS111
pKS111-N	3.5		pCF44, pKS111-K
pKS111-NpBt	3.5		MBT3(<u>Nco</u>), M13-3A/B18a,
pKS111-N			
pKS111-PpBt	4.5		M13-PpBt, pKS111-K
pNS5	3.1		pBR322, pCF44A
ppH1J1	9		
pRK290	2.2, 9		
pRK2013	9		
pRZ102	2.5		ColE1, Tn5
pTiA66	2.4		
pTi15955	2.4	2	
p8.8	4.1		pBR322, 177.4
p11-83a	2.3	3	pKS- <u>proI</u> (<u>Bam</u>), pKS-4
p11-83b	2.3	3	p11-83a, M13-Bt-A(<u>Bam</u>)
p123/58-3	1.1	1	<u>B. thuringiensis</u> var.
			<u>kurstaki</u> HD-73, pBR322
p123/58-10.	1.1	1	<u>B. thuringiensis</u> var.
			<u>kurstaki</u> HD-73, pBR322
p403	2.2	2	pBR322, pTi15955
"1.6"	2.2	2	(= transcript 24, see
			also Detailed Description)
177.4	4.1		Charon 24A, <u>P. vulgaris</u>
			cv. Tendergreen
pBt73-161	11.1		<u>B. thuringiensis</u> var.
			<u>kurstaki</u> HD-73, pBR322
pBt73-10(<u>Bam</u>)	11.2		p123/58-10
pBt73-16	11.2	4	pBt73-10(<u>Bam</u>), pBt73-161
pTR- <u>proI</u>	12.1	4	= pKS- <u>proI</u>

pTR- <u>proI</u> (Bam)	12.1	4	pTR- <u>proI</u>
pBR325	12.3	4	
pBR325aBB	12.3	4	pBR325
p403B	12.1	4	pBR325aBB, pTR- <u>proI</u> (Bam)
M13mp19	12.4	4	
1.6.4	12.4	4	M13mp19, pBt73-16
1.6.4B-3.8.3	12.4	4	1.6.4
p403B/BTBΔ3	12.3	4	1.6.4B-3.8.3, p403B
pH4-1	12.1		pSUP106, pTi15955, CaMV, Tn5
			pH4-1
pH400	12.1		CaMV, pBR322
pD0B412	12.2		pD0B412
pD0B512	12.2		p403B
p403BRL1	12.3		pD0B513
pD0B514	12.3		p403BRL1, pD0B514
pMAN514	12.3		pKS4
pKS4.2	12.3		pKS4.2
pKS4.3	12.3		pBR322, <u>lambda</u>
pBR322Bam	12.3		pKS4.3, pBR322 Bam ⁻
p11-83c	12.3		1.6.4B-3.8.3, pCG116
pCJ161	12.5		pCJ161, pH400
pH450	12.6		p11-83c, pTR- <u>proI</u> (Bam)
pCG116	12.3		

TABLE 5

Deposited Strains

NRRL B-4488	<u>B. thuringiensis</u> var. <u>kurstaki</u> HD-73
NRRL B-15394	<u>E. coli</u> C600 (pKS-4)
NRRL B-11371	<u>E. coli</u> HB101
NRRL B-12014	<u>E. coli</u> RR1 (pBR322)
ATCC 37017	pBR322
ATCC 15955	<u>A. tumefaciens</u> (pTi15955)
NRRL B-15393	<u>E. coli</u> HB101 (p8.8)
NRRL B-15612	<u>E. coli</u> HB101 (p123/58-10)
NRRL B-5759	<u>E. coli</u> HB101 (pBt73-16)
NRRL B-18009	<u>E. coli</u> K802 (pH4-1)
NRRL B-15486	<u>E. coli</u> CSH52 (pSUP106)

TABLE 6

<u>MS Medium</u>		
NH_4HCO_3	1.65	g/l
KNO_3	1.9	g/l
$\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$	440	mg/l
$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$	370	mg/l
KH_2PO_4	170	mg/l
KI	0.83	mg/l
H_3BO_3	6.2	mg/l
$\text{MnSO}_4 \cdot 4\text{H}_2\text{O}$	22.3	mg/l
$\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$	8.6	mg/l
$\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$	0.25	mg/l
$\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$	0.025	mg/l
$\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$	0.025	mg/l
Na_2EDTA	37.23	mg/l
$\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	27.85	mg/l
Inositol	1	g/l
Nicotinic acid	50	mg/l
Pyroxidine HCl	50	mg/l
Thiamine HCl	50	mg/l
Sucrose	30	g/l
Agar	8	g/l

TABLE 7

Clone	Experiment 1 ⁵		Experiment 2 ⁶	
	A ₄₉₂ ¹	CP ²	A ₄₉₂	CP
100	0.17 ± 0.03 ⁷	+	0.14	+
101	0.02 ± 0.01	-	0.07	±
102	0.06 ± 0.02	±	0.21	+
103	0.00 ± 0.00	-	ND	ND
104	ND ³	ND	0.11	+
105	0.00 ± 0.00	-	0.10	+
106	0.10 ± 0.02	+	0.01	-
107	0.06 ± 0.03	±	0.06	-
109	0.11 ± 0.05	+	0.03	-
110	0.06 ± 0.01	±	0.13	+
111	0.12 ± 0.02	+	0.00	-
NX ⁴	-0-		-0-	

¹Average of three ELISA determinations. Absorbance at 492 nm, corrected by subtracting the value for the NX control.

²Rated as having (+) or not having (-) crystal protein. ± indicates a marginal rating.

³Not Determined.

⁴Untransformed *Nicotiana tabacum* var. "Xanthi" control. In experiments 1 and 2, respectively, 18 and 19 control leaves obtained from different plants were averaged.

⁵Experiment 1 was standardized for equal tissue wet weight.

⁶Experiment 2 was standardized for equal plant protein concentrations.

⁷± standard error of the mean.

TABLE 8

<u>Clone Number</u>	<u>Replicate Number</u>	<u>Total Larvae</u>	<u>Dead Larvae</u>
100	1	4	1
	2	5	0
	3	5	4
103	1	5	0
	2	5	0
	3	5	0
106	1	5	0
	2	5	0
	3	5	0
109	1	5	2
	2	5	0
	3	5	3
111	1	5	2
	2	5	1
	3	5	0

TABLE 9

<u>Clone Number</u>	<u>Replicate Number</u>	<u>Total Larvae</u>	<u>Dead Larvae</u>
100	1	4	2
	2	4	3
	3	6	3
103	1	6	0
	2	6	2
	3	6	2
106	1	6	2
	2	8	3
	3	10	6
109	1	14	2
	2	6	1
	3	6	4
111	1	6	3
	2	6	2
	3	6	3

CLAIMS

We claim:

1. A plant comprising a genetically modified plant cell containing an insecticide structural gene under control of a plant expressible promoter.
2. A plant according to claim 1 wherein the cell contains T-DNA, and the insecticide structural gene is inserted therein.
3. A plant according to claim 2 wherein the insecticide gene is inserted at an actively transcribed site within either the tml gene, the ocs gene, or the "1.6" region of the T-DNA.
4. A plant according to claim 2 wherein the T-DNA is modified to inactivate tml, tms, or tmr.
5. A plant according to claim 2 wherein the T-DNA additionally comprises a functional ocs gene.
6. A plant according to claim 1 wherein the insecticide structural gene is obtained from B. thuringiensis.
7. A plant tissue comprising a genetically modified plant cell containing an insecticide structural gene under control of a plant expressible promoter.
8. A tissue according to claim 7 wherein the cell contains T-DNA, and the insecticide gene is inserted within the T-DNA.
9. A tissue according to claim 8 wherein the insecticide gene is inserted at an actively transcribed site within either the tml gene, the ocs gene, or the "1.6" region of the T-DNA.
10. A tissue according to claim 8 wherein the T-DNA is modified.
11. A tissue according to claim 10 wherein the modification inactivates tml, tmr, or tms.
12. A tissue according to claim 8 wherein the T-DNA additionally comprises a functional ocs gene.
13. A tissue according to claim 8 wherein the plant is a gymnosperm.
14. A tissue according to claim 8 wherein the plant is an angiosperm.
15. A tissue according to claim 14 wherein the plant is a monocot.
16. A tissue according to claim 14 wherein the plant is a dicot.
17. A tissue according to claim 16 wherein the plant is a member of the family Solanaceae.

18. A tissue according to claim 16 wherein the plant is a member of the family Leguminosae.
19. A tissue according to claim 16 wherein the plant is a member of the family Compositae.
20. A tissue according to claim 16 wherein the plant is a member of the family Malvaceae.
21. A tissue according to claim 14 wherein the plant is a vegetable.
22. A tissue according to claim 7 wherein the structural gene is under control of a T-DNA promoter.
23. A tissue according to claim 22 wherein the structural gene is under control of a promoter selected from the group of T-DNA genes consisting of tmr, tml, tms, nos, ocs, and the "1.6" transcript.
24. A tissue according to claim 7 wherein the insecticide structural gene is under control of a plant promoter.
25. A tissue according to claim 24 wherein the structural gene is under control of a promoter of a plant gene selected from a group of plant genes consisting of phaseolin and the small subunit of ribulose-1,5-bisphosphate carboxylase.
26. A tissue as described in claim 7 wherein the insecticide structural gene is under control of a CaMV promoter selected from the group of promoters which control the 35S and 19S transcripts.
27. A tissue according to claim 7 wherein the insecticide structural gene is modified.
28. A tissue according to claim 27 wherein the insecticide structural gene is modified to code for added amino acids at either end of the insecticidal protein.
29. A tissue according to claim 27 wherein the modification comprises an insertion, a deletion, or a substitution of one or more nucleotides in the structural gene nucleotide sequence.
30. A tissue according to claim 7 wherein the insecticide structural gene is obtained from B. thuringiensis.
31. A tissue according to claim ~~30~~ wherein the structural gene is hybridizable to that of p123/58-10.
32. A DNA vector comprising an insecticide structural gene and a plant expressible promoter, the gene and the promoter being in such posi-

tion and orientation with respect to each other that the gene is expressible under control of the promoter in a plant cell.

33. A vector according to claim 32 wherein the insecticide gene is inserted in T-DNA.
34. A claim according to claim 33 wherein the T-DNA is modified.
35. A vector according to claim 33 wherein the gene is inserted at an actively transcribed site within either the tml gene, ocs gene, or the "1.6" region of the T-DNA.
36. A vector according to claim 33 wherein the T-DNA additionally comprises a functional ocs gene.
37. A vector according to claim 32 wherein the promoter is a T-DNA promoter.
38. A vector according to claim 32 wherein the promoter is a plant promoter.
39. A vector as described in claim 38 wherein the plant promoter is selected from the group of plant genes consisting of phaseolin and the small subunit of ribulose-1,5-bisphosphate carboxylase.
40. A vector as described in claim 32 wherein the promoter is a CaMV promoter selected from the group of promoters which control the 35S and 19S transcripts.
41. A vector according to claim 32 wherein the insecticide structural gene is modified, the modification comprising an insertion, a deletion, or a substitution of one or more nucleotides in the structural gene-nucleotide sequence.
42. A vector according to claim 32 wherein the insecticide structural gene is from B. thuringiensis.
43. A vector according to claim 42 wherein the structural gene is hybridizable to that of p123/58-10.
44. A vector according to claim 32 wherein the promoter is modified, the modification comprising an insertion, a deletion, or a substitution of one or more nucleotides in the promoter sequence.
45. A bacterial strain comprising an insecticide structural gene and a plant expressible promoter, the gene and the promoter being in such position and orientation with respect to each other that the gene is expressible under control of the promoter in a plant cell. ✓

46. A strain according to claim 45 wherein the expressible gene is within T-DNA.
47. A strain according to claim 46 wherein the T-DNA is within a cell selected from the genus Agrobacterium.
48. A strain according to claim 45 wherein the insecticide structural gene is from B. thuringiensis.
49. A strain according to claim 48 wherein the structural gene is hybridizable to that of p123/58-10.
50. A plasmid selected from the group p123/58-10 and p123/58-3.
51. A bacterial strain comprising a plasmid selected from the group p123/58-10 and p123/58-3.
52. A strain according to claim 51 wherein the bacteria is E. coli HB101.
53. A method of genetically modifying a plant cell by transforming the cell to contain an insecticide structural gene and a plant expressible promoter whereby the gene is expressible in the plant cell under control of the promoter.
54. A method according to claim 53 wherein the insecticide gene is under control of a plant promoter.
55. A method according to claim 54 wherein the insecticide gene is under control of a promoter selected from the group of plant genes consisting of phaseolin and the small subunit of ribulose-1,5-bisphosphate carboxylase.
56. A method according to claim 53 wherein the insecticide structural gene is under control of a T-DNA promoter.
57. A method according to claim 56 wherein the structural is under control of a promoter selected from a group of T-DNA genes consisting of tmr, tml, tms, nos, ocs, and the "1.6" transcript.
58. A method according to claim 53 wherein the insecticide gene is under control of a promoter selected from the group CaMV promoters which control the 35S and 19S transcripts.
59. A method according to claim 53 wherein the insecticide gene is modified, the modification comprising an insertion, a deletion, or a substitution of one or more nucleotides in the structural gene nucleotide sequence.
60. A method according to claim 53 wherein the gene is inserted into T-DNA.

61. A method according to claim 60 wherein the T-DNA is modified, the modification comprising an insertion, a deletion, or a substitution of one or more nucleotides in the T-DNA nucleotide sequence.
62. A method according to claim 60 wherein the transformed cell is from a gymnosperm.
63. A method according to claim 60 wherein the transformed cell is from an angiosperm.
64. A method according to claim 63 wherein the angiosperm is a monocot.
65. A method according to claim 63 wherein the angiosperm is a dicot.
66. A method according to claim 65 wherein the dicot is a member of the family Solanaceae.
67. A method according to claim 65 wherein the dicot is a member of the family Compositae.
68. A method according to claim 65 wherein the dicot is a member of the family Leguminosae.
69. A method according to claim 65 wherein the dicot is a member of the family Malvaceae.
70. A method according to claim 63 wherein the dicot is a vegetable.
71. A method according to claim 60 wherein the insecticide gene is inserted at an actively transcribed site within either the tml gene, the ocs gene, or the "1.6" region of the T-DNA.
72. A method according to claim 60 wherein the plant cell is transformed by transfer of DNA from a bacterium to the plant cell.
73. A method according to claim 53 wherein the plant cell is transformed by direct uptake of DNA or by microinjection of DNA into the plant cell.
74. A method according to claim 53 comprising additionally the step of regenerating said transformed cell to yield transformed regenerated plant tissue capable of expressing the insecticide structural gene.
75. A method according to claim 53 wherein the insecticide structural gene is obtained from B. thuringiensis.
76. A method according to claim 75 wherein the insecticide structural gene is hybridizable to that found in p123/58-10.

ABSTRACT

A method for introducing expressible insecticidal protein structural genes into plant genomes is provided. In the preferred embodiments this invention comprises placing a structural gene for the Bacillus thuringiensis crystal protein under control of a plant or a T-DNA promoter and ahead of a polyadenylation site followed by insertion of said promoter/structural gene combination into a plant genome by utilizing a Agrobacterium tumefaciens Ti plasmid-based transformation system. The modified Ti plasmid is then used to transform recipient plant cells. Also provided are the plants and tissues produced by this method and bacterial strains, plasmids, and vectors useful for execution of this invention.

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April 4, 1986

(Date of deposit)

Andrea L. Ryder

Name of Applicant, Assignee or Registered Representative

Signature

4-4-86

Date of signature

B 59945493

Express Mail Receipt No.

Partial Sequence of the Crystal Protein Gene

AAGTGGATTTATATATAAGTATAAAAGTAATAAGCACTTTAAATAAGTTAACGGAATACAAACCTTAATGCATTGGTTAAACATTGTAAGTCTAAA
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 HeLAspAsnAsnPr
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 SerLeuSerLeuThrGlnPheLeuLeuSerGluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIleTrpGlyIlePheGlyProSerG
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 eTyrThrAspAlaHisArgGlyIleTyrTrpSerGlyHisGlnIleMetAlaSerProValGlyPheSerGlyProGluPheThrPheProLeuTyr
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 snIleGlyIleAsnAsnGlnGlnLeuSerValLeuAspGlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaValTyrArgLysSerG

FIG. 1 - 1

AACGGTAGATTGGCTGGATCAAAATACGGCCACAGATAACACGCTGCCACCTAGGCAAGGATTAGTCATCGATTAGCCATGTTTCAATGTTTCGTCA
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 hrGlnilleProAlaValLysGlyAsnPheLeuPheAsnGlySerValilleSerGlyProGlyPheThrGlyGlyAspLeuValArgLeuAsnSerSerGly
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 sLeuGluTrpAsnProAspLeuAspCysSerCysArgAspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspilleAspValGlyCysThr
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FIG. 1 - 2

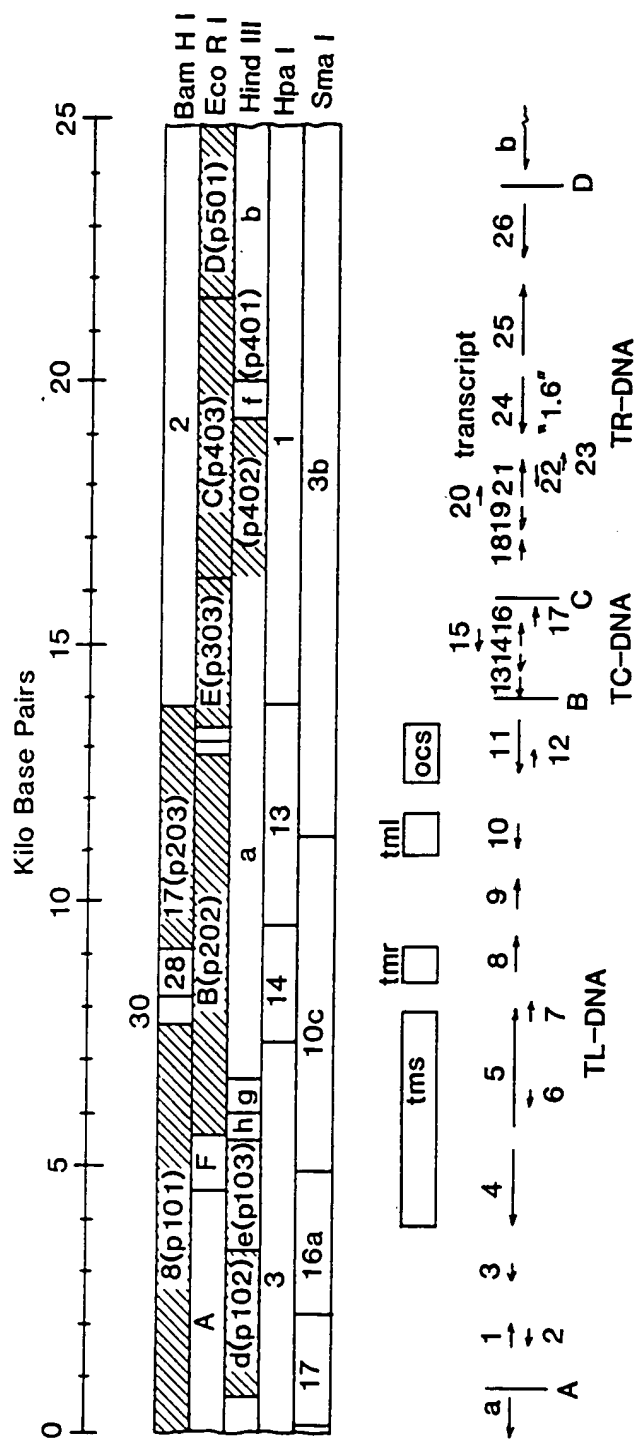


FIG. 2

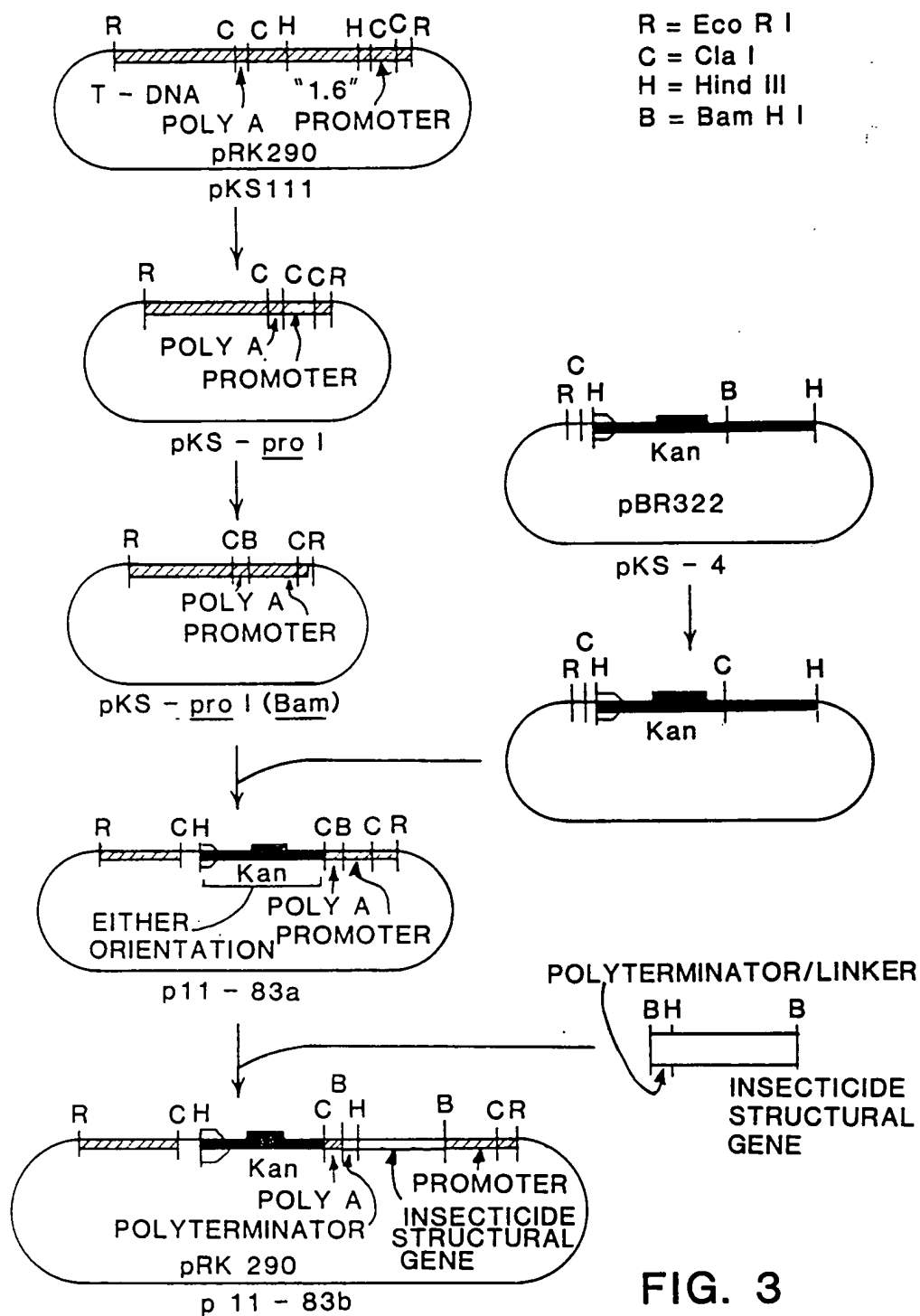


FIG. 3

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 hrGln11eProAlaValLysGlyAsnPheLeuPheAsnGlySerVal11eSerGlyProGlyPheThrGlyGlyAspLeuValArgLeuAsnSerSerG
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 yAsnAsn11eGlnAsnArgGlyTyr11eGluValPro11eHisPheProSerThrSerThrArgTyrArgValArgValArgTyrAlaSerValThrPro
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 AGCATTCGAGAGCTT
 Ser11eArgGluAlaT 3116

FIG. 1 - 2

Restriction map of the BamHI-EcoRI fragment of the pBR322 plasmid. The map shows a linear representation of the 25 kb plasmid with various restriction sites and fragments labeled. The fragments are: 8(p101), 28, 17(p203), B(p202), F, E(p303), C(p403), D(p501), A, d(p102), e(p103), h, g, a, 13, 14, 10c, 17, 16a, 3, 10, 1, 3b, 25. The fragments are arranged in a circular fashion, with the BamHI site at the top and the EcoRI site at the bottom.

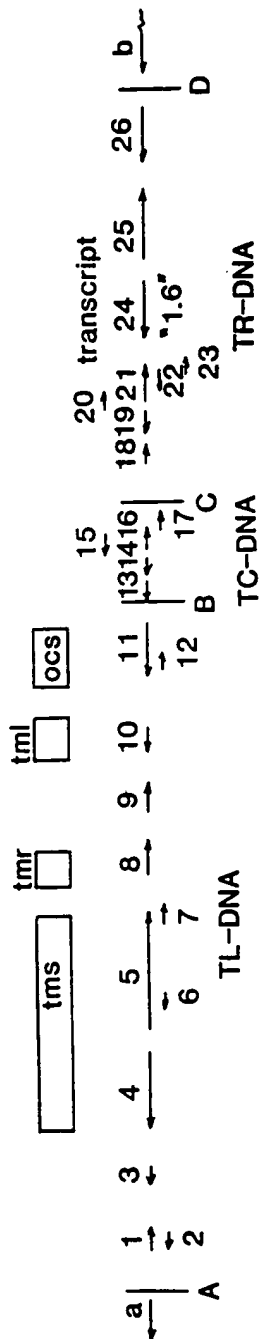


FIG. 2

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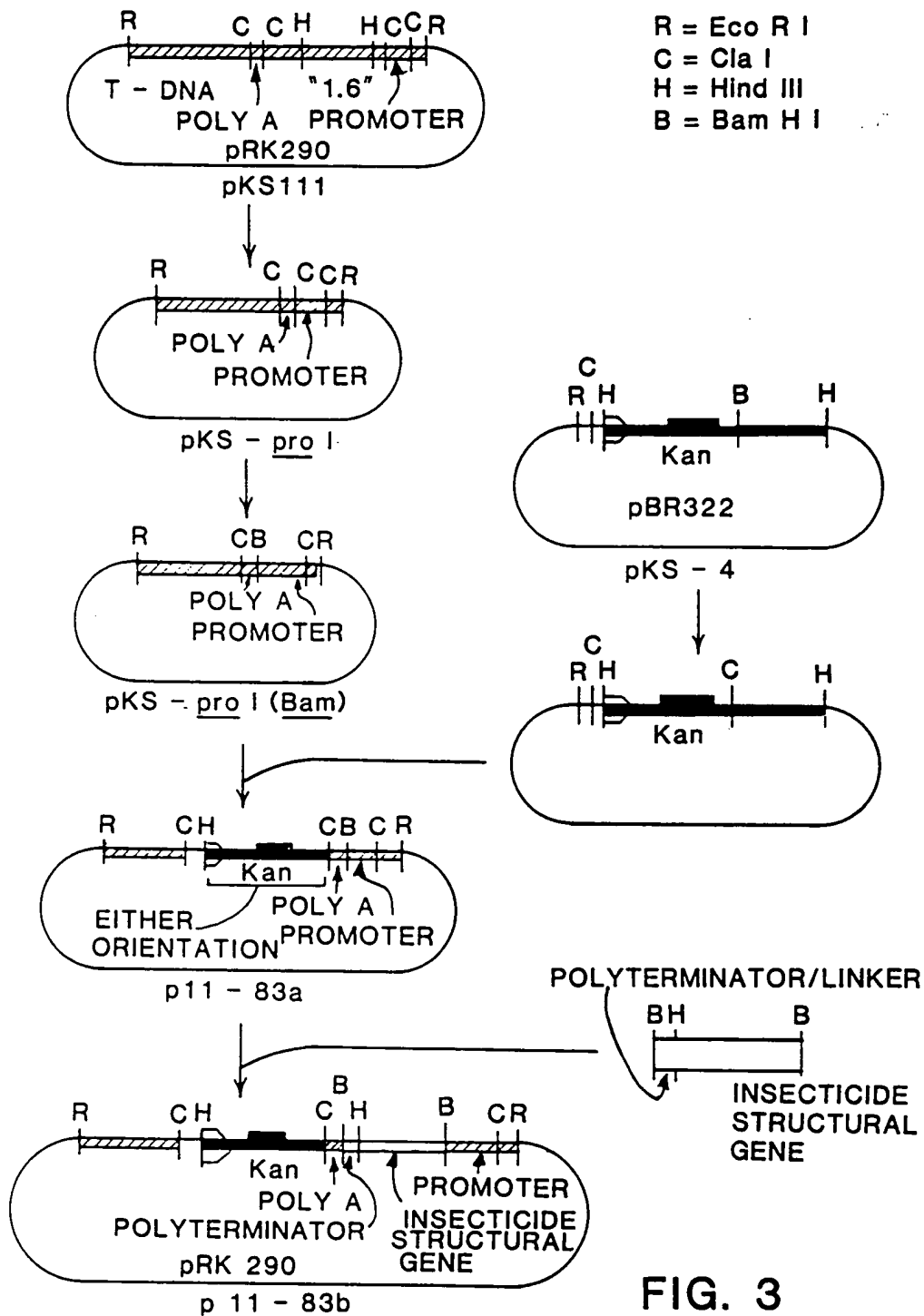


FIG. 3

Partial Sequence of the Crystal Protein Gene

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 MetAspAsnAsnPr
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 ThrValLeuAspIleValAlaLeuPheProAsnTyrAspSerArgArgTyrProIleArgThrValSerGlnLeuThrArgGluIleTyrThrAsnProV
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 oLeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGluArgSerIleArgSerSerHisLeuMetAspIleLeuAsnSerIleThrI
 CTATACGATGCTCATAGGGGTTATTATTATGTCAGGGCATCAAAATATGGCTTCTCTGTAGGGTTTTTCGGGGCCAGAATTCACTTTTCCGCTATAT
 eTyrThrAspAlaHisArgGlyTyrTyrTyrTrpSerGlyHisGlnIleMetAlaSerProValGlyPheSerGlyProGluPheThrPheProLeuTyr
 GCACTATCGGAAATCGAGCTCCACAAACCTATTGTTGCTCAACTAGGTCAAGGCGTGTATAGAACATTATGTCACCTTTATATAGAACCTTTTA
 GlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAlaGlnLeuGlyGlnGlyValTyrArgThrLeuSerSerThrLeuTyrArgArgProPheA
 ATATAGGGATAAATCAACAACTATCTGTTCTTACCGGACAGAAATTTGCTTATGGAACCTCCTCAAAATTTGCCATCCGCTGTATACAGAAAAAGCGG
 snIleGlyIleAsnAsnGlnGlnLeuSerValLeuAspGlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaValTyrArgLysSerGI

FIG. 1 - 1

AACGGTAGATTCCTGGATGAATACGCCACAGAAACACGTCGCCACATAGCAAGGATTTAGTCATGATTAAGCCATGTTTCAATGTTTCCTTCA
yThrValAspSerLeuAspGlu11leProProGlnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHisValSerMetPheArgSer
GGCTTTAGTAATAGTAGTGTAAATATAAAGAGCTCCTATGTTCTCTGGATACATCGTAGTGTGAATTTAATAATAATATGATCGGATAGTATTA
GlyPheSerAsnSerSerValSer11leArgAlaProMetPheSerTrp11leHisArgSerAlaGluPheAsnAsn11leAlaSerAspSer11leT
CTCAATCCCTGCAGTGAAGGAAACCTTTCTTTTAAATGTTCTGTAAATTTAGGACCAAGGATTTACTGGTGGGACTTAGTTAGATTAATAGTATB
hrGln11leProAlaValLysGlyAsnPheLeuPheAsnGlySerVal11leSerGlyProGlyPheThrGlyGlyAspLeuValArgLeuAsnSerSerG
AAATAACATTCAGAATAGAGGTATATTGAATTCATTCACATCCCATCCACATCTACCAGATATCGAGTTCGTGTACGGTATGCTTCTGTAAACCC
AsnAsn11leGlnAsnArgGlyTyr11leGluValPro11leHisPheProSerThrSerThrArgTyrArgValArgValArgTyrAlaSerValThrPro
ATTCACTCAACCTTAATGGGGTAATTCATCCATTTTTTCCAATACAGTACCAGCTACAGCTACGTCATTAGATAATCTACAATCAAGTATTGTTGTT
11leHisLeuAsnValAsnTrpGlyAsnSerSer11lePheSerAsnThrValProAlaThrAlaThrSerLeuAspAsnLeuGlnSerSerAspPheGlyT
ATTTTGAAGTCCCAATGCTTTTACATCTTCATAGGTAATAGTAGGTTTGAATTTTGGGACTGCAGGAGTGAATAACAGATTTGAATTT
yPheGluSerAlaAsnAlaPheThrSerSerLeuGlyAsn11leValGlyValArgAsnPheSerGlyThrAlaGlyVal11leAspArgPheGluPh
TATTCACTTACTGCAACACTCGAGGCTGAATATAATCTGGAAAGAGCCGAGAAGCGGTGAATGGCTGTTTACGTCTACAAACCACTAGGCTAAAA
AlaProValThrAlaThrLeuGlu11leGluTyrAsnLeuGluArgAlaGlnLysAlaValAlaAlaLeuPheThrSerThrAsnGlnLeuGlyLeuLys
ACAAATGTAAAGGATATCATATTGATCAAGTGTCCATTTTGTAGTACGATTTTATCGGATGAATTTTGTCTGGATGAAGGAGGAAATTTGTCGAGAAAG
ThrAsnValThrAspTyrHis11leAspGlnValSerAsnLeuValThrTyrLeuSerAspGluPheCysLeuAspGluLysArgGluLeuSerGluLysV
TCAAACATGCGAAGGCACTCAGTGAACGCAATTTACTCCAAGATTTCAAATTTCAAAGACATTAATAGGCAACGAGACGTGGTGGGCGGAAATAC
AlLysHisAlaLysAlaLeuSerAspGluArgAsnLeuGlnAspSerAsnPheLysAsp11leAsnArgGlnProGluArgGlyTrpGlyGlySerTh
AGGATTAACATCCAAAGAGGGGATGAGCTATTAAAGAAATATACGTACACATATCAGTACCTTTGATGAGTGTATCCAACATATTGTTATCAAAAA
Gly11leThr11leGlnGlyGlyAspValPheLysGluAsnTyrValThrLeuSerGlyThrPheAspGluCysTyrProThrTyrLeuTyrGlnLys
ATCGATGAATCAAAATTAAGCCCTTACCGGTATCAATTAAGAGGTATATCGAAGATAGTCAAGACTTAGAAATCTATTAAATTCGCTACAAATGCAA
11leAspGluSerLysLeuLysAlaPheThrArgTyrGlnLeuArgGlyTyr11leGluAspSerGlnAspLeuGlu11leTyrLeu11leArgTyrAsnAlaL
AACATGAACAGTAATGTGCCAGGTACGGGTCTTATGGCCGCTTTCAGCCCAAGTCCAATCGGAAAGTGTGGAGAGCCGAATCGATCGCGCCACA
ysHisGluThrValAsnValProGlyThrGlySerLeuTrpProLeuSerAlaGlnSerPro11leGlyLysCysGlyGluProAsnArgCysAlaProH
CCTTGAATGGAATCCTGATCTAGATTTGCTGTAGGATGGAGAAAGTGTGCCCATCATTCGCATCATTTCTCCTTAGACATTTGATAGATGTACA
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GACTTAAATCAGGACCTAGGTGTATGGGTGATCTTAAAGATTAAAGCCCAAGATGGGCACGCAAGACTAGGAAATCTACAGTTCTCGAAGAGAAACAT
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TAGTAGCAGAACGCTAGCTCGTGTGAAAAGACCGGAGAAAAATGGAGAGCAACGTAAGAAATTCGAATGGCAACAAATATGTTTATAAGAGGC
euValGlyGluAlaLeuAlaArgValLysArgAlaGluLysLysTrpArgAspLysArgGluLysLeuGluTrpGluThrAsn11leValTyrLysGluAl
AAAAGAACTCTAGATGCTTTATTTGTAACCTCTCAATATGATCAATTACAAGCGGATACGAATATTGCCATGATTCATCGCGCAGATAACGCTTTTCAT
LysGluSerValAspAlaLeuPheValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsn11leAlaMet11leHisAlaAlaAspLysArgValHis
AGCATTCAGAACGCTT
Ser11leArgGluAlaT

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FIG. 1 - 2

848733

Map of pTi15955 T-DNA

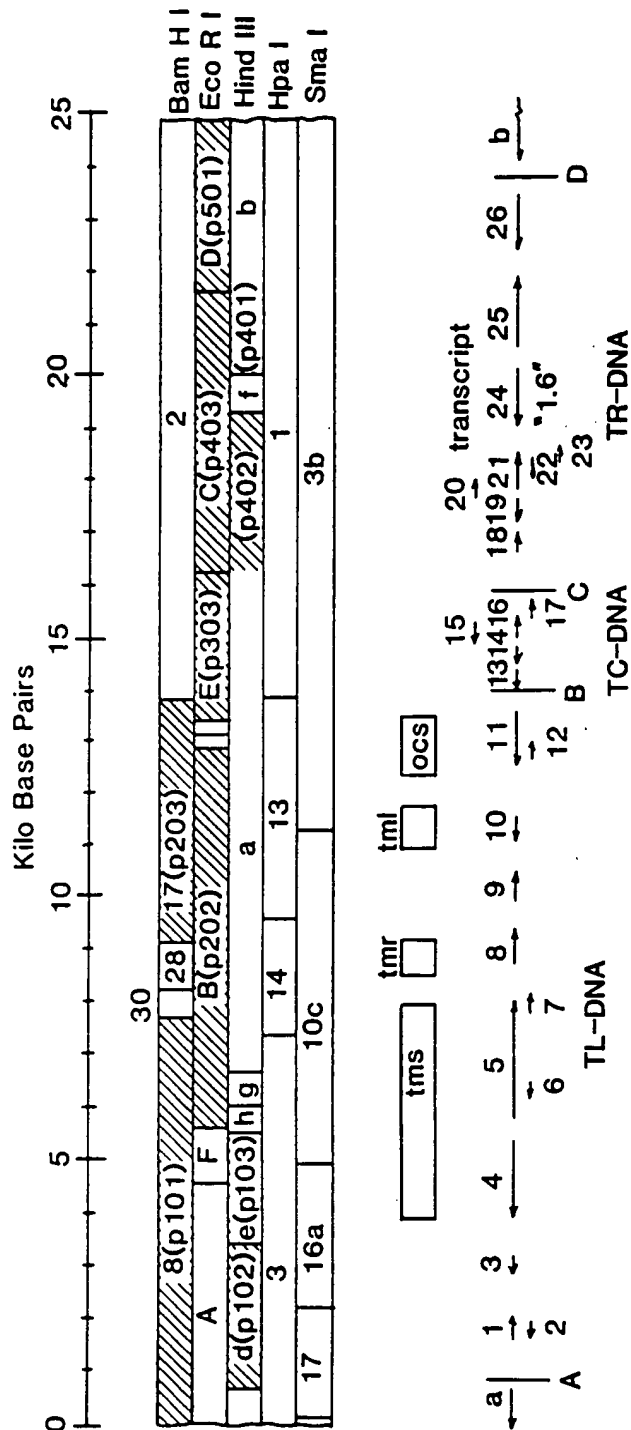


FIG. 2

848733

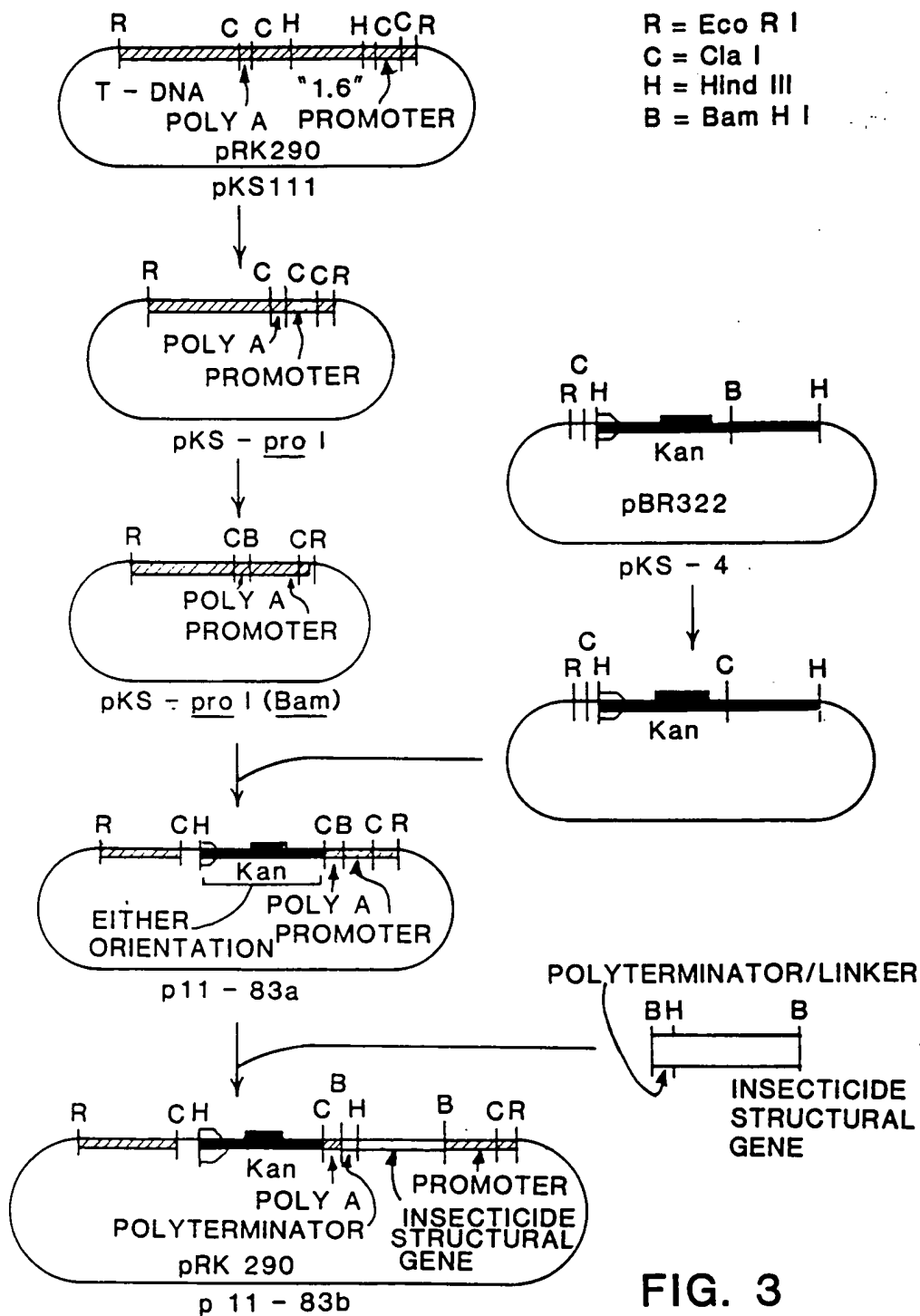


FIG. 3



UNITED STATES DEPARTMENT OF COMMERCE
Patent and Trademark Office

Address: COMMISSIONER OF PATENTS AND TRADEMARKS
Washington, D.C. 20231

Agrigenetics Corporation
3375 Mitchell Lane
Boulder, Colorado 80301

MAILED

MAY 23 1986

APPLICATION DIVISION
PATENT & TRADEMARK OFFICE

Applicant(s): Michael J. Adang et al
Serial Number: 848,733
Filing Date: April 4, 1986
Title: Insect Resistant Plants

Notice to File Missing Parts of Application-
Filing Date Granted

If all missing parts are filed within the period set below, the total amount owed by applicant as a ☒ large entity, ☐ small entity (verified statement filed), is \$ **110.00**

1. ☐ The statutory basic filing fee is: ☐ missing ☐ insufficient. Applicant as a ☐ large entity, ☐ small entity, must submit \$ _____ to complete the basic filing fee and MUST ALSO SUBMIT THE SURCHARGE AS INDICATED BELOW.
2. ☐ Additional claim fees of \$ _____ as a ☐ large entity, ☐ small entity, including any required multiple dependent claim fee, are required. Applicant must submit the additional claim fees or cancel the additional claims for which fees are due. NO SURCHARGE IS REQUIRED FOR THIS ITEM.
3. ☐ The oath or declaration is:
☐ missing.
☐ does not cover items omitted at the time of execution.
An oath or declaration in compliance with 37 CFR 1.63, identifying the application by the above Serial Number and Filing Date is required. A SURCHARGE MUST ALSO BE SUBMITTED AS INDICATED BELOW.
4. ☐ The oath or declaration does not identify the application to which it applies. An oath or declaration in compliance with 37 CFR 1.63 identifying the application by the above Serial Number and Filing Date is required. A SURCHARGE MUST ALSO BE SUBMITTED AS INDICATED BELOW.
5. The signature to the oath or declaration is: ☒ missing; ☐ a reproduction; ☐ by a person other than the inventor or a person qualified under 37 CFR 1.42, 1.43, or 1.47. A properly signed oath or declaration in compliance with 37 CFR 1.63, identifying the application by the above Serial Number and Filing Date is required. A SURCHARGE MUST ALSO BE SUBMITTED AS INDICATED BELOW.
6. ☐ The signature of the following joint inventor(s) is missing from the oath or declaration: _____. Applicant(s) should provide, if possible, an oath or declaration signed by the omitted inventor(s), identifying this application by the above Serial Number and Filing Date. A SURCHARGE MUST ALSO BE SUBMITTED AS INDICATED BELOW.
7. ☐ The application was filed in a language other than English. Applicant must file a verified English translation of the application and a fee of \$26.00 under 37 CFR 1.17(k), unless this fee has already been paid. NO SURCHARGE IS REQUIRED FOR THIS ITEM.
8. ☒ Other: **surcharge Due 110.00**

A Serial Number and Filing Date have been assigned to this application. However, to avoid abandonment under 37 CFR 1.53(d), the missing parts and fees identified above in items 1 and 3-6 must be timely provided ALONG WITH THE PAYMENT OF A SURCHARGE OF \$110.00 for large entities or \$55.00 for small entities who have filed a verified statement claiming such status. The surcharge is set forth in 37 CFR 1.16(e). Applicant is given ONE MONTH FROM THE DATE OF THIS LETTER, OR TWO MONTHS FROM THE FILING DATE of this application, WHICHEVER IS LATER, within which to file all missing parts and pay any fees. Extensions of time may be obtained by filing a petition accompanied by the extension fee under the provisions of 37 CFR 1.136(a).

Direct the response to, and any questions about, this notice to the undersigned, Attention, Application Branch, and include the above Serial Number and Filing Date.

M. LASALLE
For Manager, Application Branch
7-3254

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